Growth and characterization of pyrene crystals on carbon nanofibers

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Abstract

Pyrene crystals were grown on carbon nanofibers (CNFs) by dispersing pyrene polycrystals and CNFs in water during ultrasonic irradiation, and they were characterized by scanning electron microscopy, X-ray diffractometry (XRD) and spectroscopy. The XRD measurements indicated that the orientation and size of the pyrene crystals on the CNF aggregates were different from that of the added pyrene polycrystals. Based on the spectroscopic properties of the pyrene crystals on the CNFs, the pyrene crystals on the CNF aggregates and on the individual CNFs were determined to be polycrystals and single crystals, respectively. These results indicate that pyrene crystals are produced on the CNFs by recrystallization of the added pyrene polycrystals and their crystal states depend on the aggregation state of the CNFs.

Keywords: Carbon nanofibers; Pyrene; Single crystals; Absorption; Fluorescence

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1. Introduction

Nanocarbons, such as carbon nanotubes (CNTs) [1,2] and carbon nanofibers (CNFs), [3–5] are promising materials for use in various mechanical, electronic and optical devices because of their remarkable mechanical and unique electronic properties [6-11]. The preparation of the CNFs is technologically easier and economically more favorable for practical applications compared to that of the CNTs. The CNFs bonded to organic molecules are available for optoelectronic devices utilizing the excited-state energy transfer or electron transfer from the tethered molecules to the nanocarbons [12–14]. In addition, noncovalent modifications of the CNFs using aromatic molecules through a π - π interaction have also been utilized [15-19]. Instead of an aromatic molecular modification, the deposition of aromatic crystals on the CNF surfaces enhances two properties of the devices; one is an increase in the absorption efficiency of the aromatic molecules in the crystal state on the CNFs due to higher density of the packed molecules, and the other is a shift to longer wavelengths and an extension of the absorption range due to the formation of the aromatic crystal bands. However, in the case of the crystal modification on the CNFs, the crystal size affects the energy transfer efficiency due to the confined exciton migration range. Therefore, the formation of a smaller crystal on the CNFs is required for a higher energy transfer efficiency.

We have previously shown that pyrene molecules are adsorbed onto the CNFs through a π - π interaction by observing the fluorescence spectra of pyrene on the CNFs [19]. During this procedure, pyrene molecules were adsorbed on the CNFs dispersed by ultrasonic irradiation in a mixed water/ethanol solution of pyrene. To enhance the dispersibility of the CNFs in solution, some functional groups were introduced on the surface of the CNFs by acid treatment. The acid treatment of the CNFs with nitric acid and sulfuric acid leads to oxidized groups such as –COOH and –C=O in the graphene sheet of the materials [18,21–23].

In this study, we tried to prepare the pyrene crystals on the acid-treated CNFs by adding a considerable amount of pyrene that exceeded the solubility and the acid-treated CNFs in water and ultrasonically agitating. The pyrene molecules can be easily adsorbed on the CNFs dispersed by ultrasonic irradiation in water due to their hydrophobic characteristics. Furthermore, the ultrasonication can create a supersaturated solution of pyrene near the CNFs by rapid local cooling after an increase in the local temperature created by bubble collapse and concomitant release of shock waves [24]. Therefore, it is expected that the heterogeneous nucleation and the crystallization of pyrene on the CNFs are attained by agitating the starting material of the pyrene and the acid-treated CNFs in only water during ultrasonic irradiation. Characterization of the pyrene on the CNFs was carried out by scanning electron microscopy (SEM), X-ray diffractometry (XRD) and spectroscopy.

2. Experimental

The pyrene (Wako Pure Chemicals, S grade) was recrystallized three times from ethanol. It is referred to as the original pyrene polycrystals in this report. The CNFs (VGCF-G) having a diameter of ca. 200 nm, a length of ca. 10 - 20 μ m, and a surface area of ca. 15 m² g⁻¹ were provided by Showa Denko Co., Ltd. The cutting and functionalization of the CNFs was carried out using a mixture of concentrated nitric acid and sulfuric acid, as previously reported [18–21]. The acid-treated CNF sample was designated AA-CNF in this report.

The AA-CNF (0.50 mg) and the original pyrene crystals (10 mg) were added to water (10 ml) and ultrasonically agitated. The crystallization of pyrene on the CNFs is expected to be attained by this procedure. Therefore, the resultant suspension is expected to contain individual CNFs loading pyrene crystals, CNF aggregates loading pyrene crystals, pyrene molecules and added pyrene crystals.

The deposition of the pyrene crystals on the CNF aggregates and on the individual CNFs was confirmed by SEM (Hitachi S-4100) and XRD (Rigaku Rint 2200V). The preparation procedures of the samples for SEM observation and XRD measurement are shown in Scheme 1. The two different states of the CNF aggregates were separated from the suspension by filtration. The aqueous suspension containing the added pyrene polycrystals and AA-CNF ultrasonically agitated for 15 days was filtered through a coarse

filter paper having 5.0 μ m pores to separate the larger aggregates of the added pyrene polycrystals and AA-CNF. The separated substance was designated Sample A1. The resultant filtrate was then filtered through a finespun filter paper having 0.2 μ m pores to separate the smaller CNF aggregates. The separated substance was designated Sample A2.

(Scheme 1)

The following two reference samples were prepared in order to investigate whether or not the crystallization of the pyrene on the CNF aggregates occurred by ultrasonic irradiation. The AA-CNF (0.50 mg) and the original pyrene crystals (10 mg) were then added to water (10 ml). The solutions were left or stirred at 313 K for 15 days without ultrasonication. This temperature was the same as in the ultrasonication bath. Since almost all of the AA-CNF and the pyrene crystals in the system were aggregated, they were separated by a coarse filter paper having 5.0 µm pores. The separated substances from the suspensions not stirred or stirred were designated Sample CA1 and Sample CA2, respectively.

The individual CNFs were separated from the suspension as follows. The supernatant was obtained by centrifugation of the suspension at 4,000 g. Since the centrifugation cannot spin down the individual CNFs, the individual CNFs loading pyrene crystals and pyrene molecules are expected to be contained in the supernatant. The supernatant was then dried and this specimen was designated Sample B.

The following reference sample was prepared in order to investigate whether the crystallization of pyrene on the individual CNFs occurred during the solvent evaporation. The AA-CNF (0.50 mg) was dispersed in water (10 ml) by ultrasonic irradiation. They were centrifuged in order to leave only the individual CNFs in the supernatant. The, excess pyrene was then added to this supernatant. After saturation of the pyrene at 313 K was attained, the suspension was filtered though a finespun filter paper having 0.2 µm pores in order to remove the undissolved pyrene crystal. The resultant filtrate was dried and this specimen was designated Sample CB.

The UV absorption (Shimadzu UV-3150 spectrophotometer), fluorescence and fluorescence excitation spectra (Shimadzu RF-5300 spectrofluorophotometer) of the pyrene on the CNFs were measured. The aim of these analyses was to investigate the pyrene crystal states on the CNF aggregates or on the individual CNFs. The preparation procedures of the samples for the spectroscopy are shown in Scheme 2. For the pyrene crystals on the individual CNFs, the spectral measurements were done for the supernatant obtained by centrifuging the suspensions containing the added pyrene polycrystals and AA-CNF ultrasonically agitated for 15 days. This supernatant was designated Supernatant A.

(Scheme 2)

Furthermore, the absorption spectra of the following three reference supernatants were The first sample was prepared as follows in order to investigate whether or not measured. the heterogeneous nucleation of the pyrene on the AA-CNF occurred. The supernatant obtained by centrifuging the suspension of the pyrene polycrystals without the AA-CNF ultrasonically agitated for 15 days. This supernatant was designated Supernatant CA1. The second sample was prepared as follows in order to investigate whether or not the crystallization occurred by stirring. The supernatant was obtained by centrifuging the suspensions containing the added pyrene polycrystals and AA-CNF stirred for 15 days at 313 K. This supernatant was designated Supernatant CA2. The third sample was prepared as follows in order to investigate whether or not the crystallization in the AA-CNF dispersed water occurred by stirring. The AA-CNF (1.0 mg) was dispersed in water (20 ml) by ultrasonic irradiation, and then it was centrifuged to leave only the individual CNFs in the Pyrene (10 mg) was added to this supernatant (10 ml). supernatant. The solution was stirred for 15 days at 313 K. Finally, the supernatant was obtained by centrifugation of the stirred solution. This supernatant was designated Supernatant CA3.

3. Results and discussion

3.1. Confirmation of deposition of pyrene crystals on carbon nanofibers

Figure 1 shows the SEM images of Samples CA1, CA2, A1, A2, CB and B. In the image of Sample CA1 shown in Fig.1 (a), the AA-CNF does not seem to be covered by the The diameter of the AA-CNF in Sample CA1 is not different from that of pyrene crystals. the added AA-CNF. On the other hand, the AA-CNF in Sample CA2 shown in Fig.1 (b) has a larger diameter compared to that of Sample CA1. Thus the pyrene crystals are In the images of Samples A1 and A2 shown in Fig. 1 (c) and (d), attached to the AA-CNF. respectively, the tubular shape of the CNFs is not clearly observed because a large amount of the pyrene crystals surround the AA-CNF. The AA-CNF in Sample A1 seems to be surrounded by more pyrene crystals than that in Sample A2. These results suggest that the pyrene crystals exist on the AA-CNF aggregates by stirring or ultrasonication.

Figures 1 (e) and (f) show Samples CB and B, respectively. These samples seem to be the individual CNFs on the based on the high magnification SEM images. The images of Samples CB and B displayed microasperities on the AA-CNF. These asperities are predicted to be pyrene nanocrystals on the AA-CNF. From the SEM images, it was not clear that the crystallization of pyrene occurred or not by ultrasonication because the AA-CNF in both samples have microasperities. Therefore, we will try to confirm the crystallization by spectroscopic measurements in § 3.2.

(Figure 1)

Figure 2 shows the XRD patterns of the AA-CNF and Samples CA1, CA2, A1 and A2. Although we have observed the XRD patterns of Samples CB and B, we could not obtain a significant signal due to low density of the CNFs in these samples. In the XRD patterns for Samples CA1, CA2, A1 and A2, the peak positions of the pyrene crystals were identical. Since each characteristic interplane spacing of the pyrene crystal was unchanged in Samples CA1, CA2, A1 and A2, they have the same crystal structure.

The pyrene crystal belongs to the sandwich-herringbone class; two parallel molecules are arranged in a sandwich motif through π - π interactions, and each motif is arranged in a herringbone fashion favoring C - $H \cdots \pi$ interactions[25]. The relative intensity of the peaks for the pyrene crystals in Samples CA1, CA2, A1 and A2 differ from each other. We focused our discussion on the $(0\ 0\ 1)$ plane of the pyrene crystal. The relative intensities of the (0 0 1) plane of the pyrene crystal for Samples CA2, A1 and A2 are lower than that for The order of the relative intensity of the (0 0 1) plane for Samples CA2, A1 Sample CA1. and A2 was observed as Sample A2 < Sample CA2 < Sample A1. The relative intensities of the peak at 26.6° assigned to the AA-CNF for Samples CA2, A1and A2 show the order of Sample A1 < Sample CA2 < Sample A2. Therefore, the relative intensities of the $(0\ 0\ 1)$ plane decreased with an increase in the relevant amount of the AA-CNF. This means that a relatively small amount of pyrene molecules was aligned in the c-axis direction on the AA-CNF.

The full width at half maximum (FWHM) of the diffraction peak for the pyrene crystals in Samples CA2 and A1 is almost identical to that of Sample CA1. On the other hand, that of Sample A2 differs from that of Sample CA1. Based on the Scherrer formula, the crystallite sizes of the pyrene in the sample were estimated. For estimation using 2θ $=10.8^{\circ}$, which corresponds to the (0 0 1) plane, the crystallite sizes of the pyrene in the direction in Samples CA1, CA2, A1 cannot be evaluated due to their small FWHM. It is inferred that these are more than 200 nm. On the other hand, that in Sample A2 was For estimation using $2 \theta = 23.6^{\circ}$, which corresponds to the (2.2) evaluated to be 100 nm. 0) plane, the crystallite sizes of the pyrene in the direction in Samples CA1, CA2, A1 and A2 were evaluated to be 80 nm, 80 nm, 80 nm and 60 nm, respectively. The crystallite sizes of the pyrene in Sample A2 were smaller than that of Sample CA1. Also, the difference between the crystallite sizes of the $(0\ 0\ 1)$ and $(2\ 2\ 0)$ directions of pyrene in Sample A2 was smaller than that for Sample CA1. This means that a relative amount of pyrene molecules aligned in the c-axis in the pyrene crystallite in Sample A2 was smaller compared to that of Sample CA1. These results indicate smaller and relatively isotopic pyrene crystals were grown on the AA-CNF in Sample A2.

(Figure 2)

3.2. Identification of pyrene crystal states on carbon nanofibers by spectroscopic measurements

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The absorption spectra of the pyrene crystals on the AA-CNF were measured in order to understand the pyrene crystal states on the AA-CNF. Figure 3 shows the UV absorption spectra of the pyrene on the CNF aggregates and on the individual CNFs. Fig. 3(a) shows the absorption spectra of the pyrene molecules in water, the original pyrene polycrystals, and Sample A2. The absorption spectrum of the original pyrene polycrystals exhibits bands with peaks at 322, 338, and ca. 376 nm in the 300 - 400 nm region. The former two peaks are assigned to the transitions from the ground state to the ${}^{1}L_{a}$ exciton states. These peaks shifted to the lower energy side compared to that of the pyrene molecules in water. This shift is attributed to the interaction on the dimeric structure in the pyrene crystal [26]. The peak at ca. 376 nm is attributed to the transition from the ground state to the bottom of the free exciton state, the ${}^{1}L_{b}$ exciton state, in the pyrene polycrystal [27–29]. The absorption spectra of Sample A2 almost corresponds to that of the original pyrene polycrystals, whereas the XRD patterns of these samples are different from that of the added pyrene polycrystals. This indicates that the orientation and size of the resultant crystals on the CNF aggregates is different from that of the added pyrene polycrystals, and the crystal state remained as polycrystals.

Figure 3(b) shows the absorption spectra of Supernatants A, CA1, CA2, and CA3. The absorption spectrum of Supernatant A exhibits three bands with peaks at 320, 336, and 353 nm. This absorption profile revealed a significantly different shape from that of the original pyrene polycrystals. The absorption spectrum of Supernatant A is similar to that of the pyrene single crystals exhibiting three sharp peaks at ca. 320, 338, and 355 nm as reported by Takahashi et al. [30]. The spectrum of the pyrene single crystals is shifted to the short wavelength side compared to that of the original pyrene polycrystals. This shift is attributed to the formation of a broader band in the polycrystals compared to the single crystals due to perturbation between the crystals grain boundaries.

On the other hand, the spectrum of Supernatant CA1 indicated the same spectrum as that of the pyrene molecules in water instead of the pyrene crystal band. The crystallization of the pyrene single crystals did not occur in the absence of the AA-CNF. The spectrum of Supernatant CA2 is similar to that of Supernatant CA1. Since the absorption of the AA-CNF did not appear in the spectrum of Supernatant CA2, the AA-CNF was not dispersed in water by stirring. Consequently, the single pyrene crystals were not The spectrum of Supernatant CA3 indicated the peaks of pyrene molecules in produced. water on the background due to absorption of the AA-CNF. In spite of the presence of the AA-CNF, the crystallization of pyrene did not occur by stirring. These results mean that the heterogeneous nucleation and crystal growth of the pyrene single crystals on the AA-CNF dispersed by ultrasonication are attained from the pyrene supersaturated solution created by ultrasonication.

(Figure 3)

In order to investigate in more detail the state of the pyrene on the individual AA-CNF in Supernatant A, the fluorescence and fluorescence excitation spectra were measured. Figure 4 shows the fluorescence spectra of the original pyrene polycrystals and Supernatant A. All of the fluorescence spectra were corrected by subtracting the scattering of incident light. The fluorescence spectra of the original pyrene polycrystals and Supernatant A excited at 360 nm show similar spectral features and exhibit only one peak at 460 nm. The peak at 460 nm corresponds to the fluorescence from the excimer-like state induced by local self-trapping of the excitation energy and is related to a specific dimeric interaction in the crystal lattice [27-29,31]. Although the free exciton fluorescence is generally observed at 376 nm in the pyrene polycrystals, it was apparently not observed here. This suggested that the free exciton fluorescence was concealed behind the intense scattering of light. The fluorescence spectrum of Supernatant A excited at 335 nm exhibits a dual fluorescence. One is the vibronic bands seen at 372-392 nm and is assigned to the pyrene monomer in water The other is the broad band, which has a peak at 460 nm, due to an excimer-like [26,32,33]. fluorescence, as observed in the spectrum at the 360 nm excitation. These results indicate that the absorption band at ca. 360 nm is attributed to only the pyrene crystals, and that at ca. 335 nm is attributed to not only the pyrene crystals, but also the pyrene molecules in water. Furthermore, according to the report by Takahashi et al., the fluorescence of the pyrene single crystals indicates the same excimer-like fluorescence as that of the original pyrene

polycrystals [34]. Therefore, the fluorescence having a peak at 460 nm in Supernatant A will be attributed to the pyrene single crystals. We will try to confirm this by measurement of fluorescence excitation spectrum.

(Figure 4)

Figure 5 shows the fluorescence excitation spectra of the pyrene molecules and crystals under various conditions. The fluorescence excitation spectra of Supernatant A depend on the observed wavelength. The spectrum of Supernatant A monitored at 390 nm exhibits peaks at 307, 320 and 335 nm. These peaks correspond to the transition to the ${}^{1}L_{a}$ molecular state of the pyrene in solution [26]. On the other hand, the spectrum of Supernatant A monitored at 480 nm exhibits peaks at 340, 360 and 376 nm. There are two obvious differences between the spectra of Supernatant A monitored at 480 nm and the original pyrene polycrystals having peaks at 343, 367 and 376 nm. One is the blue shift of the 360 nm peak of the pyrene crystals in Supernatant A compared to the peak at 367 nm of the original pyrene polycrystals. As already mentioned about the absorption spectra, this blue shift is attributed to the pyrene single crystals growing on the CNF surface. However, there is a red shift in the peak around 360 nm in the excitation spectrum compared to that in the UV absorption spectrum. The absorption peak shift to the longer wavelength side accompanied by an increase in the crystal size in the perylene crystal has been reported [35]. Therefore, the red shift suggests that the fluorescence efficiently originates from the larger crystals having an absorption peak on the longer wavelength side. The other fact is the relative intensity of the 376 nm peak of the pyrene crystals in Supernatant A which is lower than that of the original pyrene polycrystals. These results suggest that the fluorescence at 460 nm in Supernatant A was mostly generated from the pyrene single crystals and a small amount from the pyrene polycrystals. The free exciton band in the pyrene polycrystals at 376 nm that appeared in the fluorescence excitation spectrum of Supernatant A did not distinctly appear in the absorption spectrum. This suggests that the fluorescence efficiency of the pyrene single crystals on the AA-CNF is lower than that of the pyrene polycrystals on the AA-CNF. Therefore, this result implies that the energy transfer originating from the pyrene single crystals to the CNFs is more efficient than that from the pyrene polycrystals due to enhancement of the interaction between the smaller pyrene single crystal and the AA-CNF.

(Figure 5)

4. Conclusions

We confirmed the growth of pyrene crystals on the CNFs by dispersing the original pyrene polycrystals and the CNFs in water during ultrasonication. Based on the XRD analysis, it was determined that the orientation and size of the pyrene crystals on the CNFs is different from that of the added pyrene polycrystals. This suggests that the crystallization of the pyrene on the CNF aggregates occurred. From the spectroscopic measurements, it was determined that the pyrene crystals grew as single crystals on the individual CNFs and as polycrystals on the CNF aggregates. On the surface of the individual CNFs dispersed by ultrasonication, the nucleation occurs from the pyrene supersaturated solution created by ultrasonication. Consequently, the pyrene single crystals are formed on the AA-CNF. The growth mechanism of pyrene crystals on the CNF aggregates was suggested as follows. The single pyrene crystals grew during the initial stage. They then gradually became larger polycrystals due to the closer spacing of the CNFs to each other. Therefore, the single crystals on the individual CNFs have a smaller crystal size compared to the polycrystals on the CNF aggregates.

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References

- [1] S. Iijima, Nature 354 (1991) 56.
- [2] S. Iijima, T. Ichihashi, Nature 363 (1993) 603.

- [3] A. Oberlin, M. Endo, T. Koyama, J. Cryst. Growth. 32 (1976) 335.
- [4] M. Endo, Chemtech. 18 (1988) 568.

[5] M. Endo, Y. A. Kim, T. Hayashi, K. Nishimura, T. Matusita, K. Miyashita, M. S. Dresselhaus, Carbon 39 (2001) 1287.

- [6] R. H. Baughman, A. A. Zakhidov, W. A. de Heer, Science 297 (2002) 787.
- [7] R. Singh, D. Pantarotto, D. McCarthy, O. Chaloin, J. Hoebeke, C. D. Partidos, J-P.

Briand, M. Prato, A. Bianco, K. Kostarelos, J. Am. Chem. Soc. 127 (2005) 4388.

- [8] J. S. Tans, R. M. A. Verschueren, C. Dekker, Nature 393 (1998) 49.
- [9] R. Saito, G. Dresselhaus, M.S. Dresselhaus, Phys. Rev. B 61 (2000) 2981.
- [10] M. J. O'Connell, S. M. Bachilo, C. B. Huffman, V. C. Moore, M. S. Strano, E. H.

Haroz, K. L. Rialon, P. J. Boul, W. H. Noon, C. Kittrell, J. Ma, R. H. Hauge, R. B. Weisman,R. E. Smalley, Science 297 (2002) 593.

[11] Y. Maniwa, K. Matsuda, H. Kyakuno, S. Ogasawara, T. Hibi, H. Kadowaki, S.Suzuki, Y. Achiba, H. Kataura, Nature materials 6 (2007) 135.

[12] L. Qu, B. R. Martin, W. Huang, K. Fu, D. Zweifel, Y. Lin, J. Chem. Phys. 117 (2002) 8089. [13] H. Li, E. M. Kose, L. Qu, Y. Lin, B. R. Martin, B. Zhou, A. B. Harruff, F. L. Allard,Y. Sun, J. Photochem. Photobio. A 185 (2007) 94.

[14] M. Álvaro, P. Atienzar, L. J. Bourdelande, H. García, Chem. Phys. Lett. 384 (2004)119.

[15] J. R. Chen, Y. Zhang, D. Wang, H. Dai, J. Am.Chem. Soc. 123 (2001) 3838.

[16] H. Nishikiori, N. Tanaka, S. Kubota, M. Endo, T. Fujii, Chem. Phys. Lett. 390 (2004) 389.

[17] S. Kubota, H. Nishikiori, N. Tanaka, M. Endo, T. Fujii, Chem. Phys. Lett. 412 (2005) 223.

[18] S. Kubota, H. Nishikiori, N. Tanaka, M. Endo, T. Fujii, J. Phys. Chem. B 109 (2005)23170.

[19] T. Tanigaki, H. Nishikiori, S. Kubota, N. Tanaka, M. Endo, T. Fujii, Chem. Phys.Lett. 448 (2007) 218.

[20] Nishikiori, H.; Tanaka, N.; Tanigaki, T.; Endo, M.; Fujii, T. J. Photochem. Photobio.A 193 (2008) 161.

[21] J.; Liu, A. G. Rinzler, H. Dai, J. H. Hafner, R. K. Bradley, P. J. Boul, A. Lu, T. Iverson, K. Shelimov, C. B. Huffman, F. R. Macias, Y. S. Shon, T. R. Lee, D. T. Colbert, R. E. Smallery, Science 280 (1998) 1253.

[22] P. V. Lakshminarayanan, H. Toghiani, C. U. Pittmann Jr., Carbon 42 (2004) 2433.

[23] M. A. Hamon, H. Hui, P. Bhowmik, H. M. E. Itkis, R. C. Haddon, Appl. Phys. A 74 (2002) 333.

[24] M. D. Luque de Castro, F. Priego-Capote, Ultra. Sonochem. 14 (2007) 717.

[25] F. P. A. Fabbiani, D. R. Allam, S. Parsons, C. R. Pulham, Acta Cryst. B62 (2006)826.

[26] T. Fujii, E. Shimizu, S. Suzuki, J.Chem. Soc. Faraday Trans. 1 84 (1988) 4387.

[27] A. Matsui, H. Nishimura, J. Phys. Soc. Jpn. 49 (1980) 657.

[28] T. Nakanishi, K. Mizuno, A. Matsui, H. Nishimura, J. Luminescence 47 (1991) 303.

[29] A. Matsui, T. Nakamura, S. Nakatani, T. Ohno, K. Mizuno, Syntheic Metals 64 (1994) 177.

[30] N. Takahashi, B Gombojav, T. Yoshinari, S. Nagasaka, Y. Takahashi, A. Yamamoto,T. Goto, A. Kasuya, J. Sol. Sta. Chem. 177 (2004) 3479.

[31] R. Seyfang, H. Port, P. Fischer, H. C. Wolf, J. luminescence 51 (1992) 197.

[32] T. Fujii, K. Murayama, N. Negishi, M. Anpo, E. J. Winder, D. R. Neu, A. B. Ellis,Bull. Chem. Soc. Jpn. 66 (1993) 739.

[33] A. Nakajima, Bull. Chem. Soc. Jpn. 44 (1971) 3272.

[34] N. Takahashi, B Gombojav, S.Yoshinari, Y. Takahashi, T. Nagasaka, A. Yamamoto,T. Goto, A. Kasuya, J. Phys. Soc. Jpn. 76 (2007) 034703.

[35] H. Kasai, H. Oikawa, S. Okada, H. Nakanishi, Bull. Chem. Soc. Jpn. 71 (1998) 2597.

Figure captions

Scheme 1

The preparation procedures of (a) ultrasonically agitated samples and (b) control samples for

SEM observation and XRD measurement

Scheme 2

The preparation procedures of (a) ultrasonically agitated samples and (b) control samples for

spectroscopy

Figure 1

SEM images of (a) Sample CA1, (b) Sample CA2, (c) Sample A1, (d) Sample A2, (e) Sample

CB, and (f) Sample B.

Figure 2

XRD patterns of (1) the AA-CNF, (2) Sample CA1, (3) Sample CA2, (4) Sample A1, and (5)

Sample A2.

Figure 3

(a) UV absorption spectra of (1) pyrene molecules in water (saturated solution), (2) the original pyrene polycrystals and (3) Sample A2.

(b) UV absorption spectra of (1) Supernatant A, (2) Supernatant CA1, (3) Supernatant CA2, and (4) Supernatant CA3

Figure 4

Fluorescence spectra of (1) the original pyrene polycrystals and (2, 3) Supernatant A. (1) and (2) were excited at 360 nm. (3) was excited at 335 nm.

Figure 5

Fluorescence excitation spectra of (1) pyrene molecules in water (saturated solution) (2) the original pyrene polycrystals and (3, 4) Supernatant A. (1) and (3) were monitored at 390 nm. (2) and (4) were monitored at 480 nm.



(a)

The AA-CNF (0.50 mg) and the original pyrene crystals (10 mg) were added to water (10 ml) and ultrasonically agitated for 15 days

Centrifugation at 4,000 g

Supernatant A

(b)

Comparison with Supernatant A



(a)



(c)









(d)







Figure 1











Figure 5

