

Obligate Anaerobic *Lactobacillus Casei* KJ686 Selectively Targets Solid Tumors and Exhibits an Antitumor Effect

Michihiko HARADA^{1)2)*} and Jun AMANO²⁾

1) *Department of Molecular Oncology, Institute of Pathogenesis and Disease Prevention,
Shinshu University Graduate School of Medicine*

2) *Department of Surgery, Shinshu University School of Medicine*

Hypoxia in solid tumors is known to confer treatment resistance and lead to rapid progression and poor prognosis. As a novel cancer drug-delivery candidate, *Lactobacillus casei* KJ686 (*L. casei* KJ686) is a mutant strain of the facultatively anaerobic *L. casei* ATCC393 that was transformed into an obligate anaerobe to suit the hypoxic conditions within solid tumors. In the present study, we intravenously administered *L. casei* KJ686 to treat tumor-bearing mice and investigated the in vitro cytotoxicity of *L. casei* KJ686 and its culture medium containing bacterial excretion products. We discovered that *L. casei* KJ686 localized exclusively in tumor tissues and suppressed tumor growth in our animal experiments. Furthermore, the culture supernatant of *L. casei* KJ686 exhibited a strong cytotoxic effect in vitro. The specific cytotoxic substances presented in its culture supernatant may have a pivotal role in direct anti-tumor effect in solid tumors. *Shinshu Med J* 63 : 375—384, 2015

(Received for publication May 19, 2015 ; accepted in revised form August 17, 2015)

Key words : obligate anaerobe, hypoxia, *Lactobacillus casei*, cytotoxicity, culturing-supernatant

Abbreviations : LLC, Lewis lung carcinoma

I Introduction

Cytotoxic chemotherapy is useful for systemic antitumor treatment, but the large doses of drugs that are often needed for effective results may also induce many adverse side effects, such as nausea, vomiting, hair loss, anemia, agranulocytosis, stomatitis, diarrhea, and dehydration. Although many molecular targeted agents have been developed up to the present, they also have many adverse side effects such as hand-foot syndrome and interstitial pneumonitis, because target molecules exist in both tumor and normal tissues.

Accordingly, more specific antitumor effects and/

* Corresponding author : Michihiko Harada

Department of Molecular Oncology, Institute of Pathogenesis and Disease Prevention, Shinshu University Graduate School of Medicine, 3-1-1 Asahi, Matsumoto, Nagano 390-8621, Japan

E-mail : mrharada2000@yahoo.co.jp

or more selective delivery systems of antitumor drugs to tumor tissues are needed^{1)–10)}.

Living bacteria have recently been considered as a drug delivery tool in cancer gene therapy ; one such example is lactic acid bacteria (LAB). LAB is a nonpathogenic anaerobic bacteria widely used in food and intestinal health supplements¹¹⁾¹²⁾. Numerous reports have documented the safety and functional characteristics of LAB.

Hypoxic conditions are characteristic of solid tumors in both rodent¹³⁾ and many types of human tumors¹⁴⁾. The oxygen pressure in human tumors has been reported to be 0–20 mmHg, whereas that in normal tissues shows a range of 20–100 mmHg⁴⁾¹⁵⁾¹⁶⁾. Tumor hypoxia, mostly stemming from poor perfusion of fresh blood, causes the development of aggressive cell clones and leads to rapid progression and poor prognosis^{17)–19)}.

Furthermore, hypoxia is known to directly and

indirectly confer resistance to radiation and some chemotherapies, which often results in treatment failure^{20,21}).

Recently, *Bifidobacterium longum* (*B.longum*), a strain of obligated LAB, has been reported to selectively germinate and grow in the hypoxic regions of solid tumors following intravenous injection and was demonstrated to be a safe and effective drug carrier to solid tumors^{1)–4)}.

Lactobacillus casei (*L.casei*), another strain of LAB, is a gram-positive facultatively anaerobic rod-shaped bacterium. *L.casei* is believed to have indirect antitumor effects due to its ability to modulate host immune responses^{22)–24)}, but no direct antitumor properties have been shown to date²⁵⁾. However, numerous recent reports have noted that the cytoplasmic fractions, proteins, and peptidoglycans of *L.casei* may suppress the proliferation of tumor cell lines^{26)–30)}.

We directed our attention to the fact that *L.casei* has thin polysaccharides in its wall and may easily excrete proteins synthesized via an expression vector, since *B.longum* with cytosine deaminase has successfully exhibited antitumor effects through intravenous injection in combination with 5-FC^{2)–4)}.

We examined whether obligated *L.casei* could also be a specific delivery tool to target tumor tissues, and produced *L.casei* KJ686, a novel *L.casei* strain that was transformed to be obligately anaerobic, to germinate and grow in the hypoxic regions of solid tumors after intravenous injection.

In the present study, we found that *L.casei* KJ686 was a tolerated and selective carrier to solid tumors and exhibited suppressive effects on tumor growth. We also assessed the direct cytotoxic effect of *L.casei* KJ686 and/or the substances excreted from this bacterium *in vitro*.

II Materials and Methods

A *In vivo*

1 Growth of *L.casei* KJ686

L.casei KJ686 with the erythromycin resistance gene was developed as a mutant strain of the facultatively anaerobic *L.casei* ATCC393 that had

been transformed to be obligately anaerobic. The mutant was maintained in MRS broth (OXOID, Hampshire, England) with erythromycin (0.2 µg/ml) at 37 °C without O₂. *L.casei* KJ686 in the mid-logarithmic growth phase was used for all experiments.

2 Preparation of tumor-bearing mice

Lewis lung carcinoma (LLC) cells were cultured in medium containing DMEM (Sigma, Louis, MO, USA), 10 % fetal bovine serum and streptomycin (10 µg/ml) at 37 °C with 5 % CO₂. Male 8-week-old C57BL/6 mice (Japan SLC, Hamamatsu, Japan) of similar size and weight were used and maintained with a standard rodent diet (Oriental Yeast, Tokyo, Japan) at the Shinshu University animal center (Matsumoto, Japan). The animal experiments in this study were carried out in accordance with the Guidelines for Animal Experimentation of Shinshu University School of Medicine. Aliquots of 10⁶ tumor cells in 100 µl Hanks' Solution (Nissui Pharmaceutical Co., Ltd., Tokyo, Japan) were inoculated under the femoral skin of the mice.

The solid tumors were palpable 5–8 days after inoculation. For each tumor, the maximum diameter (A) and diameter at a right angle to A (B) were measured daily using a sliding caliper. Tumor volume was estimated as $1/2 \times A \times B^2$. All mice survived the testing period.

3 Intravenous injection of *L.casei* KJ686 into tumor-bearing mice

L.casei KJ686 was harvested by centrifugation (3000 rpm, 1 min at 4 °C), washed twice with PBS (–), and then resuspended in PBS (–) at 5×10^8 c.f.u./100 µl. The solid tumor-bearing mice were randomly separated into 2 groups and used for subsequent studies. When tumors reached approximately 65.4 mm³ (5 mm in diameter), *L.casei* KJ686 [5×10^8 c.f.u./100 µl PBS (–)] was injected into a tail vein of one group once a day for 2 days. The other mice underwent no treatment as the control group. The tumor growth ratio was calculated every day as follows:

(tumor volume) / (tumor volume on the first day of intravenous injection).

4 Localization of *L.casei* KJ686

All tumor-bearing mice were sacrificed 17 days after the first injection of *L.casei* KJ686. Normal tissue samples from the lung, liver, and tumor were excised and placed into a homogenizer with 9-fold anaerobic diluent. The samples were plated on an MRS agar medium at 37 °C without O₂ to assess the bacterial numbers in each tissue. The localization of *L.casei* KJ686 in tumor tissue sections was visualized microscopically with Gram's and Hematoxylin-Eosin staining.

B In vitro

1 Preparation of *L.casei* KJ686 and culture supernatant of *L.casei* KJ686 to examine for cytotoxic effect

LLC cells were maintained at 37 °C with 5 % CO₂ and routinely cultured in DMEM supplemented with 10 % fetal bovine serum with or without streptomycin (10 µg/ml). *L.casei* KJ686 was harvested by centrifugation (3000 rpm, 1 min at 4 °C) and washed twice with PBS (-). The bacteria were resuspended in DMEM without streptomycin at 1×10¹⁰ c.f.u./ml (living *L.casei* KJ686). Heat-killed *L.casei* KJ686 was prepared by boiling living *L.casei* KJ686 at 95 °C for 90 min. After 24 hr of anaerobic culture in DMEM without antibiotics, the culture supernatant of *L.casei* KJ686 was harvested by centrifugation (12000 rpm, 10 min at 4 °C). The supernatant was divided into 6 fractions by molecular weight (over 100 kDa, under 100–50 kDa, under 50–10 kDa, under 10–3 kDa, under 3–1 kDa and under 1 kDa) with Amicon Ultra-0.5 ml Centrifuged Filter devices (100 k, 50 k, 10 k, and 3 k; Merck Millipore, Darmstadt, Germany), and a Microsep Centrifugal device (1 k; Pall Life Sciences, Michigan, USA). Each fraction was diluted to its original concentration with DMEM. Furthermore, the under 1 kDa supernatant (1 ml) was vortexed with ethyl acetate (1 ml) and separated into aqueous and organic layers by centrifugation. To remove the ethyl acetate, the organic layer was desiccated in draft for 48 hr. After desiccation, the organic layer powder was vortexed with 990 µl DMEM and 10 µl DMSO. The aqueous layer was threaded into a hydrophobic resin (non-methyl alcohol fraction), and then sev-

eral concentrations (20–100 %) of methyl alcohol were threaded into the hydrophobic resin and recovered (20–100 % methyl alcohol fraction). Each aqueous supernatant with alcohol was lyophilized in a desiccator and diluted with sterilized water.

2 Gram's stain

For both living and heat-killed *L.casei* KJ686, the neo B & M (Wako Pure Chemical Industries, Ltd., Osaka, Japan) method was performed according to the manufacturer's instructions.

3 Cell viability assay

The Premix WST-1 Cell Proliferation Assay System (Takara Bio Inc., Shiga, Japan) is based on the cleavage of the tetrazolium salt WST-1 by cells to form a red formazan dye. Cell viability was measured with this assay according to the manufacturer's instructions. LLC cells (1×10⁴ in 90 µl/well) on a 96-well microplate were incubated for 24 hr at 37 °C with 5 % CO₂ with various concentrations of *L.casei* KJ686 culture supernatants (1×10⁶–1×10⁸ in 10 µl/well of living or heat-killed *L.casei* KJ686). WST-1 reagent (1:10) was then added. After 60 min of incubation at 37 °C with 5 % CO₂, the absorbance of the colored formazan was determined using a Vmax automated microplate reader (Wako, Osaka, Japan) at 450 nm and 650 nm wavelengths. The mean absorbance of control wells (cells without supernatant) represented 100 % cell viability.

4 Colony-forming assay

LLC cells (500/well) on a 6-well plate were incubated for 24 hr at 37 °C with 5 % CO₂. Various conditions of *L.casei* KJ686 culture supernatants were then added to the LLC cells, which were then incubated further. After 8 days, the supernatants were removed from the wells and the cells were fixed and stained with 10 % formalin (Wako, Osaka, Japan) and crystal violet solution (Wako, Osaka, Japan) according to the manufacturer's instructions. The toxic activity of supernatants was estimated by the viability of LLC cells.

C Statistical analysis

Experimental data are expressed as the mean ± SE of the mean. Statistical analysis was performed using the Mann-Whitney *U*-test. A *p*<0.05 was

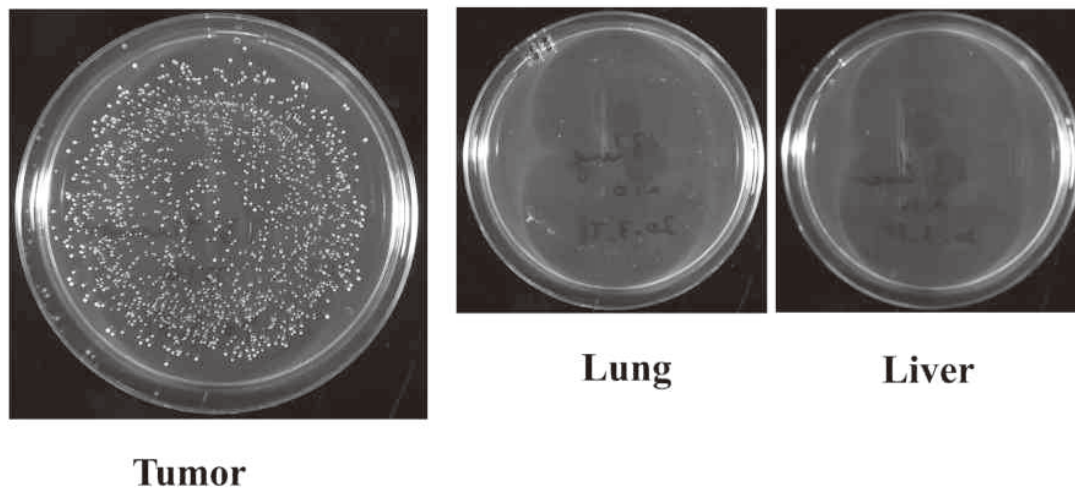


Fig. 1 Germination of *L. casei* KJ686. Seventeen days after intravenous inoculation of *L. casei* KJ686, organs and tumors were homogenized with a 9-fold anaerobic diluent. Samples of 100 μ l of homogenized tissues were cultured on an MRS agar medium at 37 °C without O₂ for 3 days.

considered to be statistically significant.

III Results

A *In vivo*

1 Selective localization of *L. casei* KJ686 in tumors

We examined the germination of *L. casei* KJ686 in the lungs, livers and tumors of KJ686-injected mice.

Another obligate anaerobic bacterium, *B. longum* was not at a detectable level in any normal tissues after seven days¹⁾. So we sacrificed KJ686-injected mice nine days after intravenous injection, *L. casei* KJ686 was selectively detected in tumors, with no bacterial growth in either the lungs or livers (Fig. 1). Each tumor had necrotic areas in its center (Fig. 2). Gram's staining of tumor tissues showed that *L. casei* KJ686 was focally scattered in the border areas between necrotic and viable regions (Fig. 2). Neutrophils existed in necrotic and viable regions. The number of *L. casei* KJ686 in solid tumors estimated by tissue homogenation appeared to reflect the bacterial number in border areas.

2 Inhibition of tumor growth with *L. casei* KJ686

Fig. 3 shows the tumor growth ratio after intravenous injection of *L. casei* KJ686 (n=25) and in controls (n=36). After administration of *L. casei* KJ686, neither suppression of activity nor death were observed in the test group. From day 5 after the first injection, the tumor growth ratio of the

KJ686 group was significantly less than that of the control group. We next estimated RNA expression of IL-2, IL-4, IL-12p, IL-1 β , and IFN- γ by real time RT-PCR of the border areas of tissue samples, but no significant differences were noted (data not shown). Cytokine levels in the serum of KJ686-injected mice were also estimated on days 3 and 9, but no significant findings were seen except for IL-1 β on day 3 (data not shown). Although IL-1 β was transiently increased, enhancement of host immunological defenses by *L. casei* KJ686, such as an increase in the number of NK cells in tumors, was not observed (data not shown).

B *In vitro*

1 Cytotoxic effect of *L. casei* KJ686 on LLC cells

The direct cytotoxic effect of *L. casei* KJ686 on LLC cells was assessed by the WST-1 method. The morphology of *L. casei* KJ686 was not visibly altered by heat treatment (Fig. 4), which indicated that the bacterial body was not ruptured. The viability of LLC cells with living *L. casei* KJ686 decreased in a concentration-dependent manner, while LLC incubated with heat-killed *L. casei* KJ686 exhibited much the same viability as controls (Fig. 5).

2 Cytotoxic effect of *L. casei* KJ686 culture supernatant on LLC cells

Conditioned media in which *L. casei* KJ686 had been cultured for 24 hr and split into 6 fractions by

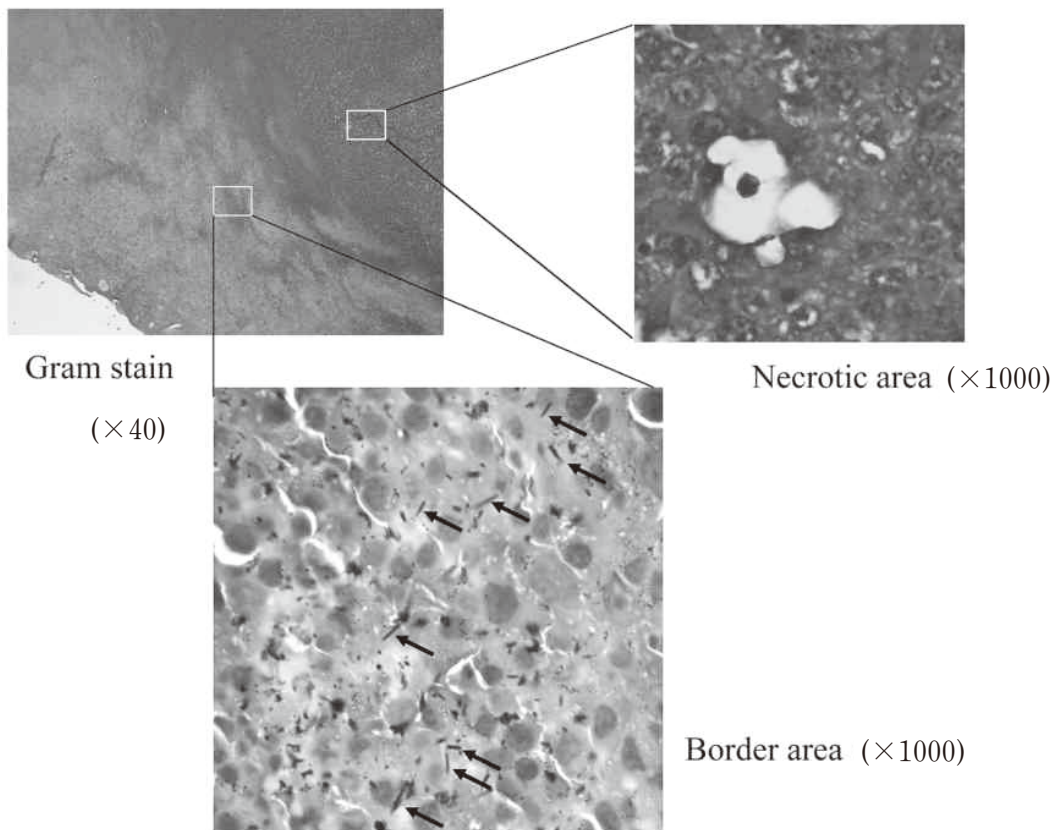


Fig. 2 Localization of *L.casei* KJ686 in tumors. On day 9, Gram's staining of histological sections indicated that *L.casei* KJ686 was scattered in the border areas between necrotic and viable regions.

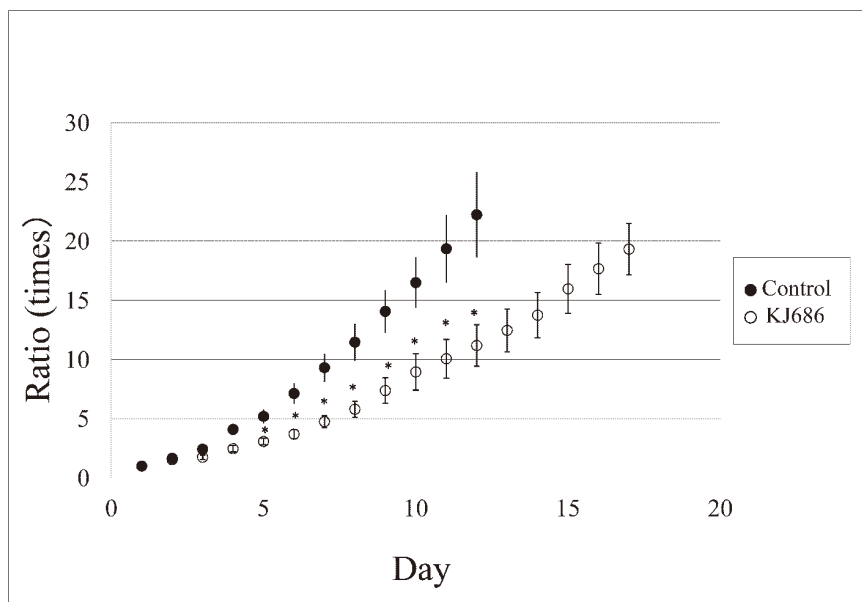


Fig. 3 *In vivo* tumor growth ratio. Controls were not given any treatment after tumor inoculation, while the test group was intravenously injected with *L.casei* KJ686. The tumor growth ratio became significantly different from day 5 (* $p < 0.01$).



Fig. 4 Gram's staining by the neo B & M method. The morphology of living and heat-killed *L. casei* KJ686 was not apparently altered.

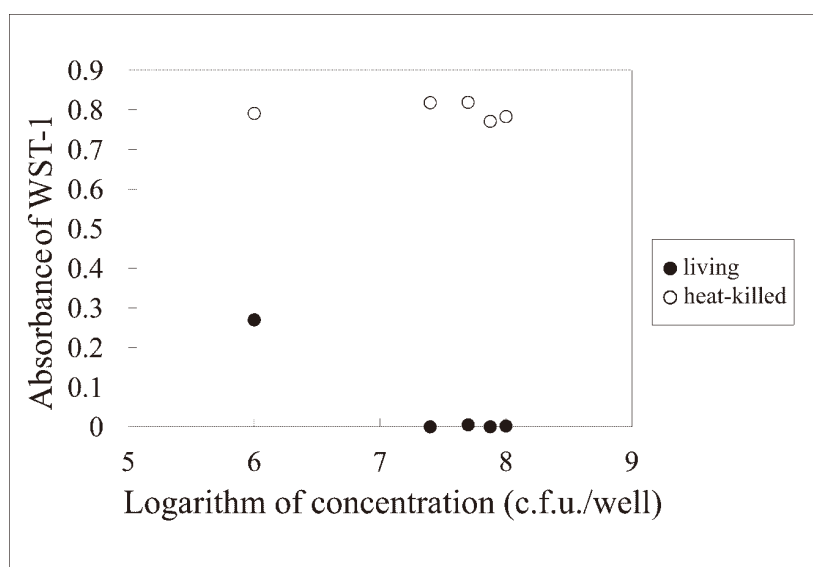


Fig. 5 Cytotoxic effect of *L. casei* KJ686 on LLC cells. The survival of cells treated with living *L. casei* KJ686 (●) was significantly less than those treated with heat-killed *L. casei* KJ686 (○). Untreated LLC cells exhibited much the same viability as heat-killed *L. casei* KJ686 cells.

molecular weight were evaluated for cytotoxic effect. The fraction with a molecular weight of less than 10 kDa exhibited a comparable cytotoxic effect as the original conditioned medium, while the other fractions did not (Fig. 6). We then compared the aqueous layer with the organic layer of the supernatant under 1 kDa, and found that only the aqueous layer had a significant cytotoxic effect (Fig. 7). However, no fraction of the under 1 kDa supernatant further separated by a hydrophobic resin column exhibited cytotoxicity (data not

shown). When conditioned medium and cytotoxic fractions were boiled at 100 °C for one minute, cytotoxicity showed only a slight decrease (data not shown). To test for pH-mediated cytotoxicity, we added a comparatively acidic pH4.0 lactic acid solution instead of the supernatant, but no cytotoxicity was observed (data not shown).

IV Discussion

Hypoxia in solid tumors is known to confer resistance to radiation and some chemotherapies, which

Direct anti-tumor effect of *L.casei* KJ686

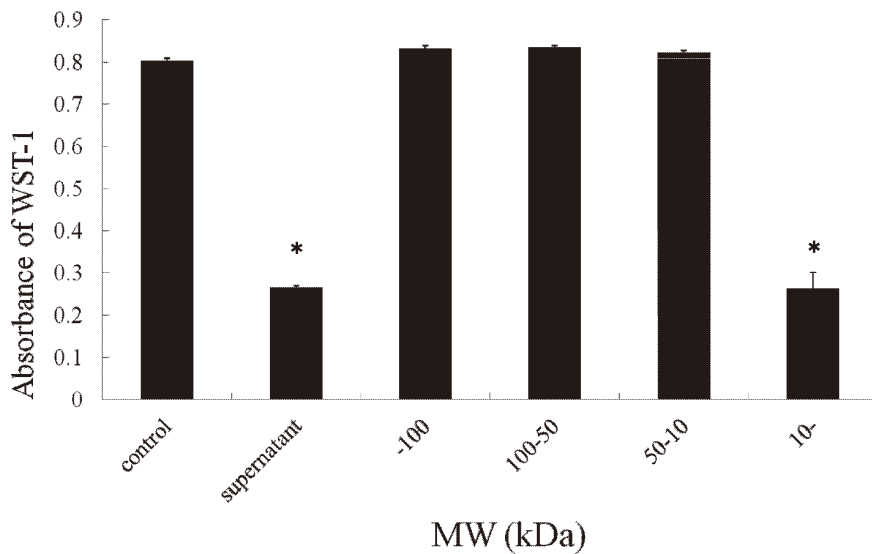


Fig. 6 Cytotoxic effect of the conditioned medium of *L.casei* KJ686 on LLC cells. The concentration of each fraction was adjusted to that of the original supernatant with DMEM (* $p < 0.01$ vs. control).

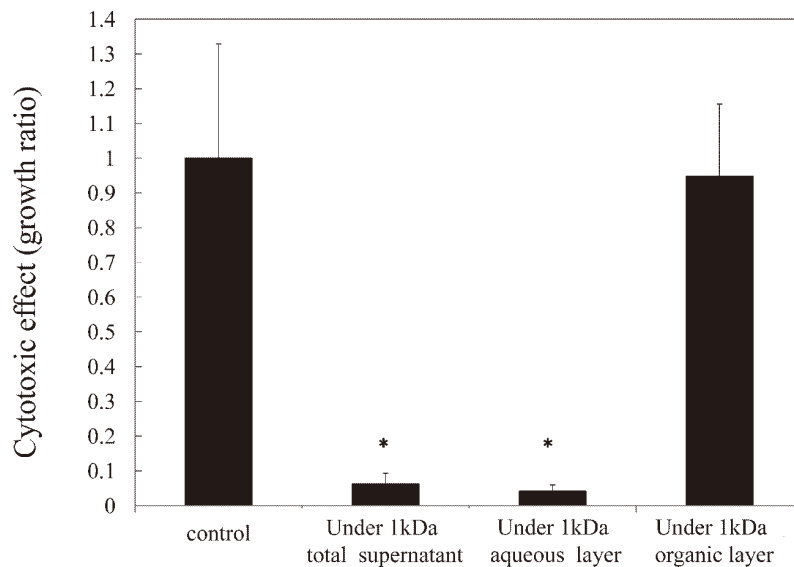


Fig. 7 Comparison of the cytotoxic effect of aqueous and organic layers of the conditioned medium fraction under 1 kDa. The concentration of each fraction was adjusted to that of the original supernatant with sterilized water (* $p < 0.05$ vs. control).

often results in treatment failure²⁰⁾²¹⁾. To overcome this impasse, several new means of delivering agents to solid tumors are being explored¹⁾⁻¹⁰⁾.

Intravenous injection of living bacteria is generally understood to induce bacteremia and sepsis. This was surprisingly not the case for *B.longum*¹⁾⁻⁴⁾, nor was it for *L.casei* KJ686, which accumulated specifically in tumor tissues without any adverse symptoms in the mice. This fact demonstrates that

L.casei KJ686 would be safely used for systemic treatment. Interestingly, a significant difference between *L.casei* KJ686 and *B.longum* is that the former has the ability to inhibit tumor growth by itself. We believe that this inhibition is due to the local effects of *L.casei* KJ686 and not immunomodulation. In previous studies, inoculation of a large dose of heat-killed bacteria had no direct anti-tumor effect, indicating that the property was not

attributed to host immunoresponses²⁵⁾²⁹⁾. Thus, we next investigated the direct anti-tumor effect of *L. casei* KJ686 *in vitro*. Following the addition of living *L. casei* KJ686, the growth of LLC cells was strongly inhibited, but this was not the case for heat-killed *L. casei* KJ686. Both living and heat-killed *L. casei* KJ686 exhibited the same morphology by Gram's stain, implying that heat treatment did not cause destruction of the cell wall. These results suggested that living *L. casei* KJ686 itself and/or substances excreted from *L. casei* KJ686 imparted a cytotoxic effect on LLC cells. We further investigated which fractions of the *L. casei* KJ686 culture supernatant had this effect. Manjunath et al. reported that a 40-fold concentrated bacterial culture supernatant exhibited cytotoxicity *in vitro*³⁰⁾. Although we used a culture supernatant that was not concentrated, we nonetheless witnessed that very low molecular weight (<1 kDa) excreted products of *L. casei* KJ686 had an inhibitory effect on the growth of LLC cells. As the inhibitory properties of the fraction were partially stable against both heat and lactic acid treatment, it can be said that *L. casei* KJ686 had a direct suppressive effect, possibly through several factors, on LLC cells rather than indirect stimulation of host immunoresponses.

In terms of safety, *L. casei* may have immunological tolerance because it can be found in the human intestinal tract just after birth³¹⁾. Although it was reported that *L. casei* sometimes induced bacteremia in patients with cancer, diabetes, broad spectrum antibiotic therapy, and transplantation,

there have been no reported cases of patient death if adequate treatment was performed³²⁾. In conclusion, it appears that specific cytotoxic substances present in the culture supernatant of *L. casei* KJ686 and their mode of action may be pivotal as direct anti-tumor agents once they have been identified.

V Acknowledgements

Our deep appreciation goes to Prof. Shun'ichiro Taniguchi of the Department of Molecular Oncology, Institute of Pathogenesis and Disease Prevention, Shinshu University Graduate School of Medicine, and Prof. Minoru Fujimori of the Department of Breast Surgery, Tokyo Medical University Ibaraki Medical Center, who provided carefully considered feedback and valuable comments. We also owe a very important obligation to Prof. Tomoyuki Fujita of the Department of Sciences of Functional Foods, Graduate School of Agriculture, Shinshu University, and Akihiro Shou of the Department of Molecular Oncology, Institute of Pathogenesis and Disease Prevention, Shinshu University Graduate School of Medicine, whose chemical analysis and technical assistance helped us greatly. Finally, we thank Dr. Shizunobu Igimi and Akinobu Kajikawa of the National Institute of Health Sciences for provision of the original stock culture of *Lactobacillus casei* KJ686 that was used in this study.

This study was supported in part by a grant-in-aid for scientific research from the Japan Society for the Promotion of Science (No. 21591678)

References

- 1) Yazawa K, Fujimori M, Amano J, Kano Y, Taniguchi S : Bifidobacterium longum as a delivery system for cancer gene therapy : selective localization and growth in hypoxic tumors. *Cancer Gene Ther* 7 : 269-274, 2000
- 2) Nakamura T, Sasaki T, Fujimori M, Yazawa K, Kano Y, Amano J, Taniguchi S : Cloned cytosine deaminase gene expression of Bifidobacterium longum and application to enzyme/pro-drug therapy of hypoxic solid tumors. *Biosci Biotechnol Biochem* 66 : 2362-2366, 2002
- 3) Sasaki T, Fujimori M, Hamaji Y, Hama Y, Ito K, Amano J, Taniguchi S : Genetically engineered Bifidobacterium longum for tumor-targeting enzyme-prodrug therapy of autochthonous mammary tumors in rats. *Cancer Sci* 97 : 649-657, 2006
- 4) Taniguchi S, Fujimori M, Sasaki T, Tsutsui H, Shimatani Y, Seki K, Amano J : Targeting solid tumors with non-pathogenic obligate anaerobic bacteria. *Cancer Sci* 101 : 1925-1932, 2010

- 5) Robbins PD, Chivizzani SC : Viral vectors for gene therapy. *Parmachol Ther* 80 : 35-47, 1998
- 6) Chawla SP, Chua VS, Fernandez L, Quon D, Blackwelder WC, Gordon EM, Hall FL : Advanced phase I/II studies of targeted gene delivery in vivo : intravenous REXIN-G for gemcitabine-resistant metastatic pancreatic cancer. *Mol Ther* 18 : 435-441, 2010
- 7) Grosse SM, Tagalakis AD, Mustapa MF, Elbs M, Meng QH, Mohammadi A, Tabor AB, Hailes HC, Hart SL : Tumor-specific gene transfer with receptor-mediated nanocomplexes modified by polyethylene glycol shielding and endosomally cleavable lipid and peptide linkers. *FASEB J* 24 : 2301-2313, 2010
- 8) Ji SR, Liu C, Zhang B, Yang F, Xu J, Long J, Jin C, Fu DL, Ni QX, Yu XJ : Carbon nanotubes in cancer diagnosis and therapy. *BBA Rev Can* 1806 : 29-35, 2010
- 9) Koppu S, Oh YJ, Edrada-Ebel R, Blatchford DR, Tetley L, Tate RJ, Dufès C : Tumor regression after systemic administration of a novel tumor-targeted gene delivery system carrying a therapeutic plasmid DNA. *J Control Release* 143 : 215-221, 2010
- 10) Dufès C, Keith WN, Bilslund A, Proutski I, Uchegbu IF, Schätzlein AG : Synthetic anticancer gene medicine exploits intrinsic antitumor activity of cationic vector to cure established tumors. *Cancer Res* 65 : 8079-8084, 2005
- 11) Salminen S, von Wright A, Morelli L, Marteau P, Brassart D, de Vos WM, Fondén R, Saxelin M, Collins K, Mogensen G, Birkeland SE, Mattila-Sandholm T : Demonstration of safety of probiotics -- a review. *Int J Food Microbiol* 44 : 93-106, 1998
- 12) Naidu AS, Bidlack W, Clemens RA : Probiotic spectra of lactic acid bacteria (LAB). *Crit Rev Food Sci Nutr* 39 : 13-126, 1999
- 13) Moulder JE, Rockwell S : Hypoxic fractions of solid tumors : experimental techniques, methods of analysis, and a survey of existing data. *Int J Radiat Oncol Biol Phys* 10 : 695-712, 1984
- 14) Vaupel PW, Hockel M : Tumor Oxygenation. Vaupel PW, Kelleher DK, Gunderoth M (eds), pp 219-232, New York, 1994
- 15) Brown JM : The hypoxic cell : a target for selective cancer therapy -- eighteenth Bruce F. Cain Memorial Award lecture. *Cancer Res* 59 : 5863-5870, 1999
- 16) Dewhirst MW, Cao Y, Moeller B : Cycling hypoxia and free radicals regulate angiogenesis and radiotherapy response. *Nat Rev Cancer* 8 : 425-437, 2008
- 17) Semenza GL : Hypoxia-inducible factor 1 : control of oxygen homeostasis in health and disease. *Pediatr Res* 49 : 614-617, 2001
- 18) Poellinger L, Johnson RS : HIF-1 and hypoxic response : the plot thickens. *Curr Opin Genet Dev* 14 : 81-85, 2004
- 19) Vaupel P : Hypoxia and aggressive tumor phenotype : implications for therapy and prognosis. *Oncologist* 13 : 21-26, 2008
- 20) Peng W, Pan F, Liu X, Ning S, Xu N, Meng F, Wang Y, Li C : Hypoxia stabilizes microtubule networks and decreases tumor cell chemosensitivity to anticancer drugs through Egr-1. *Anat Rec* 293 : 414-420, 2010
- 21) Gray LH, Conger AD, Ebert M, Hornsey S, Scott OC : The concentration of oxygen dissolved in tissues at the time of irradiation as a factor in radiotherapy. *Br J Radiol* 26 : 638-648, 1953
- 22) Takagi A, Ikemura H, Matsuzaki T, Sato M, Nomoto K, Morotomi M, Yokokura T : Relationship between the in vitro response of dendritic cells to *Lactobacillus* and prevention of tumorigenesis in the mouse. *J Gastroenterol* 43 : 661-669, 2008
- 23) Shida K, Suzuki T, Kiyoshima-Shibata J, Shimada S, Nanno M : Essential roles of monocytes in stimulating human peripheral blood mononuclear cells with *Lactobacillus casei* to produce cytokines and augment natural killer cell activity. *Clin Vaccine Immunol* 9 : 997-1003, 2006
- 24) Matsuzaki T : Immunomodulation by treatment with *Lactobacillus casei* strain Shirota. *Int J Food Microbiol* 41 : 133-140, 1998

- 25) Kato I, Kobayashi S, Yokokura T, Mutai M : Antitumor activity of *Lactobacillus casei* in mice. *Gan* (in Japanese) 72 : 517-523, 1981
- 26) Lee JW, Shin JG, Kim EH, Kang HE, Yim IB : Immunomodulatory and antitumor effects in vivo by the cytoplasmic fraction of *Lactobacillus casei* and *Bifidobacterium longum*. *J Vet Sci* 5 : 41-48, 2004
- 27) Seow SW, Rahmat JN, Mohamed AA, Mnhendran R, Lee YK, Bay BH : *Lactobacillus* species is more cytotoxic to human bladder cancer cells than *Mycobacterium Bovis* (bacillus Calmette-Guerin). *Journal Urol* 168 : 2236-2239, 2002
- 28) Fichera GA, Giese G : Non-immunologically-mediated cytotoxicity of *Lactobacillus casei* and its derivative peptidoglycan against tumor cell lines. *Cancer Lett* 85 : 93-103, 1994
- 29) Hashimoto S, Nagaoka M, Hayashi K, Yokokura T, Mutai M : Role of culture supernatant of cytotoxic/cytostatic macrophages in activation of murine resident peritoneal macrophages. *Cancer Immunol Immunother* 28 : 253-259, 1989
- 30) Manjunath N, Ranganathan B : A cytotoxic substance produced by a wild culture of *Lactobacillus casei* D-34 against tumour cells. *Indian J Exp Biol* 27 : 141-145, 1989
- 31) Sjögren YM, Jenmalm MC, Böttcher MF, Björkstén B, Sverremark-Ekström : Altered early infant gut microbiota in children developing allergy up to 5 years of age. *Clin Exp Allergy* 39 : 518-526, 2009
- 32) Cannon JP, Lee TA, Bolanos JT, Danziger LH : Pathogenic relevance of *Lactobacillus* : a retrospective review of over 200 cases. *Eur J Clin Microbiol Infect Dis* 24 : 31-40, 2005

(2015. 5. 19 received ; 2015. 8. 17 accepted)
