

**Relationship Between Expression of β 3-Adrenoceptor mRNA
in Bladder Mucosa and Urodynamic Findings
in Men with Lower Urinary Tract Symptoms**

Yoshiki Kurizaki,¹ Osamu Ishizuka,¹ Tetsuya Imamura,¹ Osamu Nishizawa,¹
and Karl-Erik Andersson²

¹*Department of Urology, Shinshu University School of Medicine, Matsumoto,
Japan*

²*Wake Forest Institute for Regenerative Medicine, Wake Forest University
School of Medicine, Winston Salem, NC, USA*

Correspondence: Osamu Ishizuka M.D., Ph.D.

Department of Urology

Shinshu University School of Medicine

3-1-1 Asahi, Matsumoto, JAPAN 390-8621

Tel.: +81 263 37 2661

Fax: +81 263 37 3082

e-mail: ishizuk@shinshu-u.ac.jp

Running head: β 3-AR mRNA in Bladder Mucosa and Urodynamic Findings

Abstract

Aims: To investigate the relationship between urinary bladder mucosal expression of β 3-adrenoceptor (AR) mRNA and urodynamic findings in patients with lower urinary tract symptoms and benign prostatic obstruction (BPO). **Methods:** During surgical prostate resection of 32 BPO patients, mucosal biopsies were collected and analyzed by reverse transcriptase polymerase chain reaction to determine the expression level of β 3-AR mRNA. First desire to void (FDV) and strong desire to void (SDV), detrusor overactivity (DO), and bladder outlet obstruction (BOO) were measured pre-operatively. Patients with FDVs <200 ml or SDVs <300 ml were assigned to the small capacity group (n=19). Patients with FDVs >201 ml and SDVs >301 ml were assigned to the large capacity group (n=13). The same patients with positive DO were also assigned to the DO+ group (n=11), and those with negative DO were assigned to the DO- group (n=21). Finally, patients whose position on the Schäfer nomogram was greater than degree V were assigned to the severe BOO group (n=17), while those with less than degree IV were assigned to the mild BOO group (n=15). **Results:** The expression level of β 3-AR mRNA was similar in both bladder capacity groups and both DO groups. However, the expression level in the severe BOO group was significantly less than in the mild BOO group (p=0.043). **Conclusions:** The expression of bladder mucosal β 3-AR mRNA was significantly decreased in patients with severe BOO, suggesting that β 3-ARs might be affected by the degree of BOO.

Key words

β 3-adrenoceptor, benign prostatic hyperplasia, bladder urothelium

INTRODUCTION

By sympathetic stimulation of β_3 -adrenoceptors (ARs) in the urinary bladder wall, urine can be stored under low pressure. Recently, bladder β_3 -ARs have become a therapeutic target for storage symptom issues. For example, a β_3 -AR selective agonist is effective for patients with symptoms such as urinary frequency, incontinence, and urgency.¹ In animal models with bladder outlet obstruction (BOO), there are few reports regarding the expression level of β_3 -AR mRNA in the bladder. Barendrecht et al., using quantitative real time PCR, reported that β_3 -AR mRNA was not up-regulated in rats 7 days after BOO.² However, Park et al., also using quantitative real time PCR, and Western blot, demonstrated an increase in β_3 -AR mRNA expression in rats with BOO for 8 weeks, although the increase was not statistically significant.³ In none of these studies, the mucosa and the detrusor muscle were analyzed separately. To the best of our knowledge, there are no reports that correlate the expression of β_3 -AR mRNA in the human bladder mucosa to urodynamic findings, especially in patients with lower urinary tract symptoms (LUTS) and benign prostatic obstruction (BPO). Thus, in this report we determined the bladder mucosal expression level of β_3 -AR mRNA in LUTS and BPO patients and correlated it with urodynamic parameters such as bladder capacity, detrusor overactivity (DO), and the degree of BOO.

METHODS

Patients

This study was performed with the approval of the Ethics Committee of School of Medicine, Shinshu University, and written informed consent was obtained from all patients. All patients were treated in accordance with the Declaration of Helsinki. None of the patients had any other significant health issues that required recent surgery or use of drugs that could affect the outcomes of this study.

Thirty-two patients with a diagnosis of BPO and who were scheduled for transurethral prostatectomy (TURP, n = 26) or retropubic prostatectomy (n = 6) at Shinshu University Hospital from October 2006 to October 2010 were enrolled in this study. Prostate specific antigen (PSA) was measured pre-operatively, and a prostatic needle biopsy was performed for the patients whose PSA values were more than 4.0 ng/ml. We did not perform biopsies in cases where the PSA elevation was thought to be due solely to prostate enlargement. Prostate volume was measured by ultrasonography.

The pre-operative international prostate symptom score (IPSS) and overactive bladder symptom score (OABSS) were recorded for each patient. The IPSS is composed of the sum score of seven symptoms: feeling of incomplete bladder emptying, frequency, intermittency, urgency, weak stream, straining, and nocturia. The IPSS is used for evaluating LUTS due to benign prostate hyperplasia (BPH). The OABSS is a composite based upon the sum

score of four symptoms: daytime frequency, nighttime frequency, urgency, and urgency incontinence.⁴

Real-time reverse transcription polymerase chain reaction (RT-PCR)

Tissue specimens were obtained from the mucosa of the posterior bladder wall during TURP or retropubic prostatectomy by means of transurethral or direct cold punch biopsy. The specimens were immediately put into the RNAlater (150 mg tissue/1.5 ml RNAlater Tissue Protect Tube; QIAGEN Inc., Valencia, CA, USA) and preserved at -20°C.

β_3 -AR mRNA was quantified by real-time RT-PCR (Bio-Rad Laboratories, Inc., Tokyo, Japan). Expression levels were determined using the ddCt value. Total RNA was extracted from the biopsied mucosa with the RNeasy Mini Kit (QIAGEN Inc.). Concentration of the total RNA was estimated by photometer (Naka Instruments, Co. Ltd., Hitachinaka, Japan). Complementary DNA (cDNA) was synthesized from 0.2 μ g of the total RNA with the High-Capacity cDNA Archive Kit (Applied Biosystems, Foster City, CA, USA). Real time RT-PCR of the cDNA was performed at 50°C for 2 minutes followed by 95°C for 10 minutes. These were followed by 40 cycles at 95°C for 15 seconds and 60°C for 1 minute. The following primer (Applied Biosystems) was used: ADRB3 (Rn00565393_m1) for β_3 -AR. Gene activity was expressed as the ratio to the internal standard eukaryotic ribosomal 18S ribosomal RNA (Applied Biosystems, Accession Number: X03205). The analysis was done in duplicate for each sample.

Video urodynamic study

A pre-operative video urodynamic study was performed for all patients. The study consisted of fluoroscopic monitoring of filling and voiding cystometry with synchronous sphincter electromyography (EMG) through a surface electrode placed on the perineum. A 14fr transurethral catheter (SAFEED Nelaton Catheter® TERUMO Co., Ltd., Tokyo, Japan) and a 4.7fr transurethral catheter (Dretler Urodynamic PFS Catheter®, Cook Urological, Inc., Spencer, IN, USA) were used for bladder filling and intravesical pressure recordings. Contrast medium (room-temperature, 30% meglumine iotalamate, Conray® Daiichi Pharmaceutical Co., Ltd., Tokyo, Japan) was instilled at a rate of 30 ml/min.

Among the cystometric variables that we recorded were the first desire to void (FDV) and the strong desire to void (SDV), which were defined by the International Continence Society.⁵ The volumes for FDV and SDV were recorded simultaneously with EMG activity using a computer-assisted urodynamic apparatus (Urovision®, Life-Tec Inc., Stafford, TX, USA). All examinations were carried out without any anesthesia or sedatives with the patients lying supine. According to criteria established by Wyndaele,^{6,7} we assigned patients with FDVs < 200 ml or SDVs < 300 ml to the small capacity group (SC group, n = 19). Patients with FDVs > 201 ml and SDVs > 301 ml were assigned to the large capacity group (LC group, n = 13). The same patients with positive DO were also assigned to the DO+ group (n=11), and those with negative DO were assigned to the DO- group (n=21). Finally, patients whose position on the Schäfer nomogram was greater than degree V

were assigned to the severe BOO group (n=17), while those with less than degree IV were assigned to the mild BOO group (n=15).

Statistical analysis

All values were calculated as means \pm standard error of means.

Comparisons were made by Mann-Whitney *U* test. P-values less than 0.05 were considered statistically significant.

RESULTS

All patients were diagnosed with benign prostatic hyperplasia. There were no significant differences in age, PSA, resected mass, pre-operative IPSS, or pre-operative OABSS between the paired groups SC vs. LC, DO+ vs. DO-, and severe BOO vs. mild BOO ($p > 0.05$ for each comparison, Table I). The FDV for the SC group was 140.64 ± 11.09 ml, which was significantly less than that of the LC group, 265.50 ± 25.00 ml ($p < 0.001$). Similarly, the SDV for the SC group, 220.21 ± 16.06 ml, was also significantly less than that of the LC group, 382.94 ± 20.43 ml ($p < 0.001$).

For $\beta 3$ -AR mRNA expression levels (ddCt), there were no significant differences in comparisons of SC vs. LC or DO+ vs. DO- groups (Fig. 1). However, patients with severe BOO expressed slightly, but significantly, lower levels of $\beta 3$ -AR mRNA than did those with mild BOO ($p = 0.043$, Fig. 1).

DISCUSSION

The β_3 -AR⁸ is the predominant β -AR in the human bladder⁹⁻¹¹, and has recently become a therapeutic target for bladder storage symptoms.^{12, 13} However, the site(s) of action and mechanisms involved have not been established. There are several structures in the bladder that could be responsive to β_3 -AR agents, including the detrusor smooth muscle, detrusor vasculature, afferent and efferent nerve terminals, intramural ganglia, urothelium, and interstitial cells.¹⁴

Nomiya et al.¹⁵ reported that 97% of β -AR mRNA expressed in human bladder detrusor is β_3 -AR, and 1.5% and 1.4% of β -AR mRNA are β_1 -AR and β_2 -AR mRNAs, respectively. In that report, they demonstrated that β_3 -AR mRNA was slightly, but not significantly, up-regulated in the obstructed bladder group. Masunaga et al.¹⁶ reported that in pig bladder, β_3 -ARs mediate the inhibitory effect of agonists on detrusor contractions via the urothelium. Also Otsuka et al. who reported the presence of β_1 -, β_2 - and β_3 -ARs in human bladder urothelium¹⁷ suggested that β -AR agonists stimulated the release of an unidentified inhibitory factor from the urothelium that reduced detrusor contraction.

Based on these reports, we found it of interest to investigate the possible relationship between urodynamic findings and the mucosal levels of β_3 -AR mRNA. Between the SC and LC groups, and between the DO+ and DO- groups, there were no significant differences in the expression of mucosal β_3 -AR mRNA. In contrast, in the severe BOO group, the expression

level of β 3-AR mRNA in the mucosa was significantly lower than that in the mild BOO group, suggesting that the expression of β 3-AR mRNA may be dependent on the degree of obstruction. Both the degree of obstruction and the bladder changes induced by obstruction may be time-dependent. In the animal studies of Park et al. and Barendrecht et al., mentioned previously, the expression of β 3-AR mRNA was studied at different time points after the induction of BOO.^{2,3} Barendrecht et al. studied their animals 7 days after the obstruction, whereas Park et al. made their analyses 8 weeks after BOO induction. No significant changes in the expression of β 3-AR mRNA were found at any of the time points, and this would not support the speculation that time-dependent bladder changes are of importance for the β 3-AR mRNA expression. However, in neither of these studies, the mucosa was analyzed separately from the detrusor muscle. This was done in the present study, showing a significantly lower β 3-AR mRNA expression in the severe BOO group. However, it is difficult to compare the results of obstruction in humans and in rats, and the clinical relevance of the present finding may be questioned, since it was not related to the occurrence of DO or symptom scores. On the other hand, mucosal β 3-ARs may be a target for therapeutically administered β 3-AR agonists. If the occurrence of β 3-AR mRNA corresponds to the presence of receptor protein, this could mean that severely obstructed men could be less responsive to such treatment than those with mild obstruction.

We are aware that our study has several limitations. The number of subjects is relatively small. Also, because of the limited amount of tissue contained in each biopsy, we did not perform Western blot analyses or

immunohistochemistry for the presence of the β 3-AR proteins. Thus, our findings may or may not accurately reflect the level of the receptors themselves, which can be determined by post-translational regulation and the rate of receptor degradation. Further studies are required to confirm our findings and to quantify the amounts of β 3 -ARs in both normal bladders and bladders from patients with BPO and LUTS to assess the clinical relevance, if any, of our observations.

CONCLUSIONS

In BPO patients with LUTS, there was no relationship between the expression of urinary bladder mucosal β 3-AR mRNA and bladder capacity or the existence of DO measured during urodynamic study. On the other hand, β 3-AR mRNA in the bladder mucosa of patients with severe BOO ($\geq V$) was significantly less than in patients with mild BOO ($\leq IV$). These results suggest that the presence of β 3-ARs might be affected by the degree of BOO. The exact function of β 3-ARs in the bladder urothelium awaits further investigation.

Disclosure

None of the authors have any conflicting interests. This study was not supported by any federal or industrial resources.

REFERENCES

1. Chapple C, Wyndaele JJ, Van Kerrebroeck P, Radziszewski P, Dvorak V, Boerrigter P. Dose-ranging study of once-daily mirabegron (YM178), a novel selective β_3 -adrenoceptor agonist, in patients with overactive bladder (OAB). *Eur Urol* 2010; (Supple 9):249.
2. Park MG, Park HS, Lee JG, Kim HJ. Changes in awake cystometry and expression of bladder β -adrenoceptors after partial bladder outlet obstruction in male rats. *Int Neurourol J* 2010; 14:157-163.
3. Barendrecht MM, Frazier EP, Vrydag W, Alewijnse AE, Peters SLM, Michel MC. The effect of bladder outlet obstruction on α_1 - and β -adrenoceptor expression and function. *Neurourol Urodyn* 2009; 28:349-355.
4. Homma Y, Yoshida M, Seki N, Yokoyama O, Kakizaki H, Gotoh M, et al. Symptom assessment tool for overactive bladder syndrome – overactive bladder symptom score. *Urology* 2006; 68:318–323.
5. Abrams P, Cardozo L, Fall M, Griffiths D, Rosier P, Ulmsten U, et al. Standardisation Sub-committee of the International Continence Society. The standardization of terminology of lower urinary tract function: Report from the Standardization Sub-committee of the International Continence Society. *Neurourol Urodyn* 2002; 21:167-178.

6. Wyndaele JJ. The normal pattern of perception of bladder filling during cystometry studied in 38 young healthy volunteers. *J Urol* 1998; 160:479-481.
7. Wyndaele JJ, De Wachter S. Cystometrical sensory data from a normal population: Comparison of two groups of young healthy volunteers examined with 5 years interval. *Eur Urol* 2002; 42:34-38.
8. Emorine LJ, Marullo S, Briand-Sutren MM, Patey G, Take K, Delavier-Klutchko C, et al. Molecular characterization of the human beta 3-adrenergic receptor. *Science* 1989; 245:1118-1121.
9. Igawa Y, Yamazaki Y, Takeda H, Hayakawa K, Akahane M, Ajisawa Y, et al. Functional and molecular biological evidence for a possible beta3-adrenoceptor in the human detrusor muscle. *Br J Pharmacol* 1999; 126:819-825.
10. Takeda M, Obara K, Mizusawa T, Tomita Y, Arai K, Tsutsui T, et al. Evidence for beta3-adrenoceptor subtypes in relaxation of the human urinary bladder detrusor: analysis by molecular biological and pharmacological methods. *J Pharmacol Exp Ther* 1999; 288:1367-1373.

11. Fujimura T, Tamura K, Tsutsumi T, Yamamoto T, Nakamura K, Koibuchi Y, et al. Expression and possible functional role of the beta3-adrenoceptor in human and rat detrusor muscle. *J Urol* 1999; 161:680-685.
12. Andersson KE, Wein AJ. Pharmacology of the lower urinary tract: basis for current and future treatments of urinary incontinence. *Pharmacol Rev* 2004; 56:581-631.
13. Yamaguchi O, Chapple CR. Beta3-adrenoceptors in urinary bladder. *Neurourol Urodyn* 2007; 26:752-756.
14. Andersson KE, Gratzke C. Pharmacology of alpha 1-adrenoceptor antagonists in the lower urinary tract and central nervous system. *Nat Clin Pract Urol* 2007; 4:368-378.
15. Nomiya M, Yamaguchi O. A quantitative analysis of mRNA expression of alpha 1 and beta-adrenoceptor subtypes and their functional roles in human normal and obstructed bladders. *J Urol* 2003; 170:649-653.
16. Masunaga K, Chapple CR, McKay NG, Yoshida M, Sellers DJ. The β_3 adrenoceptor mediates the inhibitory effects of β -adrenoceptor agonists via the urothelium in pig bladder dome. *Neurourol Urodyn* 2010; 29:1320-1325.

17. Otsuka A, Shinbo H, Matsumoto R, Kurita Y, Ozono S. Expression and functional role of beta-adrenoceptors in the human urinary bladder urothelium. *Naunyn Schmiedebergs Arch Pharmacol.* 2008; 377:473-481.

Figure Caption

Figure 1. Expression level of β 3-AR mRNA. There were no significant differences in β 3-AR mRNA expression between bladders of the Small Capacity and Large Capacity groups (A) or between the DO+ and DO- groups (B). However expression was significantly less in patients with severe BOO (\leq IV) group compared to the patients with mild BOO (\leq IV) group (C) ($p = 0.043$).