

Phospholipase C β 3 Expressed in Mouse DRGs is Involved in Inflammatory and Postoperative Pain

This article was published in the following Dove Press journal:
Journal of Pain Research

Susumu Ide¹
Tomoyuki Kawamata^{1,2}
Kumiko Ishida¹
Satoshi Fuseya¹
Takashi Ishida¹
Yuki Sugiyama¹
Mikito Kawamata¹
Satoshi Tanaka¹

¹Department of Anesthesiology and Resuscitology, Shinshu University School of Medicine, Matsumoto, Nagano, Japan;
²Department of Anesthesiology, Wakayama Medical University, Wakayama, Japan

Background: Previous studies suggested that phospholipase C β 3 (PLC β 3), which is a common downstream component in the signaling cascade, plays an important role in peripheral mechanisms of perception including nociception. However, detailed profiles of PLC β 3-expressing dorsal root ganglion (DRG) neurons and involvement of PLC β 3 in inflammatory and postoperative pain have not been fully investigated.

Purpose: We evaluated neurochemical characteristics of PLC β 3-expressing DRG neurons in mice and then we examined the effects of selective knockdown of PLC β 3 expression in DRGs on inflammatory and postoperative pain.

Methods: Male C57BL/6-strain mice were used. For the inflammatory model, each mouse received subcutaneous injection of complete Freund's adjuvant (CFA) in the left hindpaw. For the postoperative pain model, a plantar incision was made in the left hindpaw. PLC β 3 antisense oligodeoxynucleotide or PLC β 3 mismatch oligodeoxynucleotide was intrathecally administered once a day for three consecutive days in each model. The time courses of thermal hyperalgesia and mechanical hyperalgesia were investigated. Changes in PLC β 3 protein levels in DRGs were evaluated by Western blotting.

Results: Immunohistochemical analysis showed that high proportion of the PLC β 3-positive profiles were biotinylated isolectin B4-positive or transient receptor potential vanilloid subfamily 1-positive. PLC β 3 protein level in DRGs during CFA-induced inflammation was comparable to that at baseline. Intrathecal administration of PLC β 3 antisense oligodeoxynucleotide, which significantly suppressed PLC β 3 expression in DRGs, did not affect pain thresholds in normal conditions but inhibited CFA-induced thermal and mechanical hyperalgesia both at the early and late phases compared to that in mismatch oligodeoxynucleotide-treated mice. Intrathecal administration of PLC β 3 antisense oligodeoxynucleotide also inhibited surgical incision-induced thermal and mechanical hyperalgesia.

Conclusion: Our results uncover a unique role of PLC β 3 in the development and maintenance of inflammatory pain induced by CFA application and in those of surgical incision-induced pain, although PLC β 3 does not play a major role in thermal nociception or mechanical nociception in normal conditions.

Keywords: phospholipase C β 3, inflammatory pain, postoperative pain, thermal hyperalgesia, mechanical hyperalgesia

Correspondence: Susumu Ide; Satoshi Tanaka
Department of Anesthesiology and Resuscitology, Shinshu University School of Medicine, 3-1-1 Asahi, Matsumoto, Nagano 390-8621, Japan
Tel +812-6337-2670
Fax +812-6335-2734
Email ide@shinshu-u.ac.jp;
s_tanaka@shinshu-u.ac.jp

Introduction

Peripheral inflammation leads to the release of chemical mediators including adenosine triphosphate, bradykinin, trypsin/tryptase, endothelin-1, and prostaglandins, which play important roles in peripheral mechanisms of an inflammation-induced state of long-lasting pain.¹ These chemical mediators activate G protein-coupled receptors (GPCRs) coupling to Gq/11 expressed on dorsal root ganglion (DRG) neurons.

Then, upon ligand binding, GPCRs modulate the excitability of nociceptors through an intracellular signaling network.¹ Since phospholipase C β (PLC β) links GPCRs as a common downstream component in the signaling cascade,² targeting PLC β may be more effective than targeting individual chemical mediators for reducing pain.

Four major G protein-coupled PLC β isoforms (PLC β 1, PLC β 2, PLC β 3, and PLC β 4) have been characterized, and PLC β 1, PLC β 3 and PLC β 4, but not PLC β 2, have been shown to be expressed in DRG neurons.³ While PLC β 1 and PLC β 4 are expressed in both DRG neurons and spinal neurons,³ PLC β 3 is preferentially expressed in DRG neurons and keratinocytes^{4,5} and in the brain^{6,7} but not in spinal neurons.³ In DRG neurons, PLC β 3 has the highest level of expression among PLC β 1, PLC β 3 and PLC β 4,³ suggesting that PLC β 3 plays an important role in peripheral mechanisms of perception including nociception.

Previous studies showed that itch sensation was reduced but that thermal and cold nociception and thermal hyperalgesia caused by injection of complete Freund's adjuvant (CFA) were not affected by PLC β 3 whole-body knockout in mice.^{3,8} In contrast, another previous study showed that development of carrageenan-induced mechanical hyperalgesia was suppressed in PLC β 3 knockdown mice in which PLC β 3 in DRGs was knocked down by antisense oligodeoxynucleotide (ODN).⁹ Thus, the contribution of PLC β 3 in DRG neurons to inflammatory pain perception is still controversial. Involvement of PLC β 3 in inflammatory pain may vary depending on the degree and stage of inflammation. Postoperative pain is another pain condition in which inflammation around the surgical site is involved but is distinct from pure inflammatory pain because the pathogenesis of postoperative pain may include not only inflammatory but also neuropathic mechanisms.¹⁰ The contribution of PLC β 3 in DRG neurons to postoperative pain has not yet been elucidated.

The purpose of this study was to clarify the role of PLC β 3 in the two types of nociceptive pain, that is, inflammatory pain and postoperative pain. Inflammatory pain and postoperative pain share several pain-related phenotypes but might have different underlying signaling mechanisms.^{11,12} In this study, we investigated the expression profile of PLC β 3 in DRG neurons and the effects of selective knockdown of PLC β 3 expression in DRGs on CFA-induced pain at the early and late phases. We also examined the effects of selective knockdown of PLC β 3 expression in DRGs on surgical incision-induced pain.

Materials and Methods

All of the protocols of this study were approved by the Animal Care and Use Committee of Shinshu University School of Medicine, Matsumoto, Japan (reference No. 240050) on January 22, 2013. Mice were treated in accordance with the Ethics Guidelines for Investigations of Experimental Pain in Conscious Animals as issued by the International Association for the Study of Pain. Every effort was made to minimize animal suffering and to reduce the number of animals used in this study.

Animals

We used male C57BL/6-strain mice (18–20 g, Japan SLC, Hamamatsu, Japan). Mice housed in groups of 4 animals were maintained on a 12-h light/12-h dark cycle with food and water available ad libitum.

Inflammatory Pain Model

Under general anesthesia (1–2% isoflurane in 100% oxygen), each mouse received subcutaneous injection of a 30- μ L emulsion of undiluted CFA (Sigma-Aldrich, St. Louis, MO, USA) in the medial left plantar hindpaw using a Hamilton syringe with a 30-gauge needle. After the injection, anesthesia was discontinued and mice were allowed to recover in their cages.

Postoperative Pain Model

Under general anesthesia (2% isoflurane in 100% oxygen), a plantar incision was made according to previous reports.^{11,13} After sterile preparation of the left hindpaw, a 6-mm longitudinal incision was made with a number 11 scalpel blade through the skin, fascia, and muscle. The incision was started 2 mm from the proximal edge of the heel and extended towards the toes. After wound hemostasis, the skin was sutured with two single sutures of 7–0 nylon. The mice were allowed to recover in cages with sterile bedding after surgery.

Oligodeoxynucleotide Preparation and Intrathecal Administration

The PLC β 3 antisense ODN sequence was directed against a unique region of the mouse PLC β 3 sequence (Gen Bank Accession No. NM008874). The mismatch ODN sequence was designed as the antisense ODN sequence scrambled. The antisense ODN and mismatch ODN sequences used in this study were 5'-TGGTGGTCATCTGGGATGTA-3' and 5'-TTGCGGTGAGTAGTTCGTAG-3', respectively. The

hemagglutinating virus of the Japan envelope vector system (HVJ Envelope Vector Kit GENOMEONE-Neo; Ishihara Sangyo, Osaka, Japan) was used for *in vivo* transfection of the ODNs. This HVJ Envelope (HVJ-E) vector has been proven to be an effective ODN delivery system both *in vitro* and *in vivo*.¹⁴ The ODNs were incorporated into the HVJ-E vector according to the manufacturer's instructions. The final concentration of the ODNs was adjusted to 2 µg/µL.

For intrathecal administration of the ODNs, mice were anesthetized and implanted with an intrathecal catheter according to our previous report.¹⁵ Briefly, under general anesthesia (1–2% isoflurane in 100% oxygen), a polyurethane intrathecal catheter with an inner diameter of 0.35 mm and an outer diameter of 0.84 mm (R-ITC, Pittsburgh, PA, USA) was inserted 5 mm into the mouse lumbar subarachnoid space at the L4/5 intervertebral. To administer drugs intrathecally, the tip of the catheter was located near the lumbar enlargement of the spinal cord. The catheter was tunneled subcutaneously and externalized through the skin in the neck region. The volume of dead space of the intrathecal catheter was 2 µL. After catheter implantation, the effects of intrathecal lidocaine (2%, 2 µL) were examined. Only mice that had shown complete paralysis of the tail and bilateral hind legs after intrathecal lidocaine injection were used in subsequent experiments. Intrathecal drug administration was accomplished using a microinjection syringe (Hamilton, Reno, NV, USA) connected to an intrathecal catheter under brief general anesthesia (1–2% isoflurane in 100% oxygen). The ODNs were administered manually over a 10-s period in a single injection volume of 2 µL (4 µg of the ODN) followed by a flush of physiological saline (3 µL). After discontinuance of general anesthesia, mice fully recovered within 2–3 min.

Thermal Testing

Mice were placed in plastic chambers on a glass surface and allowed to habituate for 30 min. Paw withdrawal latency (PWL) to noxious heat stimuli was assessed by applying a focused radiant heat source (model number 37370; Ugo Basil, Comerio, Italy) to unrestrained mice.¹⁶ PWLs were recorded as an average of 3 trials per animal with an interval between trials of more than 5 min. The intensity of the heat was adjusted so that the basal PWL was 10–15 s. The cut-off time was 20 s in order to avoid tissue damage. The person performing the behavioral tests was blinded to the treatment.

Mechanical Testing

Mice were placed on a stainless mesh floor covered with a clear plastic cage top. For testing mechanical responses, calibrated von Frey filaments (0.068-, 0.166-, 0.407-, 0.692-, 1.202-, 2.041-, and 3.630-g forces) were applied to the plantar surface in unrestrained mice. Each filament was applied at least 30 s between applications. Testing was initiated with a 1.202-g force. Depending on the initial response, subsequent filaments were applied by the “up-down method” to determine the paw withdrawal threshold (PWT).^{11,17}

Immunohistochemistry

We used polyclonal antibodies raised against the following molecules: goat anti-PLCβ3 (1.0 µg/mL, Frontier Science, Sapporo, Japan), guinea-pig anti-transient receptor potential vanilloid subfamily 1 (TRPV1) (0.1 µg/mL, provided by Dr. Watanabe, Hokkaido University, Japan), and mouse anti-neurofilament 200 kD (NF200, a marker of myelinated neurons) (1:4000, N0142, Sigma-Aldrich). The specificities of the goat anti-PLCβ3 antibody and the mouse anti-NF200 antibody have been confirmed previously.^{7,16,18} Our preliminary study using TRPV1-deficient mice also confirmed the specificity of the guinea-pig anti-TRPV1 antibody. In addition, we used biotinylated isolectin B4 (IB4) (1:100, L3759, Sigma-Aldrich). Mice were deeply anesthetized with urethane (1.25 g/kg intraperitoneally) and perfused transcardially with 4% paraformaldehyde (PFA) in 0.1 M phosphate buffer (PB). L4 and L5 DRGs were removed. The samples were immersed in 4% PFA in 0.1 M PB for 2 h for post-fixation and then cryoprotected in 25% sucrose in 0.01 M phosphate-buffered saline (PBS) overnight at 4°C. The samples were placed in Tissue-Tek embedding medium (Sakura, Tokyo, Japan) and rapidly frozen. The frozen samples of DRGs were cut into 16-µm sections using a sliding cryostat (LEICA, Tokyo, Japan). The tissue sections were thaw-mounted onto gelatin-coated slides for processing. The sections were washed in 0.01 M PBS for 30 min followed by 0.2% Triton X-100 (Sigma-Aldrich) in 0.01 M phosphate-buffered saline (PBS-t) for 30 min and incubated for 60 min at room temperature in a blocking solution consisting of 1% normal donkey serum and 0.01 M PBS-t. Then the sections were incubated with mixtures of primary antibodies in the blocking solution overnight at 4°C. After rinses with 0.01 M PBS-t, the slices were incubated with Alexa Fluor 488-, Alexa 597-, or Alexa 649-labelled species-specific secondary antibodies (Invitrogen, Carlsbad, CA, USA) at a dilution of

1:500 in PBS-t for 2 h at room temperature. Photographs were taken with a confocal laser-scanning microscope (ECLIPSE C1, Nikon, Tokyo, Japan).

Immunoblot Analysis

The mice were deeply anesthetized with isoflurane and sacrificed by decapitation. The L4, L5 and L6 DRGs were rapidly removed and homogenized in the presence of 150 μ L of Extraction Buffer 1 (ProteoExtract[®], Merck, Germany) containing a protease inhibitor cocktail on ice. The crude homogenates were incubated for 10 min and centrifuged at 1,000 g for 5 min at 4°C. Supernatants of the homogenates were collected, and the protein concentration was determined using a DC protein assay (Bio-Rad Laboratories, Hercules, CA, USA). The samples were heated for 10 min at 100°C with sodium dodecylsulfate-polyacrylamide gel electrophoresis (SDS-PAGE) sample buffer. Equal amounts of the proteins (10 μ g) were loaded onto a 4–15% SDS-PAGE separation gel and transferred to nitrocellulose membranes (Millipore, Billerica, MA, USA). The membranes were incubated with rabbit anti-PLC β 1 (1.0 μ g/mL, Frontier Science), guinea-pig anti-PLC β 3 (1.0 μ g/mL, Frontier Science), or rabbit anti-PLC β 4 (1.0 μ g/mL, Frontier Science) in PBS containing 10% skim milk overnight at 4°C. Concentrations of actin, a housekeeping protein, were also measured using a rabbit anti-actin antibody (1:5,000, A2066, Sigma-Aldrich). Immunoreaction was visualized with an enhanced chemiluminescence plus Western blotting detection system (Amersham Biosciences, Piscataway, NJ, USA). The reaction product was visualized using an image analyzer (LAS-3000mini, Fuji Film, Tokyo, Japan). Previous studies showed that the PLC β 1, PLC β 3 and PLC β 4 antibodies used in this study recognized single protein bands corresponding to expected molecular weights (150 kDa, 157 kDa and 134 kDa, respectively).^{7,19,20} The person performing the experiments was blinded to the treatment. The amount of protein that can be obtained from 3 DRGs (left L4/5/6) of one mouse is very small and not sufficient for immunoblot analysis. Therefore, L4, L5, and L6 DRGs isolated from 2 mice were pooled for each sample, and data from Western blot analysis of PLC β 3 in DRGs were analyzed from 8 mice per group.

Distribution of PLC β 3 in DRG Neurons

To examine the distribution of PLC β 3, we performed multiple immunostainings of PLC β 3, TRPV1, IB4 and NF200 in naïve mouse DRGs. Only neurons with clearly visible nuclei were counted using a computerized image analysis system (EZ-C1 3.90, Nikon). The proportion of

colocalization of PLC β 3-positive profiles with TRPV1-positive or IB4-positive neurons was determined by counting 300–600 neuronal profiles from 4–7 DRG sections for each mouse. Because a stereological approach was not used in this study, quantification of data may have yielded biased estimates of the actual numbers of cells and neurons. To prevent duplicate counting of neuronal cell bodies, sections that were 112 μ m apart were counted for each DRG. An assistant who was unaware of the treatment groups of sections performed all of the counting.

Time Course of Hyperalgesia and PLC β 3 Expression Following CFA-Induced Inflammation

In order to know the time course of hyperalgesia after CFA injection, the following study was performed in naïve mice. After obtaining basal PWL and PWT, a 30- μ L emulsion of undiluted CFA (Sigma-Aldrich) was injected in the medial left plantar hindpaw using a Hamilton syringe with a 30-gauge needle. To examine the involvement of PLC β 3 both in the early and late phases of CFA-induced inflammatory pain, PWLs and PWTs were measured before the injection and at 4 h, 6 h, 12 h, 24 h, 2 days, 3 days, and 7 days after the CFA injection. In some mice, the expression of PLC β 3 proteins in L4/5/6 DRGs was examined using Western blot analysis before and 24 h, 3 days, and 7 days after CFA injection.

Involvement of PLC β 3 in CFA-Induced Hyperalgesia

First, we examined the subtype specificity of the PLC β 3 antisense ODN in naïve mice. We tested effects of the PLC β 3 antisense ODN on not only expression of PLC β 3 but also expression of PLC β 1 or PLC β 4. Mice received single daily intrathecal administration of the PLC β 3 antisense ODN or the mismatch ODN for 3 consecutive days. Expression of PLC β 1, β 3 and β 4 proteins in L4/5/6 DRGs was examined using Western blot analysis 24 h and 48 h after the last administration of the antisense ODN or mismatch ODN.

Next, to examine the role of PLC β 3 in the development of CFA-induced hyperalgesia, mice received single daily intrathecal administration of the PLC β 3 antisense ODN or the mismatch ODN for 3 consecutive days before CFA injection. On the last day of ODN administration, CFA was injected into the plantar surface of the left hindpaw. Behaviors were assessed before ODN administration,

before CFA injection and at 4 h, 6 h, 12 h, 24 h, 2 days, and 3 days after CFA injection. In some mice, expression of PLC β 3 was assessed 24 h after the last administration of the ODN. In addition, to examine the role of PLC β 3 in the late phase of CFA-induced hyperalgesia, mice received the PLC β 3 antisense ODN or the mismatch ODN once daily at 4 days, 5 days, and 6 days after CFA injection. Behaviors were assessed before CFA injection and at 1 day, 3 days, 7 days, and 8 days after the injection. In some mice, expression of PLC β 3 was assessed 24 h after the last administration of the ODN.

Involvement of PLC β 3 in Postoperative Pain

In order to know the time course of hyperalgesia after plantar incision, the following study was performed in naïve mice. After obtaining the basal PWL and PWT, a plantar incision was made in the left hindpaw. PWLs and PWTs were measured before the incision and at 2 h, 4 h, 6 h, 24 h, 2 days, 3 days, 5 days, and 7 days after the incision.

To examine the role of PLC β 3 in pain behaviors after plantar incision, mice received single daily intrathecal administration of the PLC β 3 antisense ODN or the mismatch ODN for 3 consecutive days before plantar incision. On the last day of ODN administration, a plantar incision was made in the left hindpaw. Behaviors were assessed before the ODN administration, before the incision and at 2 h, 4 h, 6 h, 24 h, 2 days, 3 days, 5 days, and 7 days after the incision.

Statistical Analysis

All results are expressed as means \pm standard deviation (SD). A one-way analysis of variance for repeated measures followed by Dunnett's test was used for within-group comparison. A two-way analysis of variance for repeated measures followed by Tukey's test was used for between-group comparison. For immunoblot analysis, the intensity of bands was determined using an image densitometer (NIH Image 1.63; National Institutes of Health, Bethesda, MD, USA) and was normalized to the intensity of β -actin. The relative intensities of the bands were compared using the Mann-Whitney *U*-test. $P < 0.05$ was considered statistically significant. Statistical analysis was performed using Statview 5.0 software (Abacus Concepts, Berkeley, CA, USA) and NP Multi (Nagata T, Tokyo, Japan).

Results

Distribution of PLC β 3 Expression in DRG Neurons

In naïve mice, most of the PLC β 3-positive neurons did not show NF200 immunoreactivity (Figure 1A). NF200-positive neurons were observed in 1.5% of all PLC β 3-positive DRG neurons (5 of the 333 PLC β 3-positive neurons). We performed triple immunostaining with PLC β 3, TRPV1, and IB4. A total of 1571 PLC β 3-positive DRG neurons were analyzed from 18 fluorescent slices from 4 mice. The results showed that 52% of TRPV1-expressing DRG neurons were PLC β 3-positive and 96% of IB4-expressing DRG neurons were PLC β 3-positive. 64%, 30% and 4% of PLC β 3-expressing DRG neurons were IB4-positive, TRPV1-positive, and both IB4- and TRPV1-positive, respectively (Figures 1B and C and [Supplementary Figure 1](#)).

Time Courses of Hyperalgesia and PLC β 3 Expression in DRGs After CFA-Induced Inflammation

After an intraplantar CFA injection, ipsilateral PWL in response to noxious heat stimuli and ipsilateral PWT in response to mechanical stimuli by von Frey filaments were significantly decreased compared to basal values (before injection) ($P < 0.01$). Significant decreases in PWL and PWT were evident 4 h after CFA injection and lasted up to 7 days (Figure 2A and B). On the other hand, the relative intensity of PLC β 3/ β -actin in L4/5/6 DRGs was not significantly changed at 24 h, 3 days, or 7 days after CFA injection compared to the basal value ($P = 0.24, 0.24, 1.00$, respectively) (Figure 2C).

Specificity of the PLC β 3 Antisense ODN

PLC β 3 antisense or mismatch ODNs were administered intrathecally once a day for 3 consecutive days. Intrathecal PLC β 3 antisense ODN, but not mismatch ODN, significantly decreased the relative intensity of PLC β 3 to β -actin in L4/5/6 DRGs compared to that in naïve mice ($P = 0.02$ and 0.31 , respectively) (Figure 3A). PLC β 3 antisense ODN significantly decreased the relative intensity of PLC β 3/ β -actin to about 60% of that in naïve mice at 24 h after the last administration of the ODN. The relative intensity in antisense-treated mice at 48 h after the last administration of the ODN was comparable to that in naïve mice. On the other hand, PLC β 3 antisense and mismatch ODNs did not change the relative intensity of

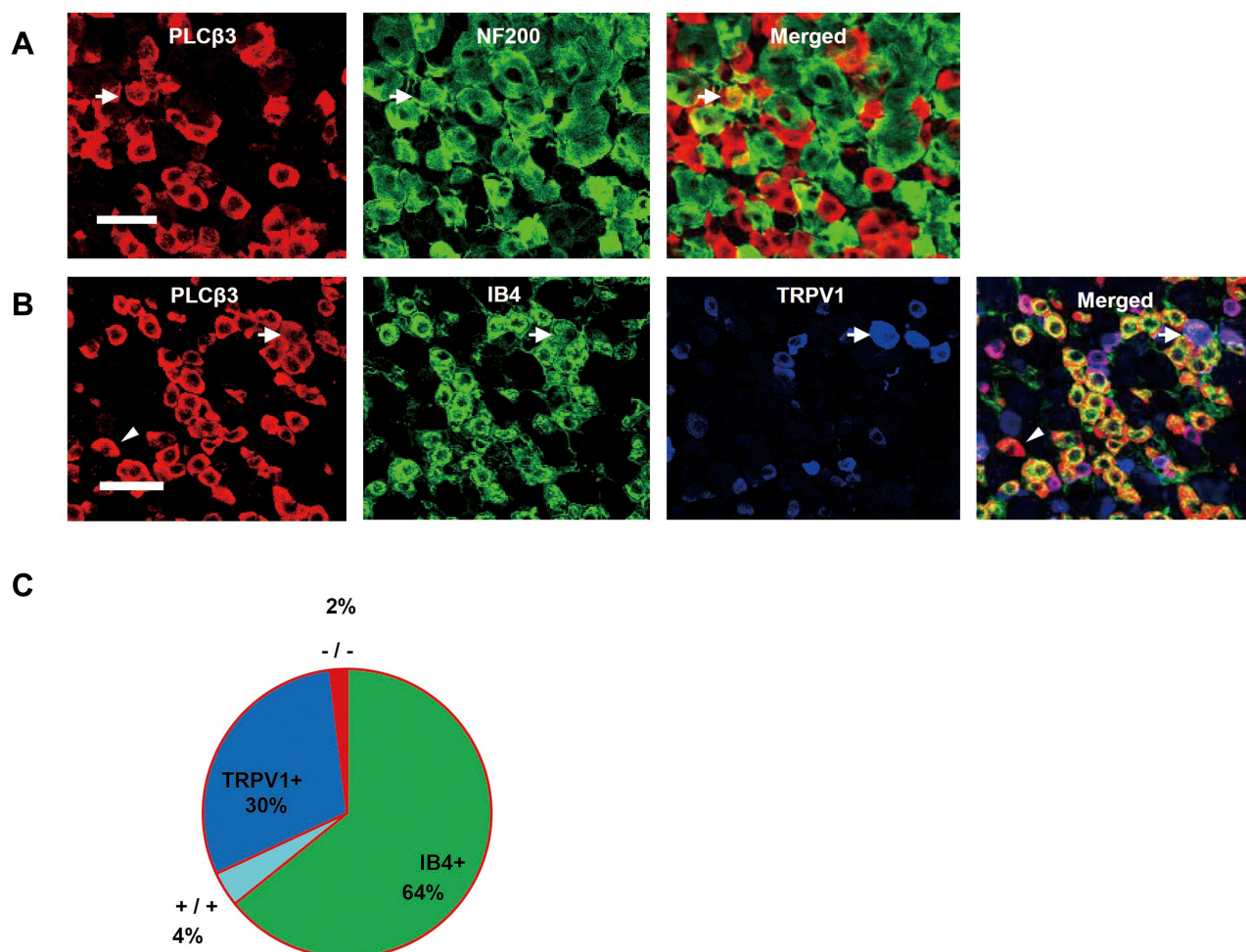


Figure 1 Expression of PLCβ3 in DRG neurons of naïve mice. **(A)** Co-localization of PLCβ3 (red) with NF200 (green). PLCβ3 immunoreactivity was found in small- to medium-sized DRG neurons. Most of the PLCβ3-positive profiles were NF200-negative. An arrow indicates both a PLCβ3-positive and an NF200-positive neuron. Scale bar = 50 μm. **(B)** Co-localization of PLCβ3 (red) with IB4 (green) and TRPV1 (blue). Many of the PLCβ3 immunoreactivity profiles were IB4-positive and/or TRPV1-positive profiles. 52% of TRPV1-expressing DRG neurons were PLCβ3-positive. An arrow indicates PLCβ3 immunoreactivity found in an IB4-positive/TRPV1-positive profile. An arrowhead indicates PLCβ3 immunoreactivity found in an IB4-negative/TRPV1-negative profile. Scale bar = 50 μm. **(C)** A pie chart summarizing the co-localization of PLCβ3 with IB4 and TRPV1. The chart shows that 64%, 30% and 4% of the PLCβ3-expressing DRG neuron profiles were IB4-positive, TRPV1-positive, and both IB4- and TRPV1-positive, respectively. Data were calculated using combined data from 4 mice (1571 PLCβ3-positive profiles from 4 mice).

Abbreviations: PLCβ3, phospholipase Cβ3; DRG, dorsal root ganglion; NF200, neurofilament 200 kD; IB4, isolectin B4; TRPV1, transient receptor potential vanilloid subfamily 1.

PLCβ1/β-actin and that of PLCβ4/β-actin compared to levels in naïve mice (Figure 3B and C). In addition, PLCβ3 antisense ODN had no effect on either the relative intensity of PLCβ3/β-actin in keratinocytes or that in the cerebellum (data not shown).

Effects of the PLCβ3 Antisense ODN on CFA-Induced Hypersensitivity to Thermal and Mechanical Stimuli at the Early Phase

To examine the involvement of PLCβ3 in the early phase of CFA-induced inflammatory pain, the PLCβ3 antisense ODN or mismatch ODN was intrathecally administered once a day for 3 consecutive days before CFA injection. Intrathecal

administration of the PLCβ3 antisense ODN significantly decreased the relative intensity of PLCβ3/β-actin at 24 h after CFA injection compared to that with intrathecal administration of the PLCβ3 mismatch ODN (0.56 ± 0.10 and 0.97 ± 0.14 , respectively; $P < 0.05$) (Figure 4A). In behavioral analyses, intrathecally administered PLCβ3 antisense ODN or mismatch ODN did not affect the basal PWL and PWT (Figure 4B and C). Mice treated with the PLCβ3 mismatch ODN (mismatch-treated mice) showed significant decreases in PWLs and PWTs from 4 h to 3 days after CFA injection compared to those before CFA injection ($**P < 0.01$). Mice treated with the PLCβ3 antisense ODN (antisense-treated mice) showed significantly higher PWTs and

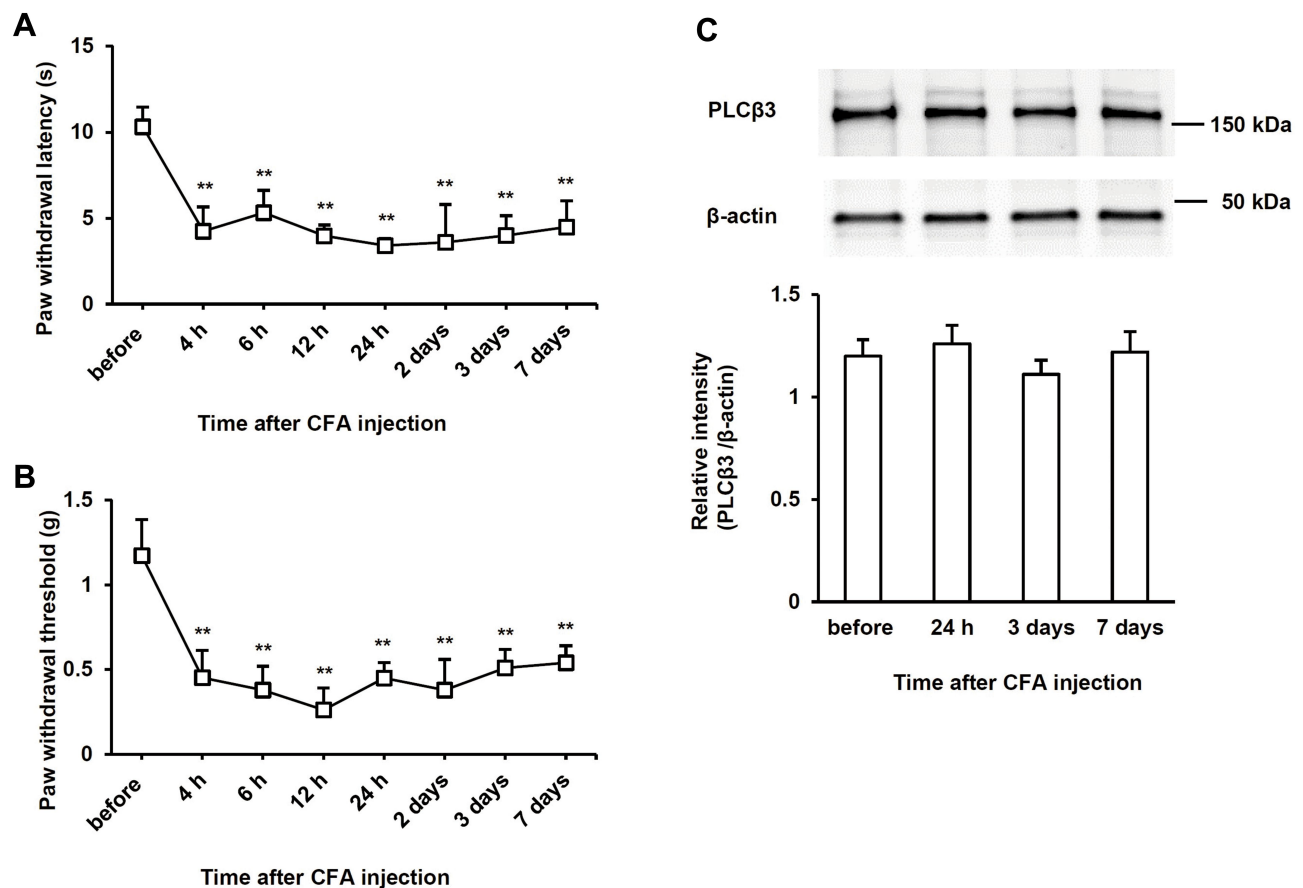


Figure 2 Time courses of hyperalgesia and PLCβ3 expression after induction of CFA-induced inflammation. **(A)** Time courses of thermal hyperalgesia after CFA injection ($n = 6$ in each group) in naïve mice. Values are expressed as means \pm SDs. $**P < 0.01$ compared to basal values (before CFA injection). **(B)** Time courses of mechanical hyperalgesia after CFA injection ($n = 6$ in each group) in naïve mice. Values are expressed as means \pm SDs. $**P < 0.01$ compared to basal values (before CFA injection). **(C)** Time courses of PLCβ3 expression in L4/5/6 DRGs after CFA injection in naïve mice. The upper panel shows representative bands of PLCβ3 and β-actin. The lower panel shows the means of relative intensity for PLCβ3/β-actin. $n = 4$ (8 mice) in each group.

Abbreviations: PLCβ3, phospholipase Cβ3; CFA, complete Freund's adjuvant; SD, standard deviation; DRG, dorsal root ganglion.

PWLs from 4 h to 24 h after CFA injection ($###P < 0.01$; $\#P < 0.05$) compared to those in the mismatch-treated mice. At 2 days and 3 days after CFA injection, PWTs and PWLs in the antisense-treated mice were comparable to those in the mismatch-treated mice. On the other hand, the antisense-treated mice showed significant decreases in PWTs from 4 to 24 h ($***P < 0.01$) but not in PWLs after CFA injection compared to those before CFA injection.

Effects of the PLCβ3 Antisense ODN on CFA-Induced Hypersensitivity to Thermal and Mechanical Stimuli at the Late Phase

To examine the involvement of PLCβ3 in the late phase of CFA-induced hypersensitivity, the PLCβ3 antisense ODN or mismatch ODN was intrathecally administered for 3 consecutive days (days 4, 5 and 6 after CFA injection) after CFA-induced hypersensitivity had been established.

Intrathecal administration of the PLCβ3 antisense ODN significantly decreased the relative intensity of PLCβ3/β-actin at 7 days after CFA injection compared to that with intrathecal administration of the PLCβ3 mismatch ODN (0.55 ± 0.09 and 0.97 ± 0.10 , respectively; $P < 0.05$) (Figure 5A). The PLCβ3 antisense ODN, but not the mismatch ODN, significantly reversed the decreased PWLs and PWTs at 7 days after CFA injection (Figure 5B and C). At 8 days after CFA injection, PWLs and PWTs in the antisense-treated mice were comparable to those in the mismatch-treated mice.

Effects of the PLCβ3 Antisense ODN on Surgical Incision-Induced Hypersensitivity to Thermal and Mechanical Stimuli

After a plantar incision, ipsilateral PWL in response to noxious heat stimuli and ipsilateral PWT in response to

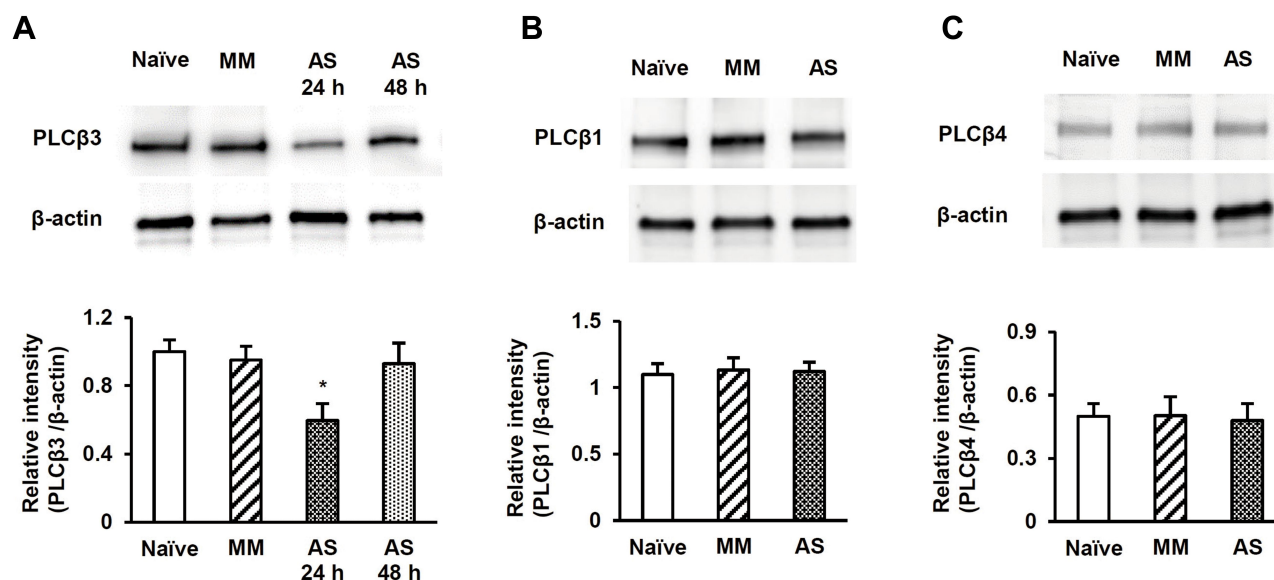


Figure 3 Effects of intrathecal PLCβ3 AS ODN and MM ODN on PLCβ1, PLCβ3, and PLCβ4 expression. **(A)** Effect of PLCβ3 AS or MM ODN on PLCβ3 expression. Intrathecal injections of PLCβ3 AS ODN, but not PLCβ3 MM ODN, significantly decreased PLCβ3 protein in L4/5/6 DRGs compared to that in naïve mice at 24 h after the last administration of the ODN. The relative intensity 48 h after the last administration of the AS ODN was comparable to that in naïve mice. The upper panel shows representative bands of PLCβ3 and β-actin. The lower panel shows the means of relative intensity for PLCβ3/β-actin. $n = 4$ (8 mice) in each group. * $P < 0.05$ compared to naïve mice. **(B)** Effect of PLCβ3 AS or MM ODN on PLCβ1 expression. Intrathecal injections of PLCβ3 AS ODN did not affect PLCβ1 protein expression in L4/5/6 DRGs compared to that in naïve mice. The upper panel shows representative bands of PLCβ1 and β-actin. The lower panel shows the means of relative intensity for PLCβ1/β-actin. $n = 4$ (8 mice) in each group. **(C)** Effect of PLCβ3 AS or MM ODN on PLCβ4 expression. Intrathecal injections of PLCβ3 AS ODN did not affect PLCβ4 protein expression in L4/5/6 DRGs compared to that in naïve mice. The upper panel shows representative bands of PLCβ4 and β-actin. The lower panel shows the means of relative intensity for PLCβ4/β-actin. $n = 4$ (8 mice) in each group.

Abbreviations: PLCβ3, phospholipase Cβ3; AS, antisense; ODN, oligodeoxynucleotide; MM, mismatch; PLCβ1, phospholipase Cβ1; PLCβ4, phospholipase Cβ4; DRG, dorsal root ganglion.

mechanical stimuli by von Frey filaments were significantly decreased compared to basal values (before injection) ($P < 0.01$) (Figure 6A and B). Significant decreases in PWL and PWT were evident 2 h after plantar incision and lasted up to 3 days and 2 days, respectively. To examine the involvement of PLCβ3 in postoperative pain, the PLCβ3 antisense ODN or mismatch ODN was intrathecally administered once a day for 3 consecutive days before plantar incision. Mismatch-treated mice showed significant decreases in PWLs and PWTs after plantar incision compared to those before plantar incision, and these significant decreases were comparable to those in naïve mice. Antisense-treated mice showed significantly higher PWLs and PWTs from 2 h to 24 h after plantar incision than those in the mismatch-treated mice ($P < 0.05$). Because PLCβ3 was not upregulated even in a long-lasting pathological state by CFA injection, we did not investigate incision-induced changes of PLCβ3 expression by Western blot analysis.

Discussion

The main findings of our study are as follows. First, most of the PLCβ3 was expressed in TRPV1-positive and/or IB4-binding primary afferents, which are considered nociceptive

(Figure 1). Second, the amount of PLCβ3 expressed in lumbar DRGs was not changed after peripheral inflammation induced by CFA injection into the hindpaw (Figure 2). Third, knockdown of PLCβ3 in lumbar DRGs before CFA injection, which did not affect basal PWL in response to thermal stimuli and PWT in response to mechanical stimuli, reversed the thermal hyperalgesia and mechanical hyperalgesia seen at 1 day (early phase) and 7 days (late phase) after CFA injection (Figures 4 and 5). The knockdown effect observed in Western blot analysis lasted until 24 h after the last administration of the antisense ODN but was wearing off at 48 h (Figure 3), consistent with the time course of behavioral analysis in CFA-induced inflammatory hyperalgesia. Depression of PLCβ3 suppressed thermal and mechanical hyperalgesia in the postoperative pain model (Figure 6). Taken together, these results suggest that PLCβ3 in DRG neurons is involved in the transmission of inflammatory and postoperative pain.

Expression of PLCβ3 in DRG Neurons

We investigated the distribution of PLCβ3-expressed DRG neurons for clarifying the mechanisms of inflammation-induced hyperalgesia via the PLCβ3 signal pathway.

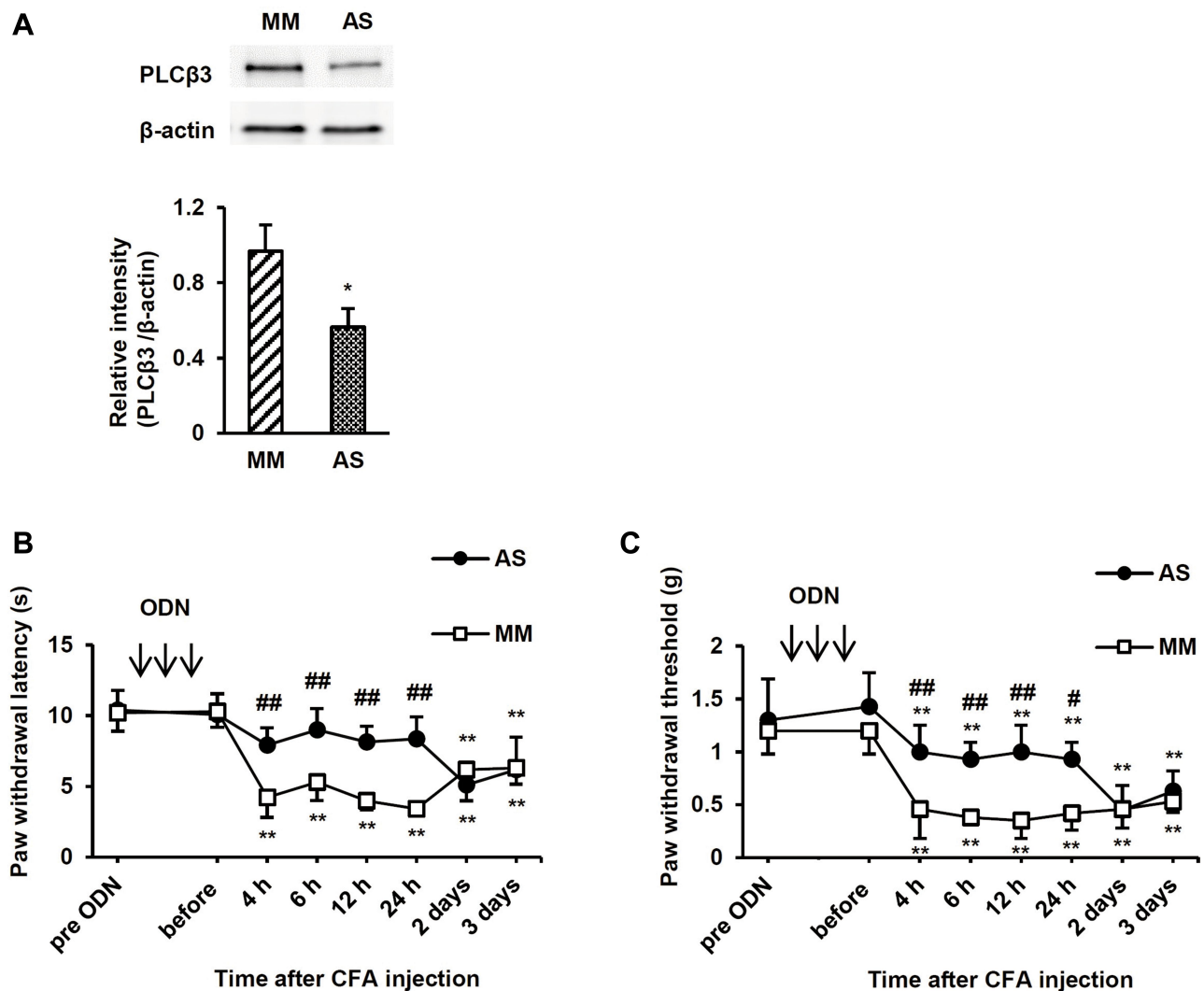


Figure 4 Effects of intrathecal PLCβ3 ODNs on the early phase of CFA-induced thermal and mechanical hyperalgesia. **(A)** Immunoblot analysis of PLCβ3 in L4/5/6 DRGs. Intrathecal injections of PLCβ3 AS ODN significantly decreased PLCβ3 expression at 24 h after CFA injection compared to that with intrathecal injection of PLCβ3 MM ODN. The upper panel shows representative bands of PLCβ3 and β-actin. The lower panel shows the means of relative intensity for PLCβ3/β-actin. $n = 4$ (8 mice) in each group. $*P < 0.05$ compared to MM-treated mice. **(B)** Effects of intrathecal injections of PLCβ3 AS ODN on CFA-induced thermal hyperalgesia. Intrathecal injections of PLCβ3 AS ODN, but not MM ODN, significantly inhibited the development of CFA-induced thermal hyperalgesia until 24 h after CFA injection. Values are expressed as means \pm SDs ($n = 6$ in each group). $**P < 0.01$ compared to the baseline value (before CFA injection). $##P < 0.01$ compared to PLCβ3 MM ODN. Arrows indicate intrathecal administration of PLCβ3 AS or MM ODN once a day for 3 consecutive days. **(C)** Effects of intrathecal injections of PLCβ3 AS ODN on CFA-induced mechanical hyperalgesia. Mechanical hyperalgesia developed in mice treated with intrathecal injections of PLCβ3 AS ODN and MM ODN. However, paw withdrawal thresholds in mice treated with intrathecal PLCβ3 AS ODN were significantly higher than those in mice treated with PLCβ3 MM ODN from 4 h to 24 h after CFA injection. Values are expressed as means \pm SDs ($n = 6$ in each group). $**P < 0.01$ compared to the baseline value (before CFA injection). $*P < 0.05$; $##P < 0.01$ compared to PLCβ3 MM ODN. Arrows indicate intrathecal administration of PLCβ3 AS or MM ODN once a day for 3 consecutive days.

Abbreviations: PLCβ3, phospholipase Cβ3; ODN, oligodeoxynucleotide; CFA, complete Freund's adjuvant; DRG, dorsal root ganglion; AS, antisense; MM, mismatch; SD, standard deviation.

Previous studies showed that PLCβ3 is expressed in DRG neurons, keratinocytes and cerebellar Purkinje cells^{3,6,21} but not in spinal neurons.³ Previous studies have suggested that, among the small- to medium-sized DRG neurons, PLCβ3 is mainly expressed in IB4-binding DRG neurons, which are unmyelinated non-peptidergic cells.^{3,21} However, more detailed information about the expression profile of PLCβ3 in DRG neurons was not obtained in those previous studies. In this study, we found that approximately half of TRPV1-

positive DRG neurons expressed PLCβ3. We also found that 64%, 30% and 4% of the PLCβ3-expressing DRG neurons were IB4-positive, TRPV1-positive, and both IB4- and TRPV1-positive, respectively (Figure 1B and C).

Since TRPV1-positive and IB4-positive DRG neurons are considered to be different types of nociceptors,^{22,23} the expression pattern of PLCβ3 in DRG neurons suggests that PLCβ3 particularly modulates the activity of nociceptors contributing to pain perception. A recent

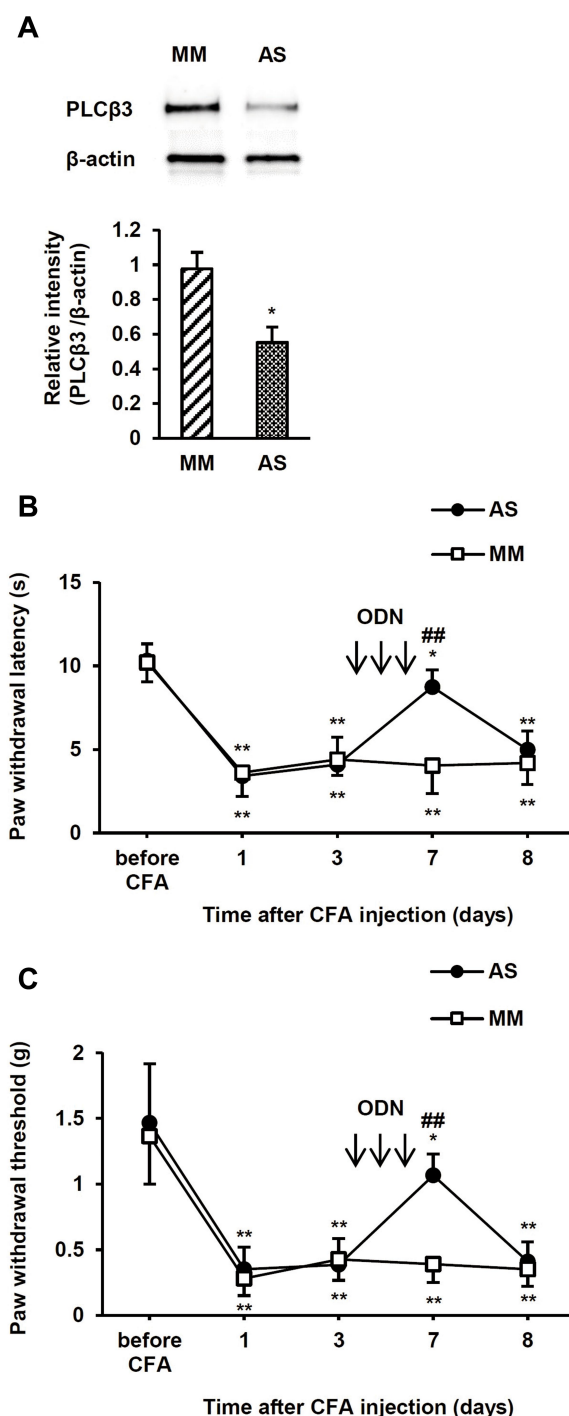


Figure 5 Effects of intrathecal PLCβ3 ODNs on the late phase of CFA-induced thermal and mechanical hyperalgesia. **(A)** Immunoblot analysis of PLCβ3 in L4/5/6 DRGs. Intrathecal injections of PLCβ3 AS ODN significantly decreased PLCβ3 expression in L4/5/6 DRGs at 7 days after CFA injection compared to that with intrathecal injection of PLCβ3 MM ODN. The upper panel shows representative bands of PLCβ3 and β-actin. The lower panel shows the means of relative intensity for PLCβ3/β-actin. $n = 4$ (8 mice) in each group. * $P < 0.05$ compared to MM-treated mice. **(B)** Effects of intrathecal injections of PLCβ3 AS ODN on CFA-induced thermal hyperalgesia. Intrathecal injections of PLCβ3 AS ODN, but not MM ODN, significantly reversed CFA-induced thermal hyperalgesia. Values are expressed as means \pm SDs ($n = 6$ in each group). * $P < 0.05$; ** $P < 0.01$ compared to the baseline value (before CFA injection). ### $P < 0.01$ compared to PLCβ3 MM ODN. Arrows indicate intrathecal administration of PLCβ3 AS ODN or MM ODN. The ODNs were administered for 3 consecutive days (4, 5 and 6 days after CFA injection). **(C)** Effects of intrathecal injections of PLCβ3 AS ODN on CFA-induced mechanical hyperalgesia. Intrathecal injections of PLCβ3 AS ODN, but not MM ODN, significantly reversed CFA-induced mechanical hyperalgesia. Values are expressed as means \pm SDs ($n = 6$ in each group). * $P < 0.05$; ** $P < 0.01$ compared to the baseline value (before CFA injection). ### $P < 0.01$ compared to PLCβ3 MM ODN. Arrows indicate intrathecal administration of PLCβ3 AS ODN or MM ODN. ODNs were administered for 3 consecutive days (4, 5 and 6 days after CFA injection).

Abbreviations: PLCβ3, phospholipase Cβ3; ODN, oligodeoxynucleotide; CFA, complete Freund's adjuvant; DRG, dorsal root ganglion; AS, antisense; MM, mismatch; SD, standard deviation.

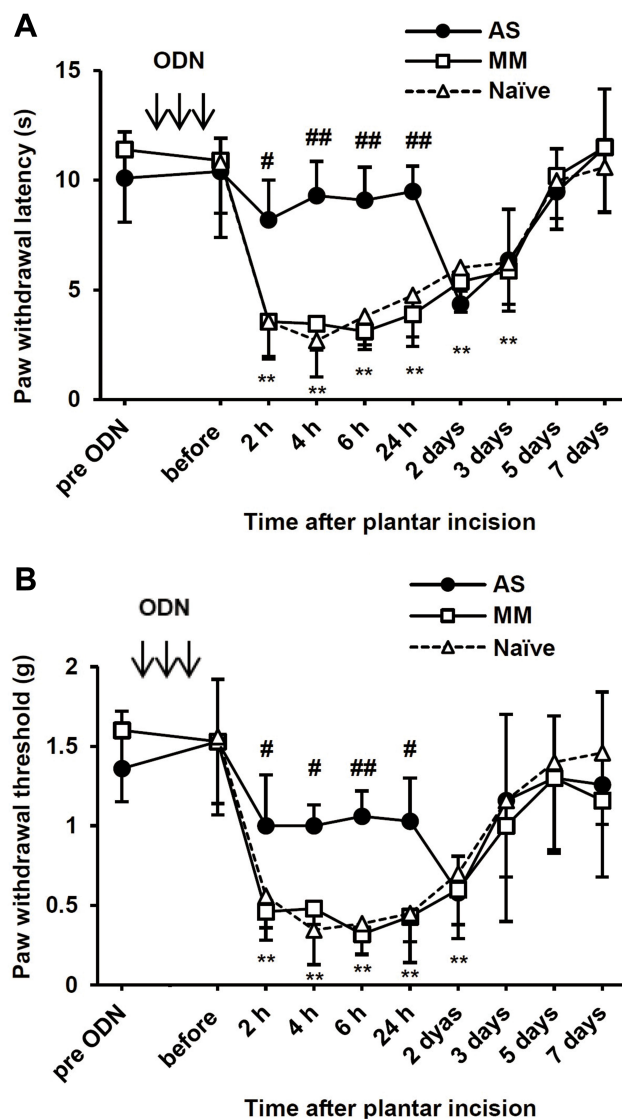


Figure 6 Time courses of thermal and mechanical hyperalgesia after plantar incision and effects of intrathecal PLC β 3 ODNs. **(A)** Time courses of thermal hyperalgesia after plantar incision in naïve mice and effects of intrathecal PLC β 3 ODNs on thermal hyperalgesia after plantar incision. After plantar incision, ipsilateral PWLs in response to noxious heat stimuli were significantly decreased compared to the basal values (before incision) at 2 h after incision and lasted up to 3 days in naïve mice and MM ODN-treated mice. Intrathecal injections of PLC β 3 AS ODN significantly inhibited thermal hyperalgesia until 24 h after plantar incision. Values are expressed as means \pm SDs ($n = 6$ in each group). ** $P < 0.01$ compared to the baseline value (before plantar incision) in naïve mice. # $P < 0.05$; ### $P < 0.01$ compared to PLC β 3 MM ODN. Arrows indicate intrathecal administration of PLC β 3 AS or MM ODN once a day for 3 consecutive days. **(B)** Time courses of mechanical hyperalgesia after plantar incision in naïve mice and effects of intrathecal PLC β 3 ODNs on mechanical hyperalgesia after plantar incision. After plantar incision, ipsilateral PWTs in response to stimuli by von Frey filaments were significantly decreased compared to the basal values (before incision) at 2 h after incision and lasted up to 2 days in naïve mice. Intrathecal injections of PLC β 3 AS ODN, but not MM ODN, significantly inhibited mechanical hyperalgesia until 24 h after plantar incision. Values are expressed as means \pm SDs ($n = 6$ in each group). ** $P < 0.01$ compared to the baseline value (before plantar incision) in naïve mice. # $P < 0.05$; ### $P < 0.01$ compared to PLC β 3 MM ODN. Arrows indicate intrathecal administration of PLC β 3 AS or MM ODN once a day for 3 consecutive days.

Abbreviations: PLC β 3, phospholipase C β 3; ODN, oligodeoxynucleotide; PWL, paw withdrawal latency; MM, mismatch; AS, antisense; SD, standard deviation; PWT, paw withdrawal threshold.

study showed that inflammation-induced mechanical hyperalgesia and thermal hyperalgesia are selectively mediated by different types of DRG neurons.²⁴ MrgprD-positive DRG neurons, which are thought to be mostly IB4-positive DRG neurons, and calcitonin gene-related peptide-positive DRG neurons, which are thought to be mostly TRPV1-positive DRG neurons, are selectively involved in inflammation-induced mechanical hyperalgesia and thermal hyperalgesia, respectively.²⁴ Therefore, knockdown of PLC β 3 expressed in IB4-positive and TRPV1-positive DRG neurons might alleviate both mechanical hyperalgesia and thermal hyperalgesia during inflammation.

Roles of PLC β 3 in Inflammatory Pain

In the present study, intrathecal administration of the PLC β 3 antisense ODN selectively inhibited PLC β 3 expression in DRG neurons but not PLC β 1 or PLC β 4 expression (Figure 3). Intrathecally administered substances probably spread to not only DRGs but also the spinal cord. Since PLC β 3 is not expressed in spinal neurons,³ alleviation of thermal and mechanical hyperalgesia after intrathecal administration of the PLC β 3 antisense ODN is likely to be due to inhibition of PLC β 3 expression in DRG neurons.

Intrathecal administration of the PLC β 3 antisense ODN for three days significantly decreased the relative intensity of PLC β 3/ β -actin to approximately 60% of that in naïve mice but did not affect basal PWLs and PWTs under non-inflammation conditions (Figure 4). These results suggested that PLC β 3 is not involved in nociception without inflammation or tissue damage. In contrast, intrathecal administration of the PLC β 3 antisense ODN for three days significantly reversed PWLs and PWTs in pathophysiological nociceptive pain states. Expression of PLC β 3 in DRGs was not up-regulated at the early and late phases of CFA-induced inflammation compared to the baseline level under normal conditions (Figure 2C). The level of activation of PLC β 3 was not determined in the current study because activation-specific antibodies were not used. PLC β 3 may be activated in a pathological state such as inflammation. It is possible that inflammation-induced activation of PLC β 3 in DRG neurons is required to augment transmission of nociceptive signals from peripheral nerve endings.

At present, it is not clear how PLC β 3 in DRGs mediates signal transmission of inflammatory pain. One possible

explanation is that continuous activation of PLC β 3 under the condition of inflammation promotes signal transmission of pain through PLC β 3-expressing primary afferents, resulting in hyperalgesia. Our results are consistent with the results of a previous study showing that intrathecal administration of PLC β 3 antisense ODN decreased mechanical hyperalgesia induced by injection of carrageenan into the hindpaw.⁹ These results support the notion that PLC β 3 has a role in signal transmission under the condition of inflammation. However, the results of our behavioral analysis are not consistent with the results of another previous study showing that PWT to thermal stimuli and PWT to mechanical stimuli after CFA injection in PLC β 3 whole-body knockout mice were similar to those in wild-type mice.⁸ Functional compensation might occur in such PLC β 3-deficient mice. On the other hand, since PLC β 3 is exclusively expressed in nociceptors,^{9,21,24} it could be expected that ablation of PLC β 3 would result in a decrease in basal pain sensitivity. However, this was not the case in our study. Although the reason for this is not clear, the results of our study suggest that targeting PLC β 3 does not affect such basal pain sensitivities but only reverses mechanical and thermal hyperalgesia in inflammatory pain states. In addition, since PLC β 3 is mostly expressed in nociceptors, targeting PLC β 3 may reduce pain perception in inflammatory pain states without affecting other sensory modalities such as touch, pressure and vibration.

Roles of PLC β 3 in Postoperative Pain

We first demonstrated that intrathecal administration of the PLC β 3 antisense ODN alleviated postoperative thermal and mechanical hyperalgesia, indicating that PLC β 3 in DRG neurons is also involved in postsurgical hyperalgesia. We performed behavioral studies at the early phase but not at the late phase because the duration of postoperative hyperalgesia is relatively short compared to that caused by CFA. Postoperative pain is distinct from pure inflammatory pain or neuropathic models of hyperalgesia.¹⁰ This is partly because TRPV1 protein upregulation occurs after inflammation but apparently not after incision, though TRPV1 mediates heat hyperalgesia after both incision and inflammation.¹² Our results suggest that PLC β 3 has an important role in the signal transmission of both CFA-induced inflammatory pain and incisional pain. On the other hand, it has been reported that TRPV1 was functionally up-regulated specifically in isolated IB4-positive DRG sensory neurons following skin incision injury and that TRPV1 was involved in heat hyperalgesia.²⁵ Therefore, it

is possible that the PLC β 3 antisense ODN alleviated incisional thermal hyperalgesia through downregulation of PLC β 3 in TRPV1-positive DRG neurons. The mechanism of postsurgical mechanical hyperalgesia has not been clearly elucidated. Further study is needed to investigate the role of PLC β 3 in postoperative hypersensitivity.

Clinical Implication

The mechanisms of early phase of hyperalgesia, that is, development of hyperalgesia induced by inflammation are partially different from those of late phase hyperalgesia, that is, maintenance of hyperalgesia. For example, the interleukin-1 β and tumor necrosis factor- α /nuclear factor-kappa B/cyclooxygenase-2 pathways are involved in the development of inflammation-induced hyperalgesia but not the maintenance of the hyperalgesia.²⁶ Acid-sensitive ion channel 3 is involved in the development but not in the maintenance of mechanical hyperalgesia in CFA-induced inflammatory pain.²⁷ Our results showed that the PLC β 3 antisense ODN had anti-hyperalgesic effects both at the early and late phases of CFA-induced inflammatory pain. In addition, the PLC β 3 antisense ODN inhibited thermal and mechanical hyperalgesia in the postoperative pain model. In addition to inflammatory pain as a type of tissue injury-induced pain such as osteoarthritis, postoperative pain is also another type of tissue injury-induced pain that is commonly encountered in a clinical setting. Nociceptor-specific deletion of Gq/11, which is upstream of PLC β 3, and systemic administration of a nonselective PLC inhibitor reduce CFA-induced inflammatory pain and neuropathic pain,²⁸ suggesting that PLC β 3 may play an important role in various stages of inflammatory pain including postoperative pain. Targeting PLC β 3 may thus lead to a new insight into drug discovery of analgesics for relieving various types of pathophysiological pain states.

Conclusion

Our results indicate that PLC β 3 in DRG neurons is involved in the development and maintenance of inflammatory pain and postoperative pain, although PLC β 3 does not play a major role in thermal sensitivity and tactile sensory threshold in normal conditions without inflammation. PLC β 3 may be an attractive target for the treatment of thermal and mechanical hyperalgesia in inflammatory pain and postoperative pain.

Funding

This study was supported by our institutional grant and Grants-in-Aid for Scientific Research (23390375 to

Tomoyuki Kawamata and 25861369 to Susumu Ide) from the Japan Society for Promotion of Science (Tokyo, Japan).

Disclosure

The authors report no conflicts of interest in this work.

References

- Hucho T, Levine JD. Signaling pathways in sensitization: toward a nociceptor cell biology. *Neuron*. 2007;55:365–376. doi:10.1016/j.neuron.2007.07.008
- Rebecchi MJ, Pentyala SN. Structure, function, and control of phosphoinositide-specific phospholipase C. *Physiol Rev*. 2000;80:1291–1335.
- Han SK, Mancino V, Simon MI. Phospholipase C β 3 mediates the scratching response activated by the histamine H1 receptor on C-fiber nociceptive neurons. *Neuron*. 2006;52:691–703. doi:10.1016/j.neuron.2006.09.036
- Haase I, Liesegang C, Binting S, Henz BM, Rosenbach T. Phospholipase C-mediated signaling is altered during HaCaT cell proliferation and differentiation. *J Invest Dermatol*. 1997;108:748–752. doi:10.1111/1523-1747.ep12292135
- Ando T, Xiao W, Gao P, et al. Critical role for mast cell stat5 activity in skin inflammation. *Cell Rep*. 2014;6:366–376. doi:10.1016/j.celrep.2013.12.029
- Watanabe M, Nakamura M, Sato K, Kano M, Simon MI, Inoue Y. Patterns of expression for the mRNA corresponding to the four isoforms of phospholipase C β in mouse brain. *Eur J Neurosci*. 1998;10:2016–2025. doi:10.1046/j.1460-9568.1998.00213.x
- Nomura S, Fukaya M, Tsujioka T, Wu D, Watanabe M. Phospholipase C β 3 is distributed in both somatodendritic and axonal compartments and localized around perisynapse and smooth endoplasmic reticulum in mouse Purkinje cell subsets. *Eur J Neurosci*. 2007;25:659–672.
- Imamachi N, Park GH, Lee H, et al. TRPV1-expressing primary afferents generate behavioral responses to pruritogens via multiple mechanisms. *Proc Natl Acad Sci U S A*. 2009;106:11330–11335. doi:10.1073/pnas.0905605106
- Joseph EK, Bogen O, Alessandri-Haber N, Levine JD. PLC- β 3 signals upstream of PKC ϵ in acute and chronic inflammatory hyperalgesia. *Pain*. 2007;132:67–73. doi:10.1016/j.pain.2007.01.027
- Brennan TJ, Zahn PK, Pogatzki-Zahn EM. Mechanisms of incisional pain. *Anesthesiol Clin North America*. 2005;23:1–20. doi:10.1016/j.atc.2004.11.009
- Ishida K, Kawamata T, Tanaka S, Shindo T, Kawamata M. Calcitonin gene-related peptide is involved in inflammatory pain but not in postoperative pain. *Anesthesiology*. 2014;121:1068–1079. doi:10.1097/ALN.0000000000000364
- Pogatzki-Zahn EM, Shimizu I, Caterina M, Raja SN. Heat hyperalgesia after incision requires TRPV1 and is distinct from pure inflammatory pain. *Pain*. 2005;115:296–307. doi:10.1016/j.pain.2005.03.010
- Banik RK, Brennan TJ. Trpv1 mediates spontaneous firing and heat sensitization of cutaneous primary afferents after plantar incision. *Pain*. 2009;141:41–51. doi:10.1016/j.pain.2008.10.004
- Kaneda Y, Nakajima T, Nishikawa T, et al. Hemagglutinating virus of Japan (HVJ) envelope vector as a versatile gene delivery system. *Mol Ther*. 2002;6:219–226. doi:10.1006/mthe.2002.0647
- Furuse S, Kawamata T, Yamamoto J, et al. Reduction of bone cancer pain by activation of spinal cannabinoid receptor 1 and its expression in the superficial dorsal horn of the spinal cord in a murine model of bone cancer pain. *Anesthesiology*. 2009;111:173–186. doi:10.1097/ALN.0b013e3181a51e0d
- Kawamata T, Ji W, Yamamoto J, Niiyama Y, Furuse S, Namiki A. Contribution of transient receptor potential vanilloid subfamily 1 to endothelin-1-induced thermal hyperalgesia. *Neuroscience*. 2008;154:1067–1076. doi:10.1016/j.neuroscience.2008.04.010
- Chaplan SR, Bach FW, Pogrel JW, Chung JM, Yaksh TL. Quantitative assessment of tactile allodynia in the rat paw. *J Neurosci Methods*. 1994;53:55–63. doi:10.1016/0165-0270(94)90144-9
- Baiou D, Santha P, Avelino A, et al. Neurochemical characterization of insulin receptor-expressing primary sensory neurons in wild-type and vanilloid type 1 transient receptor potential receptor knockout mice. *J Comp Neurol*. 2007;503:334–347. doi:10.1002/cne.21389
- Fukaya M, Uchigashima M, Nomura S, Hasegawa Y, Kikuchi H, Watanabe M. Predominant expression of phospholipase C β 1 in telencephalic principal neurons and cerebellar interneurons, and its close association with related signaling molecules in somatodendritic neuronal elements. *Eur J Neurosci*. 2008;28:1744–1759. doi:10.1111/j.1460-9568.2008.06495.x
- Nakamura M, Sato K, Fukaya M, et al. Signaling complex formation of phospholipase C β 4 with metabotropic glutamate receptor type 1 α and 1,4,5-trisphosphate receptor at the perisynapse and endoplasmic reticulum in the mouse brain. *Eur J Neurosci*. 2004;20:2929–2944.
- Shi TJ, Liu SX, Hammarberg H, Watanabe M, Xu ZQ, Hokfelt T. Phospholipase C β 3 in mouse and human dorsal root ganglia and spinal cord is a possible target for treatment of neuropathic pain. *Proc Natl Acad Sci U S A*. 2008;105:20004–20008. doi:10.1073/pnas.0810899105
- Caterina MJ, Schumacher MA, Tominaga M, Rosen TA, Levine JD, Julius D. The capsaicin receptor: a heat-activated ion channel in the pain pathway. *Nature*. 1997;389:816–824.
- Vulchanova L, Olson TH, Stone LS, Riedl MS, Elde R, Honda CN. Cytotoxic targeting of isolectin IB4-binding sensory neurons. *Neuroscience*. 2001;108:143–155. doi:10.1016/S0306-4522(01)00377-3
- Cavanaugh DJ, Lee H, Lo L, et al. Distinct subsets of unmyelinated primary sensory fibers mediate behavioral responses to noxious thermal and mechanical stimuli. *Proc Natl Acad Sci U S A*. 2009;106:9075–9080. doi:10.1073/pnas.0901507106
- Barabas ME, Stucky CL. TRPV1, but not TRPA1, in primary sensory neurons contributes to cutaneous incision-mediated hypersensitivity. *Mol Pain*. 2013;9:9. doi:10.1186/1744-8069-9-9
- Narita M, Shimamura M, Imai S, et al. Role of interleukin-1 β and tumor necrosis factor- α -dependent expression of cyclooxygenase-2 mRNA in thermal hyperalgesia induced by chronic inflammation in mice. *Neuroscience*. 2008;152:477–486. doi:10.1016/j.neuroscience.2007.10.039
- Karczewski J, Spencer RH, Garsky VM, et al. Reversal of acid-induced and inflammatory pain by the selective ASIC3 inhibitor, APETx2. *Br J Pharmacol*. 2010;161:950–960. doi:10.1111/j.1476-5381.2010.00918.x
- Tappe-Theodor A, Constantin CE, Tegeder I, et al. Ga(q/11) signaling tonically modulates nociceptor function and contributes to activity-dependent sensitization. *Pain*. 2012;153:184–196. doi:10.1016/j.pain.2011.10.014

Journal of Pain Research**Dovepress****Publish your work in this journal**

The Journal of Pain Research is an international, peer reviewed, open access, online journal that welcomes laboratory and clinical findings in the fields of pain research and the prevention and management of pain. Original research, reviews, symposium reports, hypothesis formation and commentaries are all considered for publication. The manuscript

management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/journal-of-pain-research-journal>