## 信州医学雑誌 第67卷 第3号 掲載予定

- 1 (和文題名) ブレオマイシン誘発肺線維症ラットモデルに対するサブタイプ選択
- 2 的 E プロスタノイド受容体アゴニストの効果
- 3 (著者名)市山 崇史<sup>1)</sup>, 山本 洋<sup>1)</sup>, 本郷 一博<sup>2)</sup>
- 4 (所属名)
- 5 1) 信州大学医学部内科学第一教室,
- 6 2) 信州大学医学部創薬科学講座
- 7 (欧文題名) Effects of Subtype-selective E Prostanoid Receptor Agonists on
- 8 Bleomycin-induced Pulmonary Fibrosis in Rats
- 9 (著者名ローマ字)Takashi Ichiyama<sup>1)</sup>, Hiroshi Yamamoto<sup>1)</sup>, Kazuhiro Hongo<sup>2)</sup>,
- 10 (所属欧文名)
- 11 1) First Department of Internal Medicine, Shinshu University School of
- 12 Medicine, 3-1-1 Asahi
- 13 2) Department of Drug Discovery Science, Shinshu University School of
- 14 Medicine
- 15 Key words
- 16 pulmonary fibrosis, E prostanoid receptor agonists,
- 17 cyclic adenosine monophosphate, transforming growth factor- $\beta$ 1,
- 18 bleomycin-induced pulmonary fibrosis

- RUNNING TITLE: Effect of selective EP agonists on lung fibrosis in rats
- 本文総枚数:21頁
- 図・表の枚数:8枚
- 別刷希望部数(朱書き):20部
- Corresponding Author: Takashi Ichiyama, MD.
- First Department of Internal Medicine, Shinshu University School of Medicine,
- 3-1-1 Asahi, Matsumoto, Nagano 390-8621, Japan.
- Tel: +81-(0)263-37-2631; Fax: +81-(0)263-36-3722
- E-mail: ichiyama@shinshu-u.ac.jp

### 1 Abstract

2 Background: Idiopathic pulmonary fibrosis (IPF) is a fatal lung disease with limited treatment options and a poor prognosis. In vitro research has shown that 3 4 prostaglandin (PG) E<sub>2</sub> can suppress pulmonary fibrosis via cAMP production. 5 EP2 and EP4, which are subtypes of the receptors for PGE<sub>2</sub>, are involved in 6 cAMP production. The present study was designed to examine the effects of 7 EP2 and EP4 agonists on bleomycin (BLM)-induced pulmonary fibrosis in rats. Materials and Methods: The EP2 and EP4 agonists were subcutaneously 8 9 administered to BLM-induced pulmonary fibrosis rats for 21 days. The lung weight, mRNA expressions of transforming growth factor (TGF)-B1 and 10 11 procollagen genes, and degree of pulmonary fibrosis were compared between 12 EP2 agonist, EP4 agonist, vehicle, and pirfenidone and nintedanib administered groups. We also examined the EP2 and EP4 expressions in human lung tissues 13 14 with IPF and in rat lung tissues with BLM-induced pulmonary fibrosis by immunohistochemical staining. The human lung tissues with IPF were obtained 15 16 from autopsy cases. Results: The EP2 agonist significantly suppressed the lung 17 weight gain and inhibited the mRNA expressions of TGF-*β*1 and procollagen

1	genes. In addition, the fibrosis scores and hydroxyproline content tended to be
2	lower in the EP2 agonist administered group. However, the EP4 agonist did not
3	show such evidence in suppression of fibrosis. The enhanced expressions of
4	EP2 and EP4 were demonstrated in both human and rat lung tissues with
5	fibrosis relative to those in normal lung. Conclusions: The EP2 agonist may
6	become a novel therapeutic agent for IPF.
7	要約
8	背景:特発性肺線維症(IPF)は治療の選択肢が限られた予後不良の致命的な肺疾
9	患である. 基礎研究領域でプロスタグランジン(PG)E2が cAMP 産生を介して
10	肺線維症を抑制する事が報告されている. PGE2の受容体のサブタイプである
11	EP2およびEP4はcAMPの産生に関与している.本研究はブレオマイシン(BLM)
12	誘発肺線維症ラットモデルにおいて EP2 および EP4 アゴニストの効果を調べる
13	ために計画された. 材料および方法: EP2 および EP4 アゴニストを BLM 誘発肺
14	線維症ラットモデルに 21 日間皮下投与した. 肺の重量, TGF-β1 およびプロコラ
15	ーゲン遺伝子の mRNA の発現および肺線維症の程度を, EP2 アゴニスト, EP4 ア
16	ゴニスト, 溶媒, ピルフェニドンおよびニンテダニブを投与した群でそれぞれ
17	比較した. それに加えて免疫組織化学的染色により, BLM 誘発肺線維症ラット

1	モデルおよび IPF 患者の肺組織において EP2 および EP4 の発現を調べた. IPF
2	患者の肺組織は剖検症例から取得した. 結果:EP2 アゴニストは肺重量の増加を
3	有意に抑制し, TGF-β1 およびプロコラーゲン遺伝子の mRNA 発現を阻害した.
4	さらに, EP2 アゴニスト投与群では, 線維化スコアおよびヒドロキシプロリン含
5	量が低くなる傾向を示した. しかし, EP4 アゴニストは, 線維症の抑制を示唆す
6	る所見を示さなかった.また BLM 誘発肺線維症ラットモデルおよび IPF 患者の
7	肺組織では正常肺と比較して EP2 と EP4 の発現が強く認められた. 結論: EP2
8	アゴニストは, 新規の IPF 治療薬になり得ることが示唆された.
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# I . Introduction

2	Idiopathic pulmonary fibrosis (IPF) is a specific form of chronic, progressive,
3	fibrosing, interstitial pneumonia of unknown causes and the prognosis is very
4	poor. Currently, pirfenidone and nintedanib are clinically used in Japan for its
5	treatment. Although these drugs significantly improved the standardized mean
6	difference of change in forced vital capacity 1) 2), recent meta-analysis did not
7	show significant improvement in mortality 3). Development of new anti-fibrotic
8	drugs is expected to improve the prognosis of patients with IPF. Prostaglandin
9	(PG) E <sub>2</sub> , the major prostanoid in the lung, is an important antifibrotic lipid
10	mediator. PGE <sub>2</sub> levels were reduced in the lungs of patients with IPF 4).
11	Preventive effects of PGE2 on pulmonary fibrosis have been studied in an animal
12	model, bleomycin (BLM)-induced pulmonary fibrosis 5) 6). Administration of
13	PGE2 to BLM-induced pulmonary fibrosis in mice suppressed collagen
14	deposition and reduced mortality 5) 6). It was demonstrated that administration
15	of PGE <sub>2</sub> increased cAMP level with preventive effects of myofibroblast
16	differentiation and inhibition of extracellular membrane production of pulmonary
17	fibroblasts 7). Four E prostanoid receptor (EP) subtypes (EP1, EP2, EP3 and

1	EP4) have been identified as receptors for PGE2. EP2 and EP4 are coupled to
2	Gs protein and can increase the intracellular cAMP level 8). We have developed
3	an EP2 agonist 9) and an EP4 agonist 10) as candidates for novel antifibrotic
4	agents. In the present study, we examined the pharmacological effects of the
5	EP2 agonist and the EP4 agonist on BLM-induced pulmonary fibrosis in rats. We
6	speculate that the expressions of EP2 and EP4 in lung tissues of rats with
7	pulmonary fibrosis are fundamentaly required in the therapeutic target of the
8	EP2 agonist and EP4 agonist on IPF. The immunohistological staining was
9	performed in lung tissues of human IPF patients as well as of BLM-induced
10	pulmonary fibrosis in rats in order to confirm the expressions of EP2 and EP4 in
11	lung tissues. The present study was designed to evaluate the pharmacological
12	potential of the EP2 and EP4 agonists on human IPF.
13	II . Materials and Methods
14	A. Efficacy evaluation of EP2 and EP4 agonists in BLM-induced pulmonary
15	fibrosis rats

**1. Animals and reagents** 

1	All experimental procedures were conducted according to Shinshu University
2	Animal Experimental Rules, reviewed according to the guidelines of the National
3	Regulation and Animal Care Committee, and approved by the Shinshu
4	University Animal Research Committee (Authorization number: 270038,
5	approval date: July 1st, 2016). The 48 Wister–Hann 9-week rats were purchased
6	from Japan CRLs. We purchased BLM from Nippon Kayaku (Tokyo, Japan),
7	midazolam from Fuji Pharma Co. (Tokyo, Japan), medetomidine from
8	Kyoritsuseiyaku Co. (Tokyo, Japan) and butorphanol from Meiji Seika Pharma
9	Co. (Tokyo, Japan). We purchased pirfenidone from Tokyo Chemical Industry Co.
10	(Tokyo, Japan) and nintedanib from eNOvation Chemicals LLC (NJ, USA). The
11	EP2 agonist and EP4 agonist were synthesized by Kissei Pharmaceutical Co.
12	Ltd (Hotaka, Japan). The structural formula of the EP2 agonist is shown in
13	Fig.1a and that of the EP4 agonist in Fig.1b. The EC $_{50}$ of each compound was
14	calculated as follows. HEK293T cells forcibly expressing rat EP2 and EP4 were
15	stimulated with EP2 agonist, EP4 agonist and $PGE_2$ and analyzed according to
16	the protocol of cAMP screening system (Thermo Fisher Scientific K. K.), using
17	GraphPad Prism (GraphPad Software, California, USA) analyzed concentration

1	effect data. Pharmacokinetics, plasma protein binding of the EP2 agonist and
2	EP4 agonist in SD rats were obtained in studies conducted by Kissei
3	Pharmaceutical Co. Ltd and are shown in Fig 1c, d.
4	2. Study protocol
5	The experimental design is shown in Fig.2. The 10-week rats were divided into 6
6	groups of 8 rats each. After intraperitoneal administration of mixed anesthesia (1
7	mg/kg midazolam, 0.2 mg/kg medetomidine, and 2.5 mg/kg butorphanol), the
8	rats were intratracheally administered with 2.2 mg / kg BLM dissolved in 100 $\mu L$
9	PBS. The sham control rats received intratracheally PBS only. On the next day,
10	the administration of drugs, 0.3 mg/kg EP2 agonist (BLM + EP2), 0.3 mg/kg EP4
11	agonist (BLM + EP4), 30 mg/kg pirfenidone (BLM + pirfenidone), and 1 mL/kg
12	vehicle (50% PEG400, 50% DMSO) (BLM + vehicle), were performed by
13	subcutaneous injection three times a day, and 50 mg/kg nintedanib (BLM +
14	nintedanib) was administered orally once a day.
15	The dosage of EP2 agonist and EP4 agonist were set at 0.3 mg/kg
16	subcutaneous injection 3 times per day based on the pharmacokinetics and
17	EC <sub>50</sub> data.

1	The AUC for pirfenidone 600 mg oral single dose (single dose in clinical
2	practice) to adult males, described in the interview form 11), was 37.03 $\mu g \cdot$ hr /
3	mL and Cmax was 10.57 $\mu g$ / mL. The AUC determined by a single dose
4	administration of pirfenidone 30 mg / kg for S / D rats conducted by Kissei
5	Pharmaceutical Co. Ltd. was 33.79 $\mu g \cdot hr$ / mL and Cmax was 23.39. Because
6	blood concentrations in clinically applied doses in human were maintained, the
7	dosage and dose of pirfenidone in the present study was set to 30 mg / kg
8	subcutaneous injection three times a day.
9	The dosage of nintedanib was set according to the method in the study for
10	BLM-induced pulmonary fibrosis in rats performed by Wollin et al. 12)
11	Body weight measurements were taken daily before the first dose. We
12	euthanized the rats by abdominal aortic dissection under deep anesthesia on
13	day 21 of the administration of intervention agents. We removed both lungs to
14	evaluate the efficacy of intervention by the agents.
15	3. Measurement of lung weight

16 The right lung was removed and the weight was measured. The lung specimens

17 were quickly frozen with liquid nitrogen and stored at -30°C until the

1 measurements of hydroxyproline content and quantitative real-time polymerase 2 chain reaction (qRT-PCR). 3 4. Histological analysis 4 The left lung was removed, fixed by 10% neutral-buffered formaldehyde solution and embedded in paraffin. The sections were stained with Masson's trichrome 5 6 and examined by light microscopy. The fibrosis score was evaluated using the 7 modified Ashcroft's score 13). The lung section was examined at a magnification of 100×, and each field was visually scored from 0 points (normal lung) to 8 8 9 points (complete obstruction with fibroma mass) under light microscopy. The average score was calculated with all the observed fields for an entire lung. 10 5. Measurement of hydroxyproline content 11 12 A sample of right lung tissue was homogenized with 1,000 mg of lung tissue in 1 mL of PBS. Hydroxyproline content was measured by the assay kit (Chondrex 13 14 Inc, WA, USA) following measurement protocol.

6. mRNA expressions of the TGF-β1, procollagen1a1 (Col1a1), and Col1a2
 genes

1	The total RNA was extracted from right lung tissues using a RNeasy Kit
2	(Qiagen, Valencia, CA, USA) according to the manufacturer's designated
3	regimen. We synthesized cDNA from a 100ng total RNA template via reverse
4	transcription by using a Prime Script RT Enzyme Mix I (TakaraBio Inc., Japan).
5	The mRNA levels were quantified using the qRT-PCR system (Applied
6	Biosystems StepOnePlus; Thermo Fisher Scientific K.K.). The cDNAs of the rat
7	Col1a1, Col1a2, and TGF- $\beta$ 1 were amplified from single-stranded cDNA by PCR
8	using Taq DNA Polymerase with 0.4 $\mu M$ of each primer. The PCR primers of the
9	rat TGF- $\beta$ 1, Col1a1, Col1a2 genes, and 18S rRNA are described in Table1. The
10	qRT-PCR was performed using 10 $\mu mol$ samples consisting of 200 nM of each
11	primer, 12.5 $\mu L$ of SYBR premix Ex Taq (TakaraBio), and 10 ng of template
12	cDNA.
13	The experimental PCR protocol required 30 s of an initial denaturation at 96°C
14	followed by 40 cycles of 5 s of denaturation at 96°C, 30 s of annealing at 60°C,
15	and 60 s of extension at 60°C.
16	B. Confirmation of EP2 and EP4 expressions in the lung tissues

**1. Rat lung tissues** 

1 The lung tissues of rats with administration of PBS intratracheally were used as 2 negative control. The lung tissues of rats in BLM + vehicle group were used for the indication of pulmonary fibrosis. 3 4 2. Human lung tissues 5 Approval regarding using human lung materials in this study was obtained from 6 the Shinshu University Ethics committee (Authorization number: 2730, approval 7 date: May 12th, 2014). The lung tissues with pulmonary fibrosis were obtained from autopsy cases of 8 9 IPF in 2008 and 2009. The normal lung tissues for the control were obtained from resected lung lobes after lung lobectomy performed for the treatment of 10 lung cancer in the Department of Thoracic Surgery of Shinshu University. 11 12 Tissues were fixed in formalin and embedded in paraffin. 3. Immunohistochemical staining 13 14 We used Histofine SAB-PO kit (Nichirei Biosciences Inc., Tokyo, Japan) for immunostaining and performed the procedure following the protocol. The 15 16 SAB-PO kit contains biotin-labeled anti-mouse IgG + IgA + IgM rabbit antibody 17 as a second antibody and 3, 3-diaminobenzidine (DAB) as a chromogenic

1	substrate. The anti-EP2 rabbit monoclonal antibody (ab 167171, Abcam,
2	Cambridge, UK) was diluted at 1:1,250 with PBS and the anti-EP4 rabbit
3	polyclonal antibody (ab 133170, Abcam) was diluted at 1:50 with PBS and used
4	as the primary antibody.
5	C. Statistical analysis
6	Data were expressed as the mean $\pm$ standard deviation (SD) and processed by
7	using GraphPad Prism 7 (GraphPad Software, Inc. USA). The significance of
8	differences between each group was evaluated by one-way ANOVA with
9	post-hoc tests. A value of P < $0.05$ was considered to indicate statistical
10	significance.
11	Ⅲ. Results
12	A. Efficacy evaluation of the EP2 and EP4 agonists in the BLM-induced
13	pulmonary fibrosis rats
14	1. Histopathological effects of the intervention compounds
15	The alveolitis and patchy fibrosis with destruction around the bronchus
16	(representing the part surrounded by the red ellipse) were observed in the rat
17	BLM-induced lung fibrosis model. In the sham control, only normal lung findings

1	were observed (Fig. 3a). The pathological findings were improved in BLM + EP2
2	(Fig. 3c) and BLM + nintedanib (Fig. 3f) relative to those in BLM + vehicle (Fig.
3	3b). On the other hand, there was no pathological improvement in BLM + EP4
4	(Fig. 3d) and BLM + pirfenidone (Fig. 3e).
5	The mean fibrosis scores in groups of the sham control, BLM + vehicle, BLM +
6	EP2, BLM + EP4, BLM + pirfenidone, and BLM + nintedanib were $0.07 \pm 0.06$ ,
7	$3.10 \pm 0.75$ , $2.88 \pm 0.74$ , $3.41 \pm 0.60$ , $3.62 \pm 0.81$ , and $2.64 \pm 0.59$ , respectively.
8	The mean fibrosis scores of the BLM + EP2 and BLM + nintedanib were lower
9	than that of the BLM + vehicle; however, no statistical significance was observed.
10	(Fig. 4a).
11	2. Effects on lung weights
12	The average lung weight of each experimental group is shown in Fig. 4b. The
13	lung weight of BLM + vehicle was significantly higher than that of the sham
14	control. The average lung weights of BLM + EP2 and BLM + nintedanib were
15	significantly lower than that of BLM + vehicle. However, the EP4 and pirfenidone
16	did not show effects on the lung weights of rats with BLM-induced pulmonary

17 fibrosis.

# **3. Hydroxyproline content in lung tissues**

2	The hydroxyproline content in lung tissue is shown in Figure 4c. The content was
3	22.7 $\pm$ 13.2 µg/mL in the sham control, whereas it was increased to 85.3 $\pm$ 52.8
4	$\mu$ g/mL in the BLM + vehicle. The hydroxyproline content was slightly suppressed
5	to 71.0 $\pm$ 43.6 µg/mL in the BLM + EP2; however, there was no significant
6	difference between the BLM + EP2 and BLM + vehicle. No suppressive effect
7	was observed in the other groups, in which the hydroxyproline contents were
8	93.2 $\pm$ 61.4 µg/mL in the BLM + EP4, 96.0 $\pm$ 52.1 µg/mL in the BLM + pirfenidone,
9	and 85.0 $\pm$ 52.0 µg/mL in the BLM + nintedanib.
10	4. The mRNA expressions of TGF- $\beta$ 1, Col1a1, and Col1a2 genes
10 11	<b>4. The mRNA expressions of TGF-<math>\beta</math>1, Col1a1, and Col1a2 genes</b> The TGF- $\beta$ 1 mRNA was significantly increased in BLM + vehicle compared to
10 11 12	<b>4. The mRNA expressions of TGF-<math>\beta</math>1, Col1a1, and Col1a2 genes</b> The TGF- $\beta$ 1 mRNA was significantly increased in BLM + vehicle compared to that in the sham control. On the other hand, it was significantly suppressed in
10 11 12 13	4. The mRNA expressions of TGF- $\beta$ 1, Col1a1, and Col1a2 genes The TGF- $\beta$ 1 mRNA was significantly increased in BLM + vehicle compared to that in the sham control. On the other hand, it was significantly suppressed in BLM + EP2, BLM + EP4, BLM + pirfenidone and BLM + nintedanib compared
10 11 12 13 14	<b>4. The mRNA expressions of TGF-<math>\beta</math>1, Col1a1, and Col1a2 genes</b> The TGF- $\beta$ 1 mRNA was significantly increased in BLM + vehicle compared to that in the sham control. On the other hand, it was significantly suppressed in BLM + EP2, BLM + EP4, BLM + pirfenidone and BLM + nintedanib compared with that in BLM + vehicle (Fig. 5a).
10 11 12 13 14	<ul> <li>4. The mRNA expressions of TGF-β1, Col1a1, and Col1a2 genes</li> <li>The TGF-β1 mRNA was significantly increased in BLM + vehicle compared to</li> <li>that in the sham control. On the other hand, it was significantly suppressed in</li> <li>BLM + EP2, BLM + EP4, BLM + pirfenidone and BLM + nintedanib compared</li> <li>with that in BLM + vehicle (Fig. 5a).</li> <li>The expressions of Col1a1 mRNA were significantly enhanced in BLM + vehicle</li> </ul>
<ol> <li>10</li> <li>11</li> <li>12</li> <li>13</li> <li>14</li> <li>15</li> <li>16</li> </ol>	<ul> <li>4. The mRNA expressions of TGF-β1, Col1a1, and Col1a2 genes</li> <li>The TGF-β1 mRNA was significantly increased in BLM + vehicle compared to</li> <li>that in the sham control. On the other hand, it was significantly suppressed in</li> <li>BLM + EP2, BLM + EP4, BLM + pirfenidone and BLM + nintedanib compared</li> <li>with that in BLM + vehicle (Fig. 5a).</li> <li>The expressions of Col1a1 mRNA were significantly enhanced in BLM + vehicle</li> <li>compared with those in the sham controls. The mRNA expression of Col1a1</li> </ul>

1	compared with that in BLM + vehicle (Fig. 5b). The expression of Col1a2 mRNA
2	was increased in BLM + vehicle compared to those in the sham controls without
3	significant difference. The mRNA expression of Col1a2 gene was significantly
4	suppressed in BLM + EP2, BLM + pirfenidone, and BLM + nintedanib compared
5	with that in BLM + vehicle (Fig. 5c).
6	B. Confirmation of EP2 and EP4 expressions in the lung tissues
7	1. Immunostaining of EP2 and EP4 in lung tissues of experimental rats
8	The immunostaining showed very strong expressions of the EP2 (Fig. 6a) and
9	EP4 (Figure 6c) in the lung tissues of rats with BLM-induced pulmonary fibrosis
10	compared to those in the negative control (Fig. 6b, d).
11	2. Immunostaining of EP2 and EP4 in human lung of IPF
12	The immunostaining showed that the EP2 was positive in human IPF tissues
13	(Fig. 7a) but negative in normal lung tissues (Fig. 7b). On the other hand, the
14	EP4 showed weak expression in the alveolar epithelium of IPF tissues (Fig. 7c),
15	but negative expression in the normal lung tissues (Fig. 7d).
16	

# **IV. Discussion**

2	Among the four subtypes of the $PGE_2$ receptor, EP2 and EP4 are coupled to G
3	protein to increase cAMP. Increased intracellular cAMP levels were reported to
4	suppress the inductive activity of TGF- $\beta$ to attenuate fibroblast migration
5	14). TGF- $\beta$ 1 is a key cytokine whose sustained production causes the
6	development of tissue fibrosis. Repeated injury with continued autoinduction of
7	TGF- $\beta$ 1 overrides the normal termination signals, thus creating a chronic vicious
8	circle of TGF- $\beta$ 1 overproduction 15). In the present study, the mRNA production
9	of TGF- $\beta$ 1 gene was suppressed by the administration of the EP2 and EP4
10	agonists, suggesting a hindrance of the vicious cycle of TGF- $\beta$ 1 overproduction.
11	The suppressions of EP2 on the expressions of mRNA of TGF- $\beta$ 1 gene and
12	procollagen gene in the BLM-induced pulmonary fibrosis rats were equivalent to
13	the suppression of nintedanib in the rats with pulmonary fibrosis. Our study
14	reproduced the same results for BLM + nintedanib as those reported by Wollin
15	12). Despite the significant reductions in mRNA of TGF- $\beta$ 1 and procollagen
16	genes, there was no significant difference in the mean fibrosis scores and
17	hydroxyproline contents between the BLM + EP2 and BLM + vehicle groups.

1	A plausible explanation is the problem of the evaluation method. In the present
2	study, the pulmonary fibrosis in rats was induced by intratracheal administration
3	of BLM. Thus, the fibrosis was remarkably developed around the bronchus. The
4	BLM + vehicle formed strong fibrosis around the bronchiole, which was
5	obviously improved with the administration of EP2 and nintedanib. The fibrosis
6	score was calculated by the average score of about 100 fields under light
7	microscopic observation for each rat. The fact is that the pathological parts of the
8	fibrosis were not evenly distributed on the whole lung fields, with focus on some
9	of the lung fields and scattering on the other fields. Thus, the significant
10	differences in the fibrosis score vanished with the average calculation due to the
11	large SD in the statistics. The reason for no significant difference of
12	hydroxyproline contents between the BLM + EP2 and BLM + vehicle groups was
13	similar. In addition, the weight of the entire right lung was significantly lower in
14	BLM + EP2 and BLM + nintedanib than in BLM + vehicle. The weight of lung with
15	fibrosis is heavier than that of normal lung. We believe that antifibrosis was
16	achieved in the BLM + EP2 and BLM + nintedanib groups.

1	In addition, significant differences in the Col1a1 mRNA expression and
2	different tendencies of fibrosis were observed between the rats treated with EP2
3	and EP4 agonists. EP4 was found to have induced desensitization and
4	internalization by stimulation of $PGE_2$ 16). Therefore, by stimulation with the
5	same amount of PGE2, the expression of cAMP in EP2-expressing cells was
6	observed to be 7-fold as compared with EP4-expressing cells 17). The
7	production of cAMP inhibits collagen synthesis and prevents differentiation of
8	fibroblasts into probiogenic myofibroblasts 18). Therefore, the inhibitory effects
9	of EP2/4 agonists on the procollagen mRNA can be explained. We speculate
10	that a possible role of the cAMP level may be one of the causes of the difference
11	in the efficacy between the EP2 and EP4 agonists in the present study. Moreover,
12	it has been reported that another signaling pathway, which is independent of
13	cAMP, could be present in EP4.16)19)20)
14	Pirfenidone is clinically applied to treat patients with IPF. In the present study,
15	pirfenidone failed to reduce the BLM-induced pulmonary fibrosis. Schaefer et al.
16	reviewed experiments on the effects of pirfenidone on the animal model of
17	BLM-induced pulmonary fibrosis 21) and stated in summary that the anti - fibrotic

1	effect was mainly found in mice and hamsters 22),23),24),25). In addition,			
2	pirfenidone was administered by oral administration in these studies. Recently, i			
3	was reported that oral administration of 50mg/kg pirfenidone could reduce			
4	BLM-induced pulmonary fibrosis in rats. 26) In the present study, pirfenidone			
5	was administered to rats by 30 mg / kg subcutaneous injection three times a day.			
6	We believed that the dose would be approximately equal to that when orally			
7	administered to humans in clinical practice. However, our dose and animal			
8	setting or administration route may be related to the lack of preventive effects on			
9	BLM-induced pulmonary fibrosis in the present study. We need further			
10	experiments at various doses and oral administration of pirfenidone in			
11	BLM-induced pulmonary fibrosis.			
12	The present study demonstrated that EP2 and EP4 were expressed in human			
13	and rat lung tissues with fibrosis but not normal lung tissues, which are			
14	noteworthy new findings in pulmonary fibrosis. The expression of the receptors			
15	suggested that EP2 and EP4 are related to the development of pulmonary			
16	fibrosis. However, we found that EP4 over-expression was not apparent			
17	compared with that of EP2. The lack of preventive effects of EP4 agonist on			

1	pulmonary fibrosis could be related the low expression in the lung. However, we
2	could show that the EP2 agonist contributes to the prevention of fibrosis in an
3	experimental model. We need further studies using experimental models and
4	clinical samples; however, the present study indicates that the EP2 agonist may
5	be an effective therapeutic agent for IPF.
6	Conflicts of Interest
7	The authors have no conflicts of Interest in the present study.
8	Acknowledgments
9	The authors would like to thank our research colleagues: Dr. Hiroyuki Owaki, Dr.
10	Makoto Moro and Dr. Hideki Ohnota in the Department of Drug Discovery
11	Science of Shinshu University School of Medicine, Dr. Tomoya Onozato and Dr.
12	Morimichi Hayashi in the Research Division, Safety Research Laboratory of
13	Kissei Pharmaceutical Co., Ltd., Dr. Yasushi Takigawa and Mr. Toshihiro
14	Nishimura in the Research Division in the Central Research Laboratories of
15	Kissei Pharmaceutical Co., Ltd., Mrs. Hitomi Imamura and Mrs. Yunden Droma
16	in the First Department of Internal Medicine of Shinshu University School of

1	Medicine, and Mr. Naoyuki Tatsumi and Dr. Toshihito Hiroi in the Pharmacology		
2	and Pharmacokinetics Research Lab of Kissei Pharmaceutical Co., Ltd.		
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### Table 1

### The primer sequence of cDNA used in the present study

cDNA, (ID)	primers	sequence
TGF-β1	forward	5'-CATTGCTGTCCCGTGCAGA-3'
(RA059245)	reverse	5'-AGGTAACGCCAGGAATTGTTGCTA-3'
Col1a1	forward	5'-CTGCATCAGGGTTTCAGAGCAC-3'
(RA069752)	reverse	5'-TCCACATGCTTTATTCCAGCAATC-3'
Col1a2	forward	5'-CTGGATTGACCCTAACCAAGGATG-3'
(RA069679)	reverse	5'-TTGACAGGTTGGGCCGGA-3'
18S rRNA	forward	5'-AAGTTTCAGCACATCCTGCGAGTA-3'
(RA015374)	reverse	5'-TTGGTGAGGTCAATGTCTGCTTTC-3'

1	Figure legends
2	Fig. 1
3	a Structural formula of EP2 agonist
4	b Structural formula of EP4 agonist
5	c Pharmacokinetic study of EP2 and EP4 agonists in rats
6	d Functional assay of transient expression in HEK293T cell
7	Fig. 2
8	Study design for examining the antifibrotic effects of the EP2 and EP4
9	agonists on bleomycin (BLM)-induced pulmonary fibrosis rats.
10	Fig. 3
11	Histological observation of the left lung (Masson trichrome; original
12	magnification ×20).
13	a PBS only, as a sham control
14	b BLM + vehicle
15	c BLM + EP2 agonist
16	d BLM + EP4 agonist
17	e BLM + pirfenidone

1 <b>f BLM + nintedanib</b> .	
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2	Fig. 4
3	a Histological score by modified Aschoff`s score method: **** indicates p <
4	0.0001 vs. BLM + vehicle group
5	b The right lung weight of experimental rats: * indicates $p < 0.05$ vs. BLM +
6	vehicle, ** indicates p < 0.005 vs. BLM + vehicle and *** indicates p < 0.001 vs.
7	BLM + vehicle.
8	c Hydroxyproline contents in 1 gram of right lung tissue
9	Fig. 5
10	a The mRNA expression of TGF-β1
11	b The mRNA expression of procollagen 1a1
12	c The mRNA expression of procollagen 1a2: * indicates $p < 0.05$ vs. BLM +
13	vehicle, ** indicates $p < 0.005$ vs. BLM + vehicle and *** indicates $p < 0.001$ vs.
14	BLM + vehicle
15	Fig. 6
16	Immunostaining of rat lung; original magnification ×100.
17	a The EP2 of BLM-induced pulmonary fibrosis rat

1	b The EP2 of rat administered with PBS i.t.
2	c The EP4 of BLM-induced pulmonary fibrosis rat
3	d The EP4 of rat administered with PBS i.t. The parts with EP2 expression
4	reacted with DAB and showed a brown color. The parts without expression of
5	EP2 and EP4 exhibited a blue color.
6	Fig. 7
7	Immunostaining of human lung; original magnification ×100.
8	a The EP2 was expressed in human lung of IPF with a brown color.
9	b The EP2 was not expressed in human normal lung tissue with a blue color.
10	c The EP4 was expressed in human lung of IPF, d) The EP4 was not expressed
11	in normal lung tissue.
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Fig.1

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	EP2 agonist	EP4 agonist
C <sub>max</sub> (ng/mL)	771.3	1511
T <sub>max</sub> (min)	50.0	50.0
AUC <sub>(0-X)</sub> (ng • min/mL)	172,330.3	320,213
Protein binding (%) rat plasma 37°C, 10min	88.66	99.91

Cmax: maximum serum drug concentration, Tmax: Time to reach maximum serum concentration,

AUC(0-X): Area under the concentration-time curve over x min

# d) Functional assay of transient expression in HEK293T cell

	EP2 agonist	EP4 agonist
Rat EP2 EC50 (nM)	0.010	117.8
Rat EP4 ECso (nM)	0.12	0.0005

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