

Synthesis of legioliulin, a fluorescent isocoumarin compound, isolated from *Legionella dumoffii* using cyclic acylpalladation and Heck reaction

Masaki Asai,^a Yasunao Hattori,^b Hidefumi Makabe*^a

^aGraduate School of Science and Technology, Department of Agriculture, Division of Food Science and Biotechnology, Shinshu University, 8304 Minami-minowa, Kami-ina, Nagano, 399-4598, Japan

^bCenter for Instrumental Analysis, Kyoto Pharmaceutical University, Yamashina-ku, Kyoto 607-8412, Japan

*Corresponding author. Tel. +81 265 77 1630; fax +81 265 77 1700; e-mail: makabeh@shinshu-u.ac.jp

Abstract: Concise synthesis of legioliulin, an isocoumarin compound isolated from *Legionella dumoffii*, was achieved. Isocoumarin ring of legioliulin was constructed using cyclic acylpalladation. Chain elongation was performed using Heck reaction using *t*-butylphosphine as a ligand.

Key words: natural product, polyketides, isocoumarin, acylpalladation, Heck reaction

Legionellae, which are responsible for Legionnaire's disease, are facultative intracellular gram-negative bacteria.¹ 10 species of *Legionella* exhibit blue-white and dark-red autofluorescence. In 2004, Amemura-Maekawa and co-workers isolated legioliulin (**1**) from *Legionella dumoffii*.² Legioliulin (**1**) is a new isocoumarin compound and fluorescent substance. The study on biosynthesis of legioliulin (**1**) was reported by Bode and co-workers in 2013.³ This compound did not show any cytotoxicity against human monocytic cell line U937, neither exhibit antimicrobial activity against *Staphylococcus aureus* and *E. coli*.² The real function and the detailed biological activity of legioliulin (**1**) are still unknown. Thus we began to synthesize legioliulin (**1**) to find out its function and biological properties. Here, we wish to report the concise synthesis of legioliulin (**1**) using cyclic acylpalladation and Heck reaction as the key steps (Figure 1).

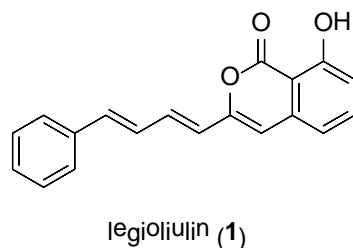
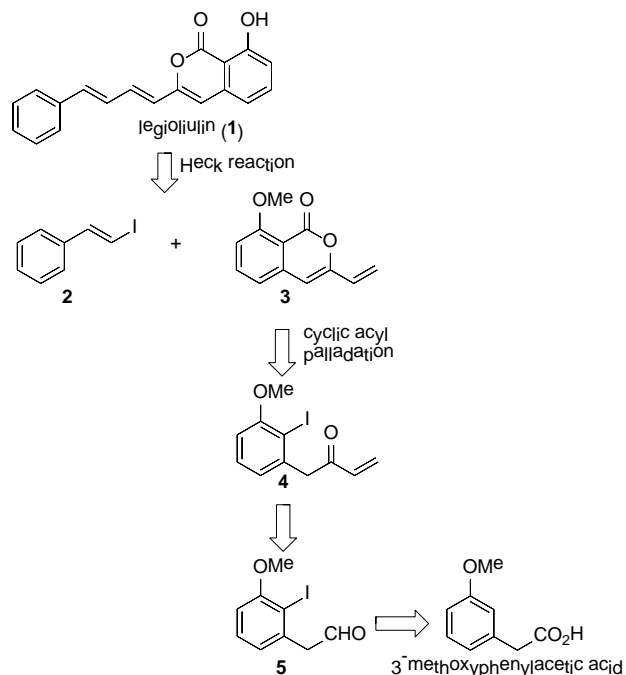


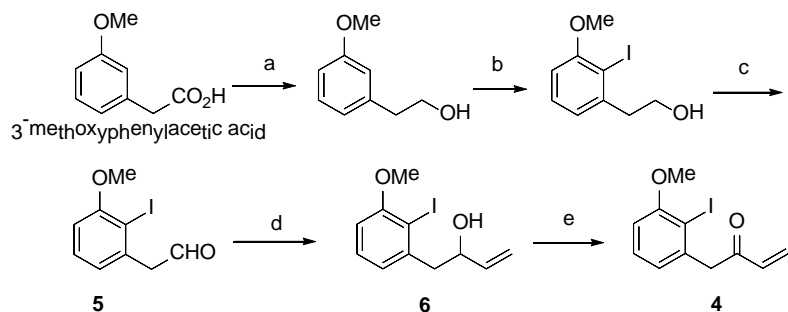
Figure 1. The structure of legioliulin (**1**).

The synthetic strategy shows in Scheme 1. The side chain would be introduced using Heck reaction. The isocoumarin part of **1** would be constructed using cyclic acyl palladation trapping by *O*-enolate from iodoenone **4**. Iodoenone **4** would be prepared from known aldehyde **5**.⁴ Aldehyde **5** would be synthesized from commercially available 3-methoxyphenylacetic acid (Scheme 1).



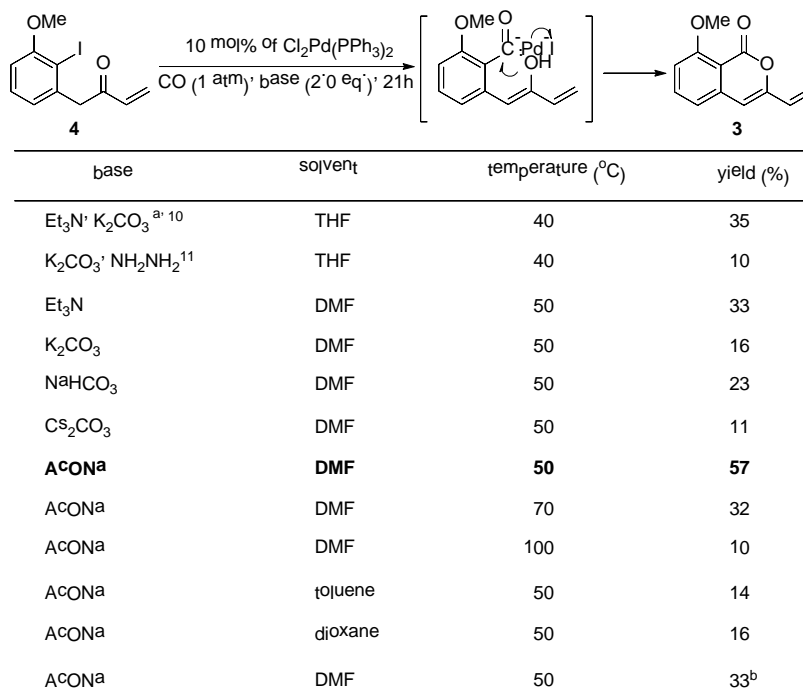
Scheme 1. Synthetic strategy of boronolide (**1**) and deacetylboronolide (**2**).

Scheme 2 shows the preparation of cyclization precursor **4**. Reduction of carboxylic acid of 3-methoxyphenylacetic acid with LiAlH_4 afforded corresponding alcohol subsequent iodination at *ortho* position and oxidation of primary alcohol gave aldehyde **5**. The yield of iodination was rather low because this compound was unstable. Alkylation of **5** using vinylmagnesium bromide afforded allylic alcohol **6**. Oxidation of the secondary hydroxy group of **6** with Dess-Martin periodinane in the presence of pyridine gave enone **4** (Scheme 2).



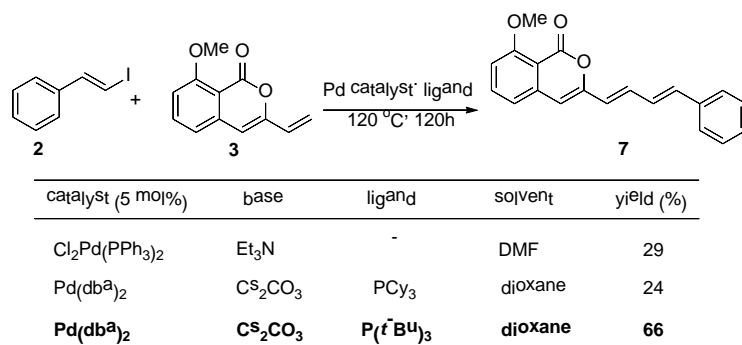
Scheme 2. Synthesis of cyclization precursor **4**. Reagents and conditions: (a) LiAlH_4 , THF, 96%; (b) $n\text{-BuLi}$, I_2 , Et_2O , 48%; (c) Dess-Martin periodinane, CH_2Cl_2 , 99%; (d) vinylmagnesium bromide, Et_2O , 89%; (e) Dess-Martin periodinane, pyridine, CH_2Cl_2 , 74%.

The cyclization precursor was in hand, we examined cyclic acylpalladation trapping by *O*-enolate to obtain **3**.⁵⁻⁹ As shown in Table 1, the choice of the base and solvent was very important. We found that AcONa as a base in DMF at 50°C furnished desired 5-*O*-endo cyclized product **3** in 57% yield. Raising reaction temperature and changing solvent did not improve the yield. We also examined other Pd catalyst such as $\text{Pd}(\text{PPh}_3)_4$ and $\text{Pd}(\text{dba})_2$ with ligand such as dppf and dppp. However, the yield of **3** was not improved (Table 1).

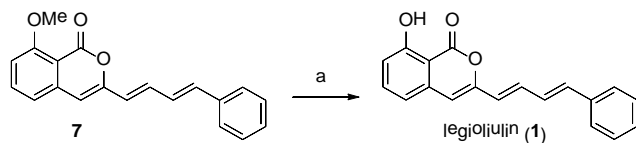
Table 1. Cyclic acylpalladation of 4.

^a4.0 eq of base was used; ^b5 mol% of Pd catalyst was used

Next, we investigated the construction of the carbon framework of legioliulin (**1**) using Heck reaction. In this reaction, we found that the phosphine ligand greatly affected the reactivity. We previously reported that P(*t*-Bu)₃ was effective ligand to form stilbene framework.¹² Fu reported that the effect of P(*t*-Bu)₃.¹³ According to his report, the effect of P(*t*-Bu)₃ is electron donating as well as accelerating reductive elimination. As a result P(*t*-Bu)₃ afforded desired product **7** in 66% yield (Table 2).

Table 2. Heck reaction of 3 with (1E)-1-iodostyrene (2).

Finally, deprotection of methyl group at C-8 position using BBr₃ in CH₂Cl₂ at -78°C gave legioliulin (**1**). The ¹H and ¹³C NMR data of the synthetic legioliulin (**1**) were consistent with those of the reported values (Scheme 3).^{2, 14}



Scheme 3. Synthesis of legioliulin (**1**). Reagent and condition: (a) BBr_3 , CH_2Cl_2 , -78°C , 76%.

In conclusion, concise synthesis of legioliulin (**1**) was achieved using acylpalladation and Heck reaction. The studies on biological activities of legioliulin (**1**) is now underway.

Acknowledgements

This work was supported by JSPS KAKENHI Grant Number 15K07408 to H. M.

References and notes

1. Fields, B. S.; Benson, R. F.; Besser, R. E. *Clin. Microbiol. Rev.* **2002**, *15*, 506.
2. Amemura-Maekawa, J.; Hayakawa, Y.; Sugie, H.; Moribayashi, A.; Kura, F.; Chang, B.; Wada, A.; Watanabe, H. *Biochem. Biophys. Res. Commun.* **2004**, *323*, 954.
3. Ahrendt, T.; Miltenberger, M.; Haneburger, I.; Kirchner, F.; Kronewerth, M.; Brachmann, A. O.; Hilbi, H.; Bode, H. B. *ChemBioChem.* **2013**, *14*, 1415.
4. Fan, L.; Takizawa, S.; Takeuchi, Y.; Takenaka, K.; Sasai, H. *Org. Biomol. Chem.* **2015**, *13*, 4837.
5. Negishi, E.; Makabe, H. *Handbook of Organopalladium Chemistry for Organic Synthesis*, John Wiley & Sons: New York, **2002**, 2455.
6. Negishi, E.; Copéret, C.; Ma, S.; Liou, S. Y.; Liu, F. *Chem. Rev.* **1996**, *96*, 365.
7. Negishi, E.; Copéret, C.; Sugihara, T.; Shimoyama, I.; Zhang, Y.; Wu, G.; Tour, J. M. *Tetrahedron* **1994**, *50*, 425.
8. Negishi, E.; Makabe, H.; Shimoyama, I.; Wu, G.; Zhang, Y.; *Tetrahedron* **1998**, *54*, 1095
9. Uozumi, Y.; Mori, E.; Mori, M.; Shibasaki, M. *J. Organomet. Chem.* **1990**, *399*, 93.
10. Makabe, H.; Okajima, M.; Konno, H.; Kamo, T.; Hirota, M. *Biosci. Biotechnol. Biochem.* **2003**, *67*, 2658.
11. Hikosaka, G.; Hattori, Y.; Makabe, H. *Tetrahedron: Asymmetry* **2014**, *25*, 1367.
12. Iijima, T.; Makabe, H. *Biosci. Biotechnol. Biochem.* **2009**, *73*, 2547.
13. Fu, G. C. *Acc. Chem. Res.* **2008**, *41*, 1555.

14. The data of legioliulin (**1**): Mp. 107-108°C; IR (KBr): 3250-3025, 2917, 2849, 1687, 1625, 1609, 1562, 1496, 1460, 1343, 1315, 1223, 1168, 1141, 1090, 1049, 982, 842, 749, 690 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ : 11.00 (1H, s), 7.55 (1H, t, $J = 7.5$ Hz), 7.45 (2H, d, $J = 7.5$ Hz), 7.34 (2H, t, $J = 7.5$ Hz), 7.26 (1H, t, $J = 7.5$ Hz), 7.22 (1H, dd, $J = 15.0, 11.0$ Hz), 6.92 (1H, d, $J = 7.5$ Hz), 6.89 (1H, dd, $J = 15.5, 11.0$ Hz), 6.86 (1H, d, $J = 7.5$ Hz), 6.81 (1H, d, $J = 15.5$ Hz), 6.39 (1H, s), 6.24 (1H, d, $J = 15.0$ Hz); ^{13}C NMR (125 MHz, CDCl_3) δ : 166.00, 161.71, 152.06, 137.94, 137.32, 137.03, 136.63, 133.81, 128.80, 128.42, 127.45, 126.83, 122.36, 116.31, 115.24, 106.39, 106.05 ppm. HRMS-EI: m/z $[\text{M}]^+$: calcd for $\text{C}_{19}\text{H}_{14}\text{O}_3$; 290.0943, found: 290.0945.