# Expression of Polypeptide N-acetylgalactosaminyl Transferase-3 and Its Association with Clinicopathological Factors in Thyroid Carcinomas

Yasuhiro Mochizuki,<sup>1</sup> Ken-ichi Ito,<sup>1</sup> Hiroto Izumi,<sup>2</sup> Kimitoshi Kohno,<sup>2</sup> and Jun Amano<sup>3</sup>

*Background:* Polypeptide N-acetylgalactosaminyl transferase-3 (GalNAc-T3) has been reportedly expressed in several human adenocarcinomas and is associated with clinicopathological features of tumors. We investigated the clinicopathological significance of GalNAc-T3 in thyroid cancer.

*Patient and Methods:* We evaluated the expression of GalNAc-T3 in 167 patients with thyroid cancer using a specific antibody and analyzed the association between its expression and clinicopathological features.

**Results:** GalNAc-T3 was expressed in 85.8% of normal follicular epithelial cells. In papillary carcinomas, positive staining was observed in 101 (73.7%) cases. Well-differentiated components (papillary and follicular) of papillary carcinomas were significantly more frequently positive than poorly differentiated components (trabecular and solid) (p < 0.01), and GalNAc-T3 was highly expressed in papillary carcinomas that had invaded beyond the thyroid capsule (p = 0.026). GalNAc-T3 was expressed in 40% and 20% of well and poorly differentiated components of follicular carcinomas, respectively. Thirteen of 15 anaplastic carcinomas were negative for GalNAc-T3 and thyroglobulin. Positive staining for GalNAc-T3 was not observed in any of the medullary carcinomas.

*Conclusions:* Our data suggest that GalNAc-T3 expression may be a useful indicator of tumor differentiation in thyroid carcinomas.

# Introduction

"HYROID CANCER IS THE MOST COMMON endocrine malignancy worldwide. Thyroid carcinomas are classified into papillary, follicular, medullary, and anaplastic, with papillary and follicular carcinomas being designated as differentiated thyroid carcinomas. Papillary, follicular, and anaplastic carcinomas originate from thyroid follicular cells, whereas anaplastic carcinoma is thought to arise mainly from differentiated carcinomas. In contrast, medullary carcinoma derives from thyroid parafollicular (neuroendocrine) C cells. The clinical behavior of thyroid carcinomas varies remarkably depending on the histological type. Differentiated thyroid carcinoma comprises the majority of thyroid carcinomas and carries a favorable prognosis. In contrast, anaplastic thyroid carcinoma is one of the most virulent human malignancies, with a mean survival time of less than one year from diagnosis regardless of the treatment administered (1–3). Although approximately half of the patients with anaplastic carcinomas have previous or coexistent differentiated thyroid carcinomas, the underlying molecular mechanisms of anaplastic transformation remain poorly understood.

It is known that structural changes in cell surface glycoproteins influence the biological behavior of cancer cells during malignant transformation and tumor progression (4,5). Mucin glycoproteins are widely distributed on the epithelial cell surface and play important functional roles in cell adhesion, cell differentiation, proliferation, carcinogenesis, cancer metastasis, and basic immunological systems (6–8). Mucintype *O*-linked carbohydrates constitute approximately 80% of the total molecular mass of these glycoproteins, and *O*-linked carbohydrate antigens, such as carcinoembryonic antigen, CA19-9, sialyl LewisX, sialyl Tn, and Tn, have been reported to be associated with invasion, recurrence, and prognosis in cancer patients (9–15). Thus, *O*-glycosylation has been shown to play important roles in tumor progression (16–18).

UDP-GalNAc:polypeptide N-acetylgalactosaminyl transferases (GalNAc-Ts) are a key enzyme family of more than 20 isoforms that catalyze the O-glycosylation reaction, the primary step of alpha-O-glycoside bond formation between

<sup>&</sup>lt;sup>1</sup>Division of Breast and Endocrine Surgery, <sup>3</sup>Department of Surgery, Shinshu University School of Medicine, Matsumoto, Nagano, Japan. <sup>2</sup>Department of Molecular Biology, University of Occupational and Environmental Health, Kitakyushu, Fukuoka, Japan.

GalNAc and serine/threonine residues, on the mucin scaffold (19-21). Although GalNAc-T3 is one of the isoforms expressed in various organs, its expression level differs among them. In normal tissues, GalNAc-T3 is expressed at higher levels in the pancreas and testis and at lower levels in the kidney, prostate, spleen, ovary, intestine, and colon, whereas GalNAc-T3 mRNA has been detected in organs containing secretary epithelial glands (19,22-24). It is hypothesized that the differential expression of GalNAc-T3 may affect the specialized functions of glycoproteins produced by normal and malignant cells, and an association between GalNAc-T3 and the biological properties of both normal and malignant cells has been demonstrated (18). Previous studies have demonstrated that the expression of GalNAc-T3 is associated with the differentiation, aggressiveness, and prognosis of gastric (25), pulmonary (26), colorectal (27), gallbladder (28), bile duct (29), pancreatic (30), and esophageal carcinomas (31). However, to the best of our knowledge, GalNAc-T3 expression has not been evaluated in thyroid carcinomas. The purpose of this study was to evaluate the relation between GalNAc-T3 expression and pathological features in thyroid carcinomas.

#### Materials and Methods

### Patients and clinical materials

This study was conducted according to the ethics guidelines of the Declaration of Helsinki, and specific approval was obtained from the Ethics Committee of Shinshu University School of Medicine. The specimens studied were obtained from 168 patients with thyroid cancers who were diagnosed and treated in Shinshu University Hospital from 1995 to 2005. Of 167 cases, 134 were papillary carcinomas, 11 follicular carcinomas, 15 anaplastic carcinomas, and 7 medullary carcinomas. All patients except eight with anaplastic carcinomas underwent surgical resection. The eight patients with anaplastic carcinomas were treated with radiation and/or chemotherapy. In 141 of 160 patients who underwent surgical resection, normal thyroid gland tissue was also observed. Clinicopathological data were obtained by retrospective chart review.

#### Immunohistochemical staining and evaluation

Polyclonal antibodies against human GalNAc-T3 were generated by multiple immunization of a New Zealand white rabbit using synthetic peptides as described previously (32). A monoclonal antibody to thyroglobulin (1D4) was purchased from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA). A formalin-fixed, paraffin-embedded 3 µm section was obtained from all 168 primary lesions. Sections were deparaffinized in xylene, hydrated through a graded series of ethanol, and immersed in 3% hydrogen peroxide in 100% methanol for 30 minutes to inhibit endogenous peroxidase activity. To activate the antigens, the sections were boiled in 10 mM citrate buffer (pH 6.0) for 12 minutes. After rinsing in phosphatebuffered saline (PBS), the sections were incubated with normal goat serum for 10 minutes, and then incubated overnight at 4°C in a humidity chamber with the primary antibody to GalNAc-T3 at 1/40,000 dilution. After washing thrice with PBS, the sections were incubated with biotinylated anti-rabbit immunoglobulin for 60 minutes. After washing again with PBS, the sections were incubated for 60 minutes with avidin and a biotinylated horseradish peroxidase macromolecular complex. Diaminobenzidine was used as a chromogen, and the sections were lightly counterstained with hematoxylin. Cells were judged positive for GalNAc-T3 expression when granulated cytoplasmic staining was observed under highpower magnification ( $10 \times 40$ ).

#### Statistical analysis

The relationship between clinicopathological features and the expression of GalNAc-T3 was examined by Fisher's exact test. Differences with a *p*-value of less than 0.05 were considered statistically significant. The chi-square test was used for immunohistochemical analysis of the clinical specimens.

# Results

# GalNAc-T3 expression in normal thyroid gland

To evaluate the expression of GalNAc-T3 in the normal thyroid gland, 141 normal thyroid tissues were stained for GalNAc-T3 expression. GalNAc-T3 was expressed in 121 of 141 (85.8%) normal follicular epithelial cells (Fig. 1). The association between GalNAc-T3 expression in the normal thyroid gland and both sex and age is summarized in Table 1. The expression of GalNAc-T3 was observed more frequently in the normal thyroid gland of male patients compared with female patients; however, there was no statistical significance between GalNAc-T3 expression and sex or age.

### GalNAc-T3 expression in thyroid carcinomas

In thyroid carcinomas, the biological behavior of the tumor is known to depend mainly on its histological type. To evaluate differences in expression of GalNAc-T3 with the histological type, 167 thyroid carcinoma tissue samples (134 papillary, 11 follicular, 15 anaplastic, 7 medullary) were immunohistochemically analyzed for GalNAc-T3 expression (Table 2; Fig. 2). In papillary carcinomas, positive staining for GalNAc-T3 was observed in 101 of 134 tumors (73.7%), whereas in follicular carcinomas, the expression of GalNAc-T3 was detected in 5 of 11 tumors (45.5%). In contrast, only 2 of 15 (13.3%) anaplastic carcinomas and none of 7 (0%) medullary tumors were positive for GalNAc-T3. Thus, Gal-NAc-T3 was significantly more frequently expressed in the differentiated carcinomas (papillary and follicular) compared with the other entities (p < 0.001).

### GalNAc-T3 expression in papillary thyroid carcinomas

As the expression of GalNAc-T3 was detected most frequently in papillary carcinomas, we further evaluated the association between GalNAc-T3 expression and its clinicopathological features. The clinicopathological features of 134 papillary carcinomas are summarized in Table 3. As differentiated papillary carcinoma often comprises diverse histological components, we examined the expression of GalNAc-T3 in individual histological components of papillary carcinoma classified pathologically (Fig. 2). The well-differentiated components of papillary thyroid carcinomas were mainly papillary (Fig. 2A) and/or follicular (Fig. 2B). In contrast, poorly differentiated components were mainly trabecular (Fig. 2C) and/ or solid (Fig. 2D). Two hundred fifteen well-differentiated



FIG. 1. Expression of GalNAc-T3 in the normal thyroid gland. (A, B) Immunohistochemical analyses using GalNAc-T3 antibody showed positive staining in the follicular epithelial cells. (C, D) Negative control staining with nonimmune rabbit serum showed no staining in the follicular epithelial cells. GalNAc-T3, polypeptide N-acetylgalactosaminyl transferase-3. Color images available online at www.liebertpub .com/thy

components (102 papillary and 113 follicular) and 89 poorly differentiated components (49 trabecular and 40 solid) were detected in 134 tumors. We next evaluated GalNAc-T3 expression in individual components. Positive staining for GalNAc-T3 was detected in 152 (70.7%) of 215 well-differentiated components, whereas only 13 (14.6%) of 89 poorly differentiated components were positive for GalNAc-T3. Thus, the well-differentiated components of papillary carcinomas showed GalNAc-T3 expression significantly more frequently compared with poorly differentiated components (p < 0.001). With regard to other clinicopathological factors, a statistically significant association between the expression of GalNac-T3 and the extent of extraglandular tumor invasion was observed (p < 0.05).

#### GalNAc-T3 expression in follicular thyroid carcinomas

With regard to follicular thyroid carcinomas, 5 (45.5%) of 11 tumors expressed GalNAc-T3. Follicular carcinomas consist of both well-differentiated (follicular) and poorly differentiated components (trabecular or solid). We evaluated the expression of GalNAc-T3 in individual components (Table 4;

Table 1.	GalNAc-T3	Expression
in Noi	rmal Thyroi	d Gland

	No. of cases	No. of positive GalNAc-T3 expression	%	р
All cases Sex	141	121	85.8	
Male	33	31	93.9	0.21
Female Age	108	90	83.3	
<45	52	42	80.8	0.19
≥45	89	79	88.8	

GalNAc-T3, polypeptide N-acetylgalactosaminyl transferase-3.

Fig. 2). Ten well-differentiated and 10 poorly differentiated components were detected in 11 follicular carcinomas (Fig. 2E, F). The expression of GalNAc-T3 was more frequently observed in the well-differentiated components compared with the poorly differentiated components (40.0% vs. 20.0%, respectively); however, no significant difference was observed.

# Expression of thyroglobulin and GalNAc-T3 in differentiated thyroid carcinomas

Thyroglobulin is a protein produced in the follicular cells of the normal thyroid gland and in differentiated carcinoma cells. Consequently, thyroglobulin is used as a marker for the differentiation of both normal and malignant thyroid tissue. To evaluate any correlation between GalNAc-T3 expression and the production of thyroglobulin in thyroid carcinomas, the expression of both GalNAc-T3 and thyroglobulin was assessed by immunostaining (Table 5; Fig. 3A, B). In papillary carcinomas, thyroglobulin was detected in 208 of 215 (96.7%) well-differentiated components and in 72 of 89 (80.9%) poorly differentiated components. However, no significant association was observed between the expression of thyroglobulin and GalNAc-T3. With regard to follicular carcinomas, thyroglobulin was detected in all tumors except one, and no

TABLE 2. GALNAC-T3 EXPRESSION IN FOUR HISTOLOGICALLY DIFFERENT THYROID CARCINOMAS

	No. of cases	No. of positive GalNAc-T3 expression	%	р
Papillary carcinoma	134	101	73.7	< 0.001
Follicular carcinoma	11	5	45.5	
Anaplastic carcinoma	15	2	13.3	
Medullary carcinoma	7	0	0	
Total	167	108	64.7	

FIG. 2. Expression of GalNAc-T3 in thyroid carcinomas. Representative findings of immunohistochemical analyses in the papillary carcinoma: positive staining in (A) the papillary component and (B) the follicular component; negative staining in (C) the trabecular component and (D) the solid component. Expression of GalNAc-T3 in the follicular thyroid carcinoma: (E) positive staining in the follicular component and (F) negative staining in the solid component. Expression of GalNAc-T3 and thyroglobulin in the medullary thyroid carcinoma: negative staining for (G) GalNAc-T3 and (H) thyroglobulin. Color images available online at www.liebertpub .com/thy



correlation between the expression of thyroglobulin and GalNAc-T3 was observed. With regard to medullary carcinomas, thyroglobulin and GalNAc-T3 were not detected in any of the studied tumors (Fig. 2G, H).

# Expression of thyroglobulin and GalNAc-T3 in anaplastic thyroid carcinomas

In anaplastic thyroid carcinomas, the expression of GalNAc-T3 was not detected in 13 of 15 tumors, and the expression of thyroglobulin was detected only in tumors positive for Gal-NAc-T3 expression (Table 6; Fig. 3C, D). Furthermore, in seven cases that underwent surgical resection, a transition from papillary to anaplastic carcinoma was observed in the resected specimens. In these cases, the papillary carcinoma components were positive for GalNAc-T3 and thyroglobulin in all tumors, whereas the expression of neither GalNac-T3 nor thyroglobulin was detected in the anaplastic carcinoma component in the same tumors, except in one case (Fig. 3E, F).

# Discussion

It is well known that the biological behavior and aggressiveness of thyroid carcinomas depend on histological type, but the mechanisms attributed to variation in biological behavior remain to be fully elucidated (33,34). Many studies

### GALNAC-T3 EXPRESSION IN THYROID CANCER

TABLE 3. GALNAC-T3 EXPRESSION AND CLINICOPATHOLOGICAL FEATURES OF PAPILLARY THYROID CARCINOMA

	No.	No. of positive		
	of cases	GalNAc-T3 expression	%	р
All cases	134	101	73.7	
Male	27	21	77.8	0.75
Female	107	80	74.8	
Age				
<45	46	31	67.4	0.16
≥45	88	69	78.4	
Tumor size				
≤1 cm	28	21	75.0	0.17
1–4 cm	88	69	78.4	
>4 cm	16	9	56.3	
Intraglandula	r metasta	asis		
Absent	36	30	83.3	0.33
Present	89	67	75.3	
Lymph node	metastas	is		
Absent	32	23	71.9	0.61
Present	97	74	76.3	
Lymphatic inf	filtration			
Absent	15	10	66.7	0.76
Present	101	75	75.2	
Vascular infilt	ration			
Absent	22	17	77.3	0.88
Present	100	73	73.0	
Distant metas	tasis			
Absent	129	95	73.6	0.42
Present	5	5	100	
Extraglandula	r invasic	n		
Absent	45	29	64.4	0.026
Present	88	72	81.8	
Histological co	omponer	nts		
Well	215	152	70.7	< 0.001
Papillary	102	74	72.5	
Follicular	113	78	69.2	
Poorly	89	13	14.6	
Trabecular	49	9	18.4	
Solid	40	4	10.0	

have demonstrated that GalNAc-T3 expression is associated with the differentiation or biological behavior of adenocarcinomas. In the present study, we demonstrated for the first time that GalNAc-T3 expression is associated with both the histological type of thyroid carcinomas and the structural

Table 4. GalNAc-T3 Expression in Follicular Thyroid Carcinoma

Component	No. of components	No. of positive GalNAc-T3 expression	%	р
Well-differentiated	10	4	40.0	0.63
Poorly differentiated component	10	2	20.0	

difference in differentiated carcinomas: the expression of GalNAc-T3 was higher in the well-differentiated components and lower in the poorly differentiated components. Furthermore, the expression of GalNAc-T3 was rarely detected in anaplastic carcinomas.

In the present study, the expression of GalNAc-T3 was detected in the majority of follicular epithelial cells in normal thyroid. Previous studies demonstrated strong positivity in normal colorectal epithelium (27), whereas weak expression of GalNAc-T3 was reported in normal breast glandular tissue and normal bile duct epithelium (29,32). Thus, the expression of GalNAc-T3 in normal epithelial cells depends on the organ involved.

With regard to the association of GalNAc-T3 expression and thyroid cancer phenotype, the expression of GalNAc-T3 was detected more frequently in the well-differentiated components compared with the poorly differentiated components, and interestingly, GalNAc-T3 was rarely detected in anaplastic thyroid carcinomas. Consistent with the association of GalNAc-T3 expression and differentiation of thyroid carcinomas, GalNAc-T3 was not detected in anaplastic lesions from specimens showing a transition from papillary to anaplastic carcinoma, except in one case. Thus, the expression of GalNAc-T3 in thyroid carcinomas was decreased in parallel with tumor transition to a more aggressive phenotype.

Although the expression of GalNAc-T3 was inversely correlated with tumor aggressiveness in thyroid carcinomas in the present study, previous studies have demonstrated that positive GalNAc-T3 expression is correlated with tumor aggressiveness in gastric and esophageal carcinomas (25,31). With regard to the expression of GalNAc-T3 in other adenocarcinomas, colorectal carcinoma has been reported to demonstrate a lower intensity than normal colorectal cells (27). In contrast, weak expression of GalNAc-T3 was correlated with an aggressive phenotype in pancreatic and gallbladder carcinomas (28,30). Moreover, in gallbladder and extrahepatic bile duct carcinomas, high-intensity GalNAc-T3 expression was detected in noninvasive or minimally invasive carcinomas, and the proportion of low-intensity GalNAc-T3 expression was increased at the invasive border of advanced carcinomas (29). Thus, our data, together with those of other studies, suggest that the expression pattern of GalNAc-T3 in endocrine and exocrine organs is different from that in epithelial cells in other organs. In the present study, the expression of GalNAc-T3 was not detected in medullary carcinomas. Considering that medullary thyroid carcinoma derives from neuroendocrine parafollicular C cells of the thyroid gland, which is different from the origin of papillary and follicular thyroid cancer, the negative expression of GalNAc-T3 in medullary carcinoma may reflect difference in the origin of the cancer cells.

In the present study, a significantly positive association between the expression of GalNAc-T3 and that of thyroglobulin was observed in tumor lesions showing a transition from papillary to anaplastic carcinoma. Human thyroglobulin is a large molecule containing 2750 amino acids, with a molecular weight of 330 kDa (35). Although 20 putative N-linked glycosylation sites have been proposed, only 16 are glycosylated in the mature protein (36). Immunohistochemical detection of thyroglobulin in surgical specimens is useful in the differential diagnosis of tumors of unknown origin, and measurement of serum thyroglobulin is primarily used as a tumor marker in

		Thyroglobulin	GalNac-T3		
Histological type	Component		Positive	Negative	р
Papillary carcinoma	Well differentiated	Positive Negative	148	60 3	0.42
	Poorly differentiated	Positive Negative	8 1	64 16	0.84
Follicular carcinoma	Well differentiated	Positive Negative	$4 \\ 0$	6 0	NA
	Poorly differentiated	Positive Negative	2 0	7 1	0.43

TABLE 5. GALNAC-T3 AND THYROGLOBULIN EXPRESSION IN DIFFERENTIATED THYROID CARCINOMAS

NA, not available.

the postoperative management of patients with differentiated thyroid cancer. Thus, thyroglobulin measurements in tissue and serum play an integral role in the evaluation of patients with thyroid cancer (37). It is known that during malignant transformation, epithelial cells modulate the glycosylation profile of their secretion products and that these posttranslational modifications may represent exploitable diagnostic or prognostic markers, which may provide important tools for elucidating the molecular mechanisms responsible for the progression of these tumors (38). In thyroid malignancies, heterogeneity in the carbohydrate chains has been reported (30,39); moreover, promising results of studies using lentil (*Lens culinaris*) agglutinin–reactive serum thyroglobulin ratios to distinguish differentiated thyroid cancer from benign

FIG. 3. Expression of GalNAc-T3 and thyroglobulin in papillary and anaplastic thyroid carcinomas. Representative findings of immunohistochemical analyses of GalNAc-T3 and thyroglobulin expression in serial sections of the tumor. Both GalNAc-T3 (A) and thyroglobulin (B) are expressed in the well-differentiated component of papillary carcinoma. Neither GalNAc-T3 (C) nor thyroglobulin (D) was expressed in the anaplastic carcinoma. In tumor sample demonstrating transition from the papillary carcinoma to the anaplastic carcinoma, both GalNAc-T3 (E) and thyroglobulin (F) were expressed in the papillary carcinoma area but their expression decreased in the anaplastic carcinoma area. Color images available online at www .liebertpub.com/thy



#### GALNAC-T3 EXPRESSION IN THYROID CANCER

TABLE 6. GALNAC-T3 AND THYROGLOBULIN EXPRESSION	N
in Anaplastic Thyroid Carcinoma	

	Anaplastic carcinoma component		Papillary carcinoma component	
Case no.	GalNAc-T3	Thyroglobulin	GalNAc-T3	Thyroglobulin
1	_	_	+	+
2	_	_	+	+
3	_	_	+	+
4	_	_	+	+
5	_	_	+	+
6	_	_	+	+
7	+	+	+	+
8	+	+	1	NA
9	_	_	NA	
10	_	_	NA	
11	_	_	NA	
12	_	_	NA	
13	_	_	NA	
14	_	_	NA	
15	_	-	NA	

NA, not available.

thyroid lesions have been reported (40,41). Thus, differences in post-translational modifications of the glycosylation sites of thyroglobulin occurring in differentiated thyroid cancer may have potential as a tumor-specific marker, but the alterations in the glycosylation of thyroglobulin occurring in the process of malignant transformation have not been precisely elucidated. The expression of thyroglobulin is known to be detectable in less than 10% of anaplastic thyroid cancers, whereas thyroglobulin is considered to be a marker in differentiated thyroid carcinomas (42-44), indicating that thyroid cancer cells lose the ability to produce thyroglobulin in the process of dedifferentiation to a more aggressive phenotype. As no direct association of GalNAc-T3 and thyroglobulin has been demonstrated in the thyroid gland, our data suggest that absent expression of GalNAc-T3 is attributed to the post-translational modification of glycosylation induced along with the anaplastic transformation of differentiated thyroid carcinoma.

In conclusion, further studies are required to elucidate the function of GalNAc-T3 in thyroid carcinomas, and GalNAc-T3 is not useful in distinguishing a malignant lesion from a benign lesion. However, the present study suggests that lower expression of GalNAc-T3 could be a useful marker for identifying differentiated thyroid carcinomas with a more aggressive phenotype.

#### Author Disclosure Statement

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

# References

- 1. Sherman SI 2003 Thyroid carcinoma. Lancet 361:501-511.
- Voutilainen PE, Multanen M, Haapiainen RK, Leppaniemi AK, Sivula AH 1999 Anaplastic thyroid carcinoma survival. World J Surg 23:975–978; discussion 978–979.

- Ito K, Hanamura T, Murayama K, Okada T, Watanabe T, Harada M, Ito T, Koyama H, Kanai T, Maeno K, Mochizuki Y, Amano J 2012 Multimodality therapeutic outcomes in anaplastic thyroid carcinoma: improved survival in subgroups of patients with localized primary tumors. Head Neck 34:230–237.
- Hakomori S 1989 Aberrant glycosylation in tumors and tumor-associated carbohydrate antigens. Adv Cancer Res 52: 257–331.
- Nakamori S, Ota DM, Cleary KR, Shirotani K, Irimura T 1994 MUC1 mucin expression as a marker of progression and metastasis of human colorectal carcinoma. Gastroenterology 106:353–361.
- Hattrup CL, Gendler SJ 2008 Structure and function of the cell surface (tethered) mucins. Annu Rev Physiol 70:431–457.
- Dube DH, Bertozzi CR 2005 Glycans in cancer and inflammation—potential for therapeutics and diagnostics. Nat Rev Drug Discov 4:477–488.
- Hollingsworth MA, Swanson BJ 2004 Mucins in cancer: protection and control of the cell surface. Nat Rev Cancer 4:45–60.
- Ogawa J, Sano A, Koide S, Shohtsu A 1994 Relation between recurrence and expression of proliferating cell nuclear antigen, sialyl LewisX, and sialyl Lewis(a) in lung cancer. J Thorac Cardiovasc Surg 108:329–336.
- Rice GE, Bevilacqua MP 1989 An inducible endothelial cell surface glycoprotein mediates melanoma adhesion. Science 246:1303–1306.
- Itzkowitz SH, Bloom EJ, Kokal WA, Modin G, Hakomori S, Kim YS 1990 Sialosyl-Tn. A novel mucin antigen associated with prognosis in colorectal cancer patients. Cancer 66:1960– 1966.
- 12. Nakamori S, Kameyama M, Imaoka S, Furukawa H, Ishikawa O, Sasaki Y, Kabuto T, Iwanaga T, Matsushita Y, Irimura T 1993 Increased expression of sialyl Lewisx antigen correlates with poor survival in patients with colorectal carcinoma: clinicopathological and immunohistochemical study. Cancer Res 53:3632–3637.
- Niklinski J, Furman M, Laudanski J, Kozlowski M 1992 Prognostic value of pretreatment CEA, SCC-Ag and CA 19– 9 levels in sera of patients with non-small cell lung cancer. Eur J Cancer Prev 1:401–406.
- 14. Diez M, Torres A, Maestro ML, Ortega MD, Gomez A, Pollan M, Lopez JA, Picardo A, Hernando F, Balibrea JL 1996 Prediction of survival and recurrence by serum and cytosolic levels of CEA, CA125 and SCC antigens in resectable nonsmall-cell lung cancer. Br J Cancer 73:1248–1254.
- 15. Ohgami A, Tsuda T, Osaki T, Mitsudomi T, Morimoto Y, Higashi T, Yasumoto K 1999 MUC1 mucin mRNA expression in stage I lung adenocarcinoma and its association with early recurrence. Ann Thorac Surg **67**:810–814.
- Bresalier RS, Niv Y, Byrd JC, Duh QY, Toribara NW, Rockwell RW, Dahiya R, Kim YS 1991 Mucin production by human colonic carcinoma cells correlates with their metastatic potential in animal models of colon cancer metastasis. J Clin Invest 87:1037–1045.
- 17. Bresalier RS 1994 Adhesion molecules and gastrointestinal malignancies. Gastroenterology **106**:1378–1382.
- Sutherlin ME, Nishimori I, Caffrey T, Bennett EP, Hassan H, Mandel U, Mack D, Iwamura T, Clausen H, Hollingsworth MA 1997 Expression of three UDP-N-acetyl-alpha-Dgalactosamine:polypeptide GalNAc N-acetylgalactosaminyltransferases in adenocarcinoma cell lines. Cancer Res 57: 4744–4748.

- Hagen FK, Van Wuyckhuyse B, Tabak LA 1993 Purification, cloning, and expression of a bovine UDP-GalNAc: polypeptide N-acetyl-galactosaminyltransferase. J Biol Chem 268:18960–18965.
- Tarp MA, Clausen H 2008 Mucin-type O-glycosylation and its potential use in drug and vaccine development. Biochim Biophys Acta 1780:546–563.
- Yoshimura Y, Nudelman AS, Levery SB, Wandall HH, Bennett EP, Hindsgaul O, Clausen H, Nishimura S 2012 Elucidation of the sugar recognition ability of the lectin domain of UDP-GalNAc:polypeptide N-acetylgalactosaminyltransferase 3 by using unnatural glycopeptide substrates. Glycobiology 22:429–438.
- 22. Homa FL, Hollander T, Lehman DJ, Thomsen DR, Elhammer AP 1993 Isolation and expression of a cDNA clone encoding a bovine UDP-GalNAc:polypeptide N-acetylgalactosaminyltransferase. J Biol Chem **268**:12609–12616.
- 23. White T, Bennett EP, Takio K, Sorensen T, Bonding N, Clausen H 1995 Purification and cDNA cloning of a human UDP-N-acetyl-alpha-D-galactosamine:polypeptide N-acetyl galactosaminyltransferase. J Biol Chem **270**:24156–24165.
- 24. Bennett EP, Hassan H, Clausen H 1996 cDNA cloning and expression of a novel human UDP-N-acetyl-alpha-D-galactosamine. Polypeptide N-acetylgalactosaminyltransferase, GalNAc-t3. J Biol Chem **271**:17006–17012.
- 25. Onitsuka K, Shibao K, Nakayama Y, Minagawa N, Hirata K, Izumi H, Matsuo K, Nagata N, Kitazato K, Kohno K, Itoh H 2003 Prognostic significance of UDP-N-acetyl-alpha-Dgalactosamine:polypeptide N-acetylgalactosaminyltransferase-3 (GalNAc-T3) expression in patients with gastric carcinoma. Cancer Sci 94:32–36.
- Dosaka-Akita H, Kinoshita I, Yamazaki K, Izumi H, Itoh T, Katoh H, Nishimura M, Matsuo K, Yamada Y, Kohno K 2002 N-acetylgalactosaminyl transferase-3 is a potential new marker for non-small cell lung cancers. Br J Cancer 87: 751–755.
- 27. Shibao K, Izumi H, Nakayama Y, Ohta R, Nagata N, Nomoto M, Matsuo K, Yamada Y, Kitazato K, Itoh H, Kohno K 2002 Expression of UDP-N-acetyl-alpha-D-galactosaminepolypeptide galNAc N-acetylgalactosaminyl transferase-3 in relation to differentiation and prognosis in patients with colorectal carcinoma. Cancer **94**:1939–1946.
- 28. Miyahara N, Shoda J, Kawamoto T, Furukawa M, Ueda T, Todoroki T, Tanaka N, Matsuo K, Yamada Y, Kohno K, Irimura T 2004 Expression of UDP-N-acetyl-alpha-Dgalactosamine-polypeptide N-acetylgalactosaminyltransferase isozyme 3 in the subserosal layer correlates with postsurgical survival of pathological tumor stage 2 carcinoma of the gallbladder. Clin Cancer Res 10:2090–2099.
- 29. Inoue T, Eguchi T, Oda Y, Nishiyama K, Fujii K, Izumi H, Kohno K, Yamaguchi K, Tanaka M, Tsuneyoshi M 2007 Expression of GalNAc-T3 and its relationships with clinicopathological factors in 61 extrahepatic bile duct carcinomas analyzed using stepwise sections—special reference to its association with lymph node metastases. Mod Pathol 20:267–276.
- 30. Yamamoto S, Nakamori S, Tsujie M, Takahashi Y, Nagano H, Dono K, Umeshita K, Sakon M, Tomita Y, Hoshida Y, Aozasa K, Kohno K, Monden M 2004 Expression of uridine diphosphate N-acetyl-alpha-D-galactosamine: polypeptide N-acetylgalactosaminyl transferase 3 in adenocarcinoma of the pancreas. Pathobiology **71**:12–18.
- 31. Ishikawa M, Kitayama J, Kohno K, Nagawa H 2005 The expression pattern of UDP-N-acetyl-alpha-D-galactosamine-

polypeptide N-acetyl-galactosaminyl transferase-3 in squamous cell carcinoma of the esophagus. Pathobiology **72**: 139–145.

- 32. Nomoto M, Izumi H, Ise T, Kato K, Takano H, Nagatani G, Shibao K, Ohta R, Imamura T, Kuwano M, Matsuo K, Yamada Y, Itoh H, Kohno K 1999 Structural basis for the regulation of UDP-N-acetyl-alpha-D-galactosamine: polypeptide N-acetylgalactosaminyl transferase-3 gene expression in adenocarcinoma cells. Cancer Res 59:6214–6222.
- 33. Ramirez AT, Gibelli B, Tradati N, Giugliano G, Zurlo V, Grosso E, Chiesa F 2007 Surgical management of thyroid cancer. Expert Rev Anticancer Ther 7:1203–1214.
- Are C, Shaha AR 2006 Anaplastic thyroid carcinoma: biology, pathogenesis, prognostic factors, and treatment approaches. Ann Surg Oncol 13:453–464.
- 35. van de Graaf SA, Pauws E, de Vijlder JJ, Ris-Stalpers CR 1997 The revised 8307 base pair coding sequence of human thyroglobulin transiently expressed in eukaryotic cells. Eur J Endocrinol **136:**508–515.
- 36. Yang SX, Pollock HG, Rawitch AB 1996 Glycosylation in human thyroglobulin: location of the N-linked oligosaccharide units and comparison with bovine thyroglobulin. Arch Biochem Biophys 327:61–70.
- 37. Whitley RJ, Ain KB 2004 Thyroglobulin: a specific serum marker for the management of thyroid carcinoma. Clin Lab Med **24**:29–47.
- Lin JD 2008 Thyroglobulin and human thyroid cancer. Clin Chim Acta 388:15–21.
- Tarutani O, Ui N 1985 Properties of thyroglobulins from normal thyroid and thyroid tumor on a concanavalin Asepharose column. J Biochem 98:851–857.
- 40. Shimizu K, Nakamura K, Kobatake S, Satomura S, Maruyama M, Kameko F, Tajiri J, Kato R 2007 The clinical utility of Lens culinaris agglutinin-reactive thyroglobulin ratio in serum for distinguishing benign from malignant conditions of the thyroid. Clin Chim Acta **379**:101–104.
- 41. Kanai T, Amakawa M, Kato R, Shimizu K, Nakamura K, Ito K, Hama Y, Fujimori M, Amano J 2009 Evaluation of a new method for the diagnosis of alterations of Lens culinaris agglutinin binding of thyroglobulin molecules in thyroid carcinoma. Clin Chem Lab Med **47**:1285–1290.
- Rosai J 2003 Immunohistochemical markers of thyroid tumors: significance and diagnostic applications. Tumori 89: 517–519.
- Ordonez NG, El-Naggar AK, Hickey RC, Samaan NA 1991 Anaplastic thyroid carcinoma. Immunocytochemical study of 32 cases. Am J Clin Pathol 96:15–24.
- 44. Wiseman SM, Masoudi H, Niblock P, Turbin D, Rajput A, Hay J, Bugis S, Filipenko D, Huntsman D, Gilks B 2007 Anaplastic thyroid carcinoma: expression profile of targets for therapy offers new insights for disease treatment. Ann Surg Oncol 14:719–729.

Address correspondence to: Ken-ichi Ito, MD, PhD Division of Breast and Endocrine Surgery Department of Surgery Shinshu University School of Medicine 3-1-1 Asahi, Matsumoto Nagano 390-8621 Japan

*E-mail:* kenito@shinshu-u.ac.jp