

**Expression of RECK in endothelial cells of glioma: comparison with CD34 and VEGF expressions**

Nunung Nur Rahmah,<sup>1</sup> Keiichi Sakai,<sup>1,2</sup> Kenji Sano,<sup>3</sup> Kazuhiro Hongo<sup>1</sup>

<sup>1</sup>Department of Neurosurgery, Shinshu University School of Medicine, Matsumoto, Japan

<sup>2</sup>Department of Neurosurgery, Shinshu Ueda Medical Center, Ueda, Japan

<sup>3</sup>Laboratory Medicine, Shinshu University Hospital, Matsumoto, Japan

Keywords: glioma, microvascular density, RECK, CD34, VEGF

Running Title: RECK in glioma angiogenesis

**Corresponding Author:**

Keiichi Sakai, M.D.

Department of Neurosurgery, Shinshu University School of Medicine

Asahi 3-1-1, Matsumoto 390-8621, Japan

Tel. +81-263-37-2690; Fax. +81-263-37-0480

e-mail: skeiichi@shinshu-u.ac.jp

## **Abstract**

**Purpose:** Angiogenesis is thought to involve in progression of glioma grades, and RECK has been said to involve in maturation of vessels. In this study, we aimed to determine whether high micro-vessel density (MVD) expressed by RECK in glioma tissue is correlated with grades of glioma. We also compared RECK expression with that of the formerly known vessel marker, CD34 and VEGF.

**Methods:** RECK, CD34 and VEGF immuno-reactivities of 72 glioma tissues were studied.

**Results:** RECK was seen in microvessels of glioma tissues. CD34 showed similar pattern to RECK, whereas VEGF showed positive staining in cytoplasm of tumor cells and endothelial cells. Average MVD with RECK was 107.6 microvessels (range: 7-290). RECK was positively correlated with grades of glioma. RECK and CD34 also showed strong correlation ( $P= 0.001$ ). Higher frequency of VEGF staining was also correlated with higher grade of glioma.

**Conclusions:** This is the first study describing expression of RECK in glioma, and its angiogenesis-related nature may provide a potential therapeutic target for glioma treatment in the future.

**Keywords:** glioma, microvascular density, RECK, CD34, VEGF

## **Introduction**

Reversion-inducing cysteine-rich protein with Kazal motifs (RECK) acts as a membrane-anchored metalloproteinase regulator[1-4] which functions as a regulator of extracellular matrix integrity in normal condition, contributing to tumor and metastasis suppressing agent. However, recent study found abundant expression of RECK in the cells associated with blood vessels undergoing rapid remodeling in the mouse implantation chambers, suggesting that RECK has a role in vascular remodeling which may involve non-sprouting mechanisms such as intussusception and pruning.[5] Positive RECK staining was also found in endothelial cells of osteosarcoma, indicating their role in angiogenesis of tumor.[6] However, controversies still remain on the function of RECK.

In glioma, angiogenesis has been told to be the cause of increased grades and malignancy. Angiogenesis markers such as platelet-derived growth factor (PDGF), basic fibroblast growth factor (bFGF), epidermal growth factor (EGF), transforming growth factor beta (TGF-B) and vascular endothelial growth factor (VEGF) have been mentioned to play a role in this process. [7-13] However, there has not been any published report explaining the angiogenesis profile of RECK in glioma. Therefore in this study, we aimed at exploring RECK role in angiogenesis of glioma by performing immunohistochemistry study and comparing it with other well-known angiogenesis markers such as CD34 and VEGF.

## **Materials and methods**

### **Patients**

This study was approved by the local Ethical Committee of Shinshu University School of Medicine under registration number 1321. Seventy two patients diagnosed with glioma from Shinshu University Hospital were included in this study. Glioma was graded based on WHO classification,[14] details are explained in Table 1. Information on glioma grades was extracted from the medical records.

### **Immunohistochemical study**

For single labeling, glioma tissues in formalin-fixed and paraffin-embedded tissue samples were stained with RECK (R&D Systems, Inc), CD34 (Beckman Coulter, Inc, Fullerton, Calif) and VEGF (Santa Cruz Biotechnology, Inc., Santa Cruz, CA). Paraffin-embedded tissue samples were washed free of fixative with phosphate-buffered saline (PBS, pH 7.4), exposed to 0.3% H<sub>2</sub>O<sub>2</sub> in PBS for 5 minutes to inactivate endogenous peroxidase, then pretreated with trypsin solution 37°C for 30 minutes for RECK and CD34, and EDTA (microwaved) for 30 minutes for VEGF. The sections were then reacted with primary antibody overnight in a humidity chamber at 4°C. Dilution of primary antibodies was 1:400 for anti-RECK IgG, 1:100 for anti-CD34, and 1:50 for anti-VEGF IgG. Polyclonal goat IgG antibody, as the secondary antibody for RECK, polyclonal mouse IgG antibody for anti-CD34 and polyclonal mouse IgG antibody for VEGF, were applied to the sections for 30 minutes. The color was developed with 0.02% 3,3'-diaminobenzidine tetrahydrochloride and 0.006% H<sub>2</sub>O<sub>2</sub> in PBS. The slides were counterstained with Mayer's hematoxylin. Between steps, the slides were washed three times in PBS. RECK expression on human pancreatic tissue served as a positive control.

To determine more specifically co-expression of RECK and CD34 in endothelial cells of glioma, paraffin-embedded tissue samples and frozen-section tissue samples were double stained. Paraffin-embedded double staining was performed using EnVision™ G|2 Doublestain System (Dako, Kyoto, Japan) with 2 different Chromogens: DAB<sup>+</sup> Chromogen (brown color) for RECK and Permanent Red Chromogen (red color) for CD34. Section incubated without the primary antibody served as negative control. Frozen-section double staining was performed using Rhodamine-labeled anti-goat IgG (Jackson ImmunoResearch, West Grove, PA) for RECK and Fluorescein isothiocyanate (FITC-labeled) anti-mouse IgG (Jackson ImmunoResearch) for CD34.

Frequency of the tumor cell stained was used to calculate VEGF expressions (percentage of stained tumor cell per total number of visible cells) in 400x high power field. RECK and CD34 staining was counted as microvascular density (MVD), which was obtained by manually counting the positive foci for slides counterstained with hematoxylin.

MVD was scored as the number of vessels found in the field, and the final score was the average of 3 most-vascularized areas on high power field (200x).

### **Statistical Analysis**

SPSS 18 for Mac (SPSS, Chicago, IL, USA) was used for the statistical analysis. Significance of correlations among RECK, CD34 and VEGF was assessed using Pearson's correlation. Mean difference of expression based on histological grade of glioma was assessed using unpaired student t-test. Probability values of less than 0.05 were considered significant.

### **Results**

#### **Characteristics of the expression in glioma**

In total 72 gliomas were included in the study. They consisted of 26 samples of grade II, 25 of grade III, and 21 of grade IV. Observers were blinded to the grades during counting. Interestingly, RECK disclosed strong staining of the endothelial cells, appearing dark brown with DAB coloring. However, there was no staining of nuclear, cytoplasm or membrane of tumor cells. RECK showed similar pattern to CD34 staining (Figure 1), whereas VEGF showed mild to moderate staining of nuclear and cytoplasm of tumor cells, and endothelial cells (Figure 2).

#### **Correlation of RECK, CD 34 and VEGF with histological grade**

MVD count using RECK gave an average score of 107.6 (range: 7-290), and CD34 was 111.6 (range: 34.7-413). CD34 is a well-known marker for endothelial cells, and in this study RECK showed similar pattern to CD34 staining, showing that RECK did stain endothelial cells of glioma tissues. RECK and CD34 expression in grade IV showed significant mean difference between grade II and grade III ( $P= 0.01, 0.03, 0.002, 0.013$ , respectively; Figure 3). Positive correlation was also observed between RECK and CD34 (Pearson's coefficient correlation= 0.56,  $P= 0.001$ ) (Table 2). In order to confirm the co-expression of RECK and CD34, double staining was performed in this study.

Paraffin-embedded double staining and frozen-section immunofluorescent double staining disclosed the similar location of expression between RECK and CD34 (Figure 4).

VEGF frequency of staining had an average score of 50.8% (range: 0-90), and higher VEGF staining correlated with higher grade of glioma ( $P= 0.001$ ,  $0.001$ , respectively; Figure 3). VEGF showed various staining patterns, such as aggregated and diffuse staining of the cytoplasm. RECK and VEGF showed different pattern of staining in glioma, and statistical analysis disclosed no correlation between the two ( $P= 0.369$ , Table 2).

## **Discussion**

Angiogenesis and the production of angiogenic factors are fundamental for tumor progression in the form of growth, invasion and metastasis.[15] There are two known types of angiogenesis: 1) the so called sprouting angiogenesis, which is characterized by the proliferation and migration of endothelial cells into avascular sites; and 2) the `nonsprouting` angiogenesis or intussusceptive microvascular growth.[16] In glioma, as with malignant tumors elsewhere in the body, angiogenesis is necessary for the growth and progression.[17] Several growth factors are said to involve in glioma angiogenesis, notably platelet-derived growth factor (PDGF),[7] basic fibroblast growth factor (bFGF),[8] epidermal growth factor (EGF),[9] transforming growth factor beta (TGF-B),[18] and vascular endothelial growth factor (VEGF).[10-13]

Among angiogenic agents, VEGF is the most studied one, and its role in driving the angiogenic process in glioma is well known.[10-13] As mentioned by the previous authors, our study also proved the correlation of VEGF expression and glioma grades. In glioblastoma group, the expression of VEGF was at least 70%. Chaudhry et al. in their study of 41 glioma cases disclosed at least 86% immunoreactivity in glioblastoma group,[17] showing the important role of VEGF in progression of glioma. Nico et al. even explained the involvement of VEGF in intussusceptive microvascular growth in glioblastoma.[19] CD34 is well known as endothelial marker, and it gives positive staining in physiologic and pathologic vessels.[20-22] CD34 is also considered an optimum marker for microvascular density studies because of its good immunoreactivity.[22] Netto et al., in

their oligodendroglioma study, found a strong expression of CD34 in the endothelial cells without any significant correlation to the grades of oligodendroglioma.[20] However, in this study, we found a strong expression of CD34 and high grade gliomas showed higher expression of CD34, which means that high grade gliomas have higher density of vessels.

On the other hand, RECK has been widely published as an MMP inhibitor. In normal condition RECK is understood to have a function of regulating extracellular matrix integrity, contributing to inhibition of tumor and metastasis progression. Lack of RECK expression has been noticed in several cancers, and it is correlated with malignancy and poorer prognosis.[23] Our previous study in skull base chordoma also showed concordance result where lack of RECK expression correlated with poorer prognosis of the patients.[24] However, recent studies have disclosed RECK involvement in angiogenesis, where RECK showed positive staining in endothelial cells of neuroblastoma[25] and osteosarcoma.[6] RECK was also found in maternal and embryonic vessels in mice showing its importance in maturation process of vessels.[26]

In this study, we found out that RECK was not expressed in tumor cells, and was the same findings through out all grades. It was found in endothelial cells with increasing number in accordance to its grade. RECK expression in endothelial cells was also proved by its co-expression with CD34. There are controversies of RECK function in tumor. RECK has been said to play a role against progression of tumor, however a recent study found that treatment with an angiogenic factor, or co-culture with hypervascular tumor-derived cells markedly induced RECK expression in human umbilical vein endothelial cells (HUVEC), which indicates that RECK may possess proangiogenic function.[25] Our finding is in discordance with the results from previously published articles, however, a recent study by Miki et al. perhaps can explain the reasons. Cell lines with higher RECK-inducing activity tend to express endogenous RECK, implicating that RECK could be induced in such tumors by an autocrine mechanism. However, depletion of endogenously expressed RECK in one of these RECK-positive tumor cell lines did not result in detectable changes in cell behavior, including proliferation, cell death, migration, and invasion.[25] Therefore, in glioma, instead of acting as inhibitor, RECK maybe

involved in a similar manner, and supplying vessels to tumor cells, thus facilitating tumor growth. Further study is needed to answer this issue. Another interesting point is that in our study, we did not find any significant relationship between RECK and VEGF, even though both play roles in angiogenesis. Perhaps they employ a different path in influencing angiogenesis of tumor, as explained by Song et al. in their study of gastric cancer.[27]. Mechanistic study in order to prove the true angiogenesis function of RECK will be the next focus of our study.

### **Conclusion**

Our immunohistochemistry study demonstrated that RECK was identified in endothelial cells of glioma and tended to increase toward higher grades. This preliminary study also identified expression of VEGF in tumor cells, and that higher expression of VEGF also correlated with grades of glioma. However, RECK and VEGF did not show any significant correlation. Further study is necessary to elucidate its role in angiogenesis of glioma, and its potentials as target therapy of glioma in the future.

### **Funding**

This study was supported in part by the Research Fund for Health Care Promotion Plan in Josho Area, Nagano Prefecture from the Ministry of Health, Labour and Welfare of Japan and the Grants-in-Aid for Foreign Researcher Program of Shinshu Public Utility Foundation for Promotion of Medical Sciences.

### **Conflict of Interest**

All authors have no financial, commercial, legal, or professional relationship with other organizations or with the people working with us that may exert an influence on this research. Therefore, we have disclosed any conflict of interest in the making of this paper.

### **Acknowledgements**



The authors are very grateful to Ms. Tomoko Nishizawa and Dr. Takashi Ehara from department of pathology, Shinshu University School of Medicine, for their help with the immunohistochemistry staining technique.

#### References

1. Takahashi C, Sheng Z, Horan TP, Kitayama H, Maki M, Hitomi K, Kitaura Y, Takai S, Sasahara RM, Horimoto A, Ikawa Y, Ratzkin BJ, Arakawa T, Noda M: Regulation of matrix metalloproteinase-9 and inhibition of tumor invasion by the membrane-anchored glycoprotein RECK. *Proc Natl Acad Sci U S A* 95: 13221-13226, 1998
2. Oh J, Takahashi R, Kondo S, Mizoguchi A, Adachi E, Sasahara RM, Nishimura S, Imamura Y, Kitayama H, Alexander DB, Ide C, Horan TP, Arakawa T, Yoshida H, Nishikawa S, Itoh Y, Seiki M, Itohara S, Takahashi C, Noda M: The membrane-anchored MMP inhibitor RECK is a key regulator of extracellular matrix integrity and angiogenesis. *Cell* 107: 789-800, 2001
3. Miki T, Takegami Y, Okawa K, Muraguchi T, Noda M, Takahashi C: The reversion-inducing cysteine-rich protein with Kazal motifs (RECK) interacts with membrane type 1 matrix metalloproteinase and CD13/aminopeptidase N and modulates their endocytic pathways. *J Biol Chem* 282: 12341-12352, 2007
4. Omura A, Matsuzaki T, Mio K, Ogura T, Yamamoto M, Fujita A, Okawa K, Kitayama H, Takahashi C, Sato C, Noda M: RECK forms cowbell-shaped dimers and inhibits matrix metalloproteinase-catalyzed cleavage of fibronectin. *J Biol Chem* 284: 3461-3469, 2009
5. Noda M, Takahashi C: Recklessness as a hallmark of aggressive cancer. *Cancer Sci* 98: 1659-1665, 2007
6. Clark JC, Akiyama T, Thomas DM, Labrinidis A, Evdokiou A, Galloway SJ, Kim HS, Dass CR, Choong PF: RECK in osteosarcoma: A novel role in tumour vasculature and inhibition of tumorigenesis in an orthotopic model. *Cancer*, 2011
7. Maxwell M, Naber SP, Wolfe HJ, Galanopoulos T, Hedley-Whyte ET, Black PM,

Antoniades HN: Coexpression of platelet-derived growth factor (PDGF) and PDGF-receptor genes by primary human astrocytomas may contribute to their development and maintenance. *J Clin Invest* 86: 131-140, 1990

8. Gately S, Soff GA, Brem S: The potential role of basic fibroblast growth factor in the transformation of cultured primary human fetal astrocytes and the proliferation of human glioma (U-87) cells. *Neurosurgery* 37: 723-730; discussion 730-722, 1995

9. Goldman CK, Kim J, Wong WL, King V, Brock T, Gillespie GY: Epidermal growth factor stimulates vascular endothelial growth factor production by human malignant glioma cells: a model of glioblastoma multiforme pathophysiology. *Mol Biol Cell* 4: 121-133, 1993

10. Plate KH, Breier G, Weich HA, Risau W: Vascular endothelial growth factor is a potential tumour angiogenesis factor in human gliomas in vivo. *Nature* 359: 845-848, 1992

11. Machein MR, Plate KH: VEGF in brain tumors. *J Neurooncol* 50: 109-120, 2000

12. Fischer I, Gagner JP, Law M, Newcomb EW, Zagzag D: Angiogenesis in gliomas: biology and molecular pathophysiology. *Brain Pathol* 15: 297-310, 2005

13. Rubenstein JL, Kim J, Ozawa T, Zhang M, Westphal M, Deen DF, Shuman MA: Anti-VEGF antibody treatment of glioblastoma prolongs survival but results in increased vascular cooption. *Neoplasia* 2: 306-314, 2000

14. Louis DN, Ohgaki H, Wiestler OD, Cavenee WK, Burger PC, Jouvet A, Scheithauer BW, Kleihues P: The 2007 WHO classification of tumours of the central nervous system. *Acta Neuropathol* 114: 97-109, 2007

15. Ribatti D, Vacca A, Dammacco F: The role of the vascular phase in solid tumor growth: a historical review. *Neoplasia* 1: 293-302, 1999

16. Yancopoulos GD, Davis S, Gale NW, Rudge JS, Wiegand SJ, Holash J: Vascular-specific growth factors and blood vessel formation. *Nature* 407: 242-248, 2000

17. Chaudhry IH, O'Donovan DG, Brenchley PE, Reid H, Roberts IS: Vascular endothelial growth factor expression correlates with tumour grade and vascularity in gliomas. *Histopathology* 39: 409-415, 2001

18. Chen TC, Hinton DR, Yong VW, Hofman FM: TGF-B2 and soluble p55 TNFR

modulate VCAM-1 expression in glioma cells and brain derived endothelial cells. *J Neuroimmunol* 73: 155-161, 1997

19. Nico B, Crivellato E, Guidolin D, Annese T, Longo V, Finato N, Vacca A, Ribatti D: Intussusceptive microvascular growth in human glioma. *Clin Exp Med* 10: 93-98, 2010

20. Netto GC, Bleil CB, Hilbig A, Coutinho LM: Immunohistochemical evaluation of the microvascular density through the expression of TGF-beta (CD 105/endoglin) and CD 34 receptors and expression of the vascular endothelial growth factor (VEGF) in oligodendrogliomas. *Neuropathology* 28: 17-23, 2008

21. Jasek E, Furgal-Borzych A, Lis GJ, Litwin JA, Rzepecka-Wozniak E, Trela F: Microvessel density and area in pituitary microadenomas. *Endocr Pathol* 20: 221-226, 2009

22. Vermeulen PB, Gasparini G, Fox SB, Toi M, Martin L, McCulloch P, Pezzella F, Viale G, Weidner N, Harris AL, Dirix LY: Quantification of angiogenesis in solid human tumours: an international consensus on the methodology and criteria of evaluation. *Eur J Cancer* 32A: 2474-2484, 1996

23. Clark JC, Thomas DM, Choong PF, Dass CR: RECK--a newly discovered inhibitor of metastasis with prognostic significance in multiple forms of cancer. *Cancer Metastasis Rev* 26: 675-683, 2007

24. Rahmah NN, Sakai K, Nakayama J, Hongo K: Reversion-inducing cysteine-rich protein with kazal motifs and matrix metalloproteinase-9 are prognostic markers in skull base chordomas. *Neurosurg Rev* 33: 167-173; discussion 173, 2010

25. Miki T, Shamma A, Kitajima S, Takegami Y, Noda M, Nakashima Y, Watanabe K, Takahashi C: The  $\alpha 1$ -integrin-dependent function of RECK in physiologic and tumor angiogenesis. *Mol Cancer Res* 8: 665-676, 2010

26. Chandana EP, Maeda Y, Ueda A, Kiyonari H, Oshima N, Yamamoto M, Kondo S, Oh J, Takahashi R, Yoshida Y, Kawashima S, Alexander DB, Kitayama H, Takahashi C, Tabata Y, Matsuzaki T, Noda M: Involvement of the Reck tumor suppressor protein in maternal and embryonic vascular remodeling in mice. *BMC Dev Biol* 10: 84, 2010

27. Song SY, Son HJ, Nam E, Rhee JC, Park C: Expression of reversion-inducing-cysteine-rich protein with Kazal motifs (RECK) as a prognostic

indicator in gastric cancer. *Eur J Cancer* 42: 101-108, 2006

### **Figure Legends**

Figure 1. Various expressions of RECK and CD 34 in glioma

RECK (A) and CD 34 (B) are expressed in endothelial cells in glioma, showing a highly dense tumor vessels found in high grade glioma (20x).

Figure 2. Expressions of RECK, CD 34 and VEGF based on glioma grades

RECK (A, D, G), CD 34 (B, E, H) and VEGF (C, F, I) are expressed in glioma and were grouped into grades (40x). A,B,C) are showing their expressions in grade II glioma. D,E,F) are showing the expressions in grade III glioma. G,H,I) are showing their expressions in grade IV glioma.

Figure 3. Distribution of MVD and frequency of expressions based on glioma grades

A) is showing MVD distribution of RECK expressions in different grades of glioma. B) is showing MVD distribution of CD 34 expressions in different grades of glioma. C) is showing distribution of VEGF expressions in different grades of glioma.

Figure 4. Double staining expression of glioma

CD34 (A, D), RECK (C, F) and double stained (B, E) expression in a glioblastoma case, showing a positive permanent red expression for CD34 (A) and DAB for RECK (C) in a paraffin-embedded sample (20x). Frozen-section immunofluorescent staining is showing positive FITC staining of the endothelial cells for CD34 (D) and Rhodamine staining of RECK (F) (40x). Colocalization of both staining is proven both in paraffin-embedded and frozen-section samples (B, E).

Table 1. Distribution of glioma cases based on WHO classification

WHO Grading	Type	Frequency
II	Pilomyxoid astrocytoma	9
	Diffuse astrocytoma	10
	Oligoastrocytoma	7
III	Anaplastic astrocytoma	25
IV	Glioblastoma	21
Total		72

Table 2. Correlation among markers (RECK, CD34 and VEGF)

Expression	Pearson Correlation & P Value		
	RECK	CD34	VEGF
RECK		0.56 P= 0.001	0.11 P= 0.39
CD34			0.106 P= 0.416
VEGF			

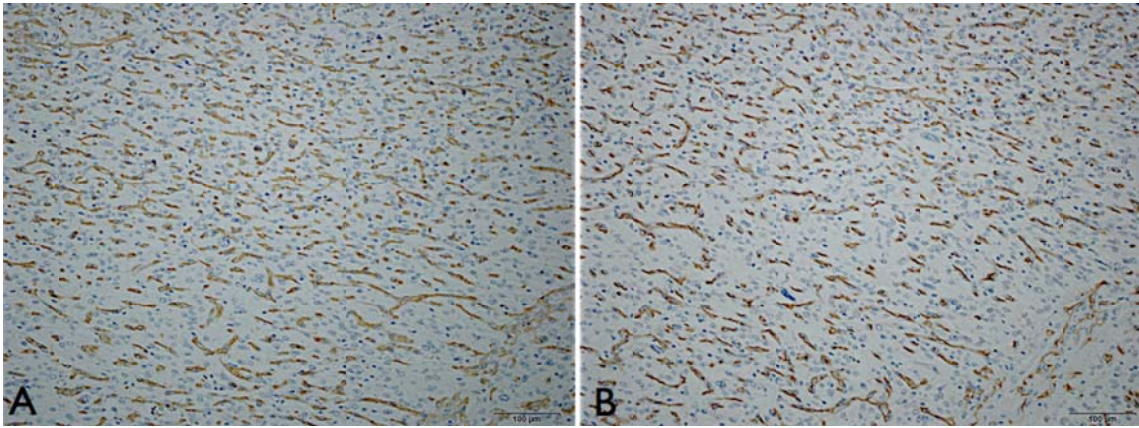


Figure 1.

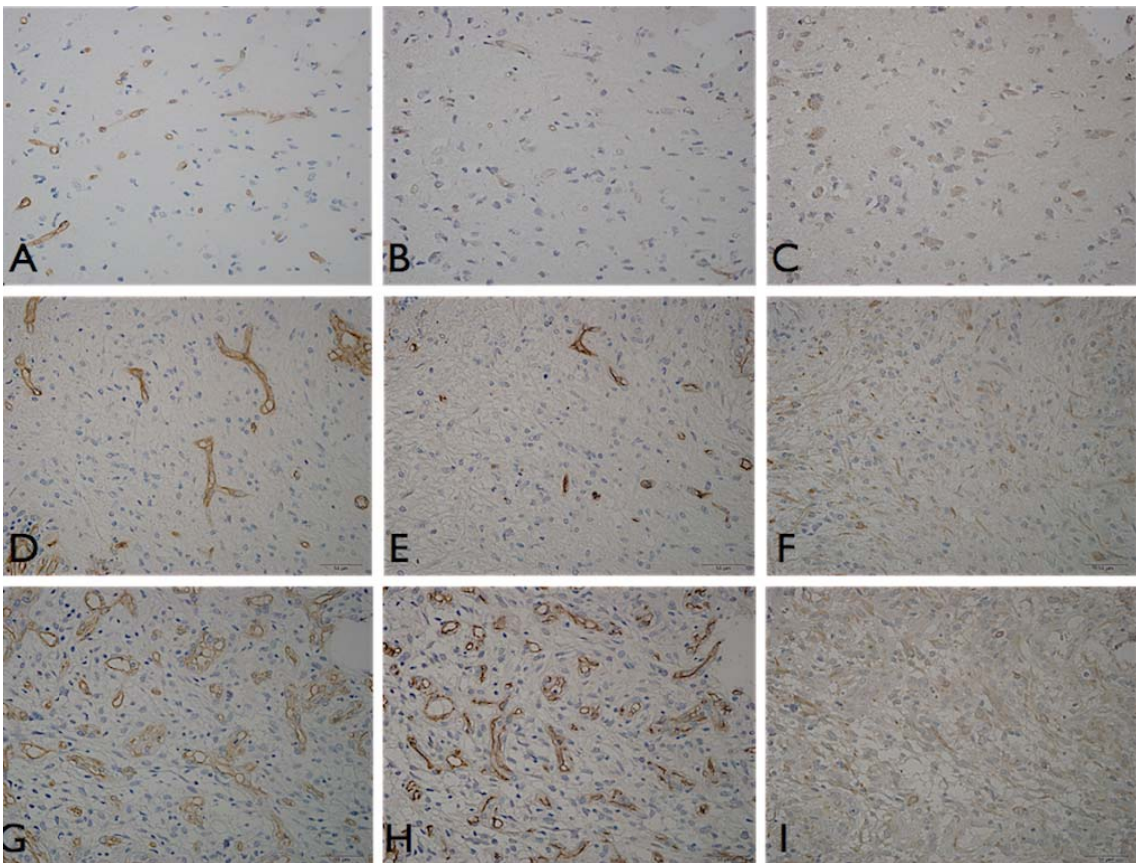


Figure 2.

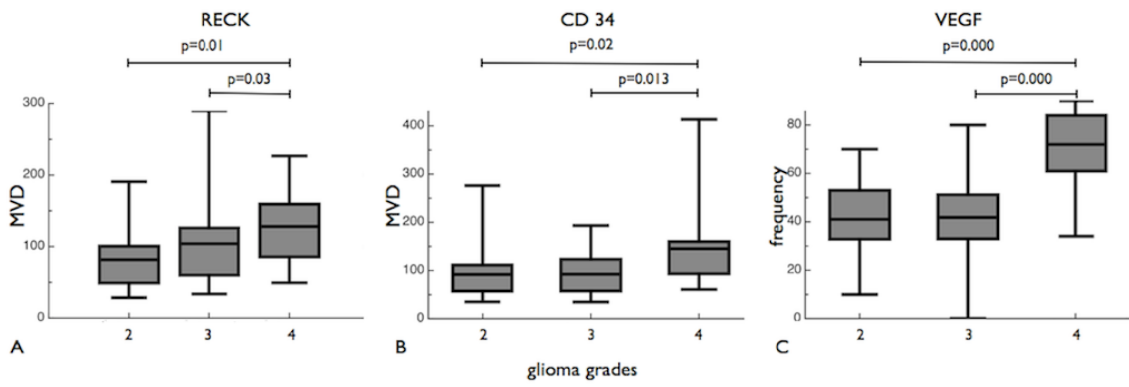


Figure 3.

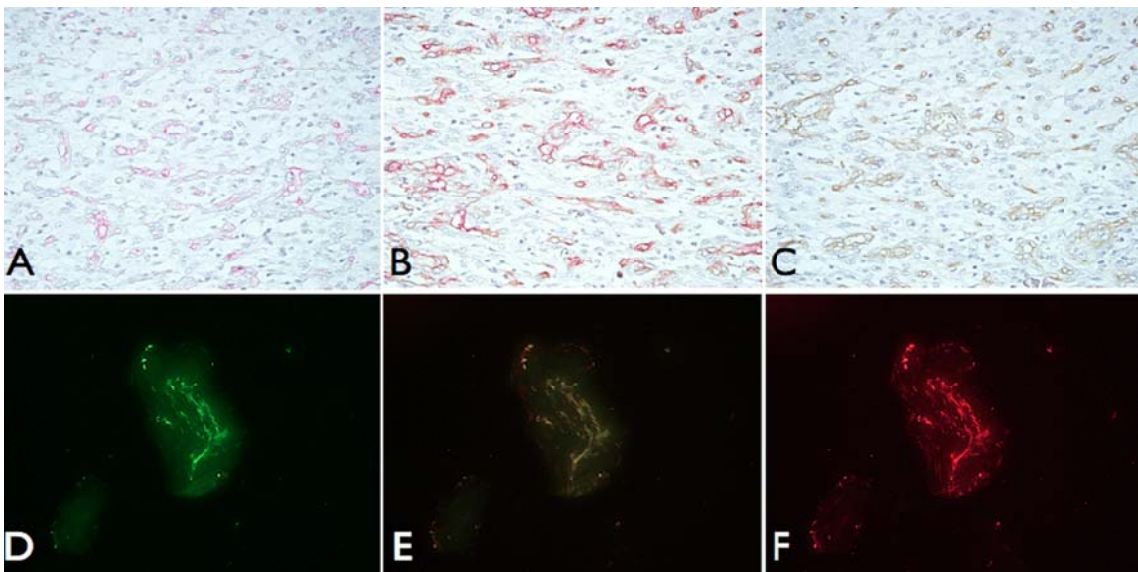


Figure 4.